



Review

Magnetic Fields and Cancer: Epidemiology, Cellular Biology, and Theranostics

Massimo E. Maffei

Department Life Sciences and Systems Biology, University of Turin, Via Quarello 15/a, 10135 Turin, Italy;
massimo.maffei@unito.it; Tel.: +39-011-670-5967

Abstract: Humans are exposed to a complex mix of man-made electric and magnetic fields (MFs) at many different frequencies, at home and at work. Epidemiological studies indicate that there is a positive relationship between residential/domestic and occupational exposure to extremely low frequency electromagnetic fields and some types of cancer, although some other studies indicate no relationship. In this review, after an introduction on the MF definition and a description of natural/anthropogenic sources, the epidemiology of residential/domestic and occupational exposure to MFs and cancer is reviewed, with reference to leukemia, brain, and breast cancer. The *in vivo* and *in vitro* effects of MFs on cancer are reviewed considering both human and animal cells, with particular reference to the involvement of reactive oxygen species (ROS). MF application on cancer diagnostic and therapy (theranostic) are also reviewed by describing the use of different magnetic resonance imaging (MRI) applications for the detection of several cancers. Finally, the use of magnetic nanoparticles is described in terms of treatment of cancer by nanomedical applications for the precise delivery of anticancer drugs, nanosurgery by magnetomechanic methods, and selective killing of cancer cells by magnetic hyperthermia. The supplementary tables provide quantitative data and methodologies in epidemiological and cell biology studies. Although scientists do not generally agree that there is a cause-effect relationship between exposure to MF and cancer, MFs might not be the direct cause of cancer but may contribute to produce ROS and generate oxidative stress, which could trigger or enhance the expression of oncogenes.



Citation: Maffei, M.E. Magnetic Fields and Cancer: Epidemiology, Cellular Biology, and Theranostics. *Int. J. Mol. Sci.* **2022**, *23*, 1339.
<https://doi.org/10.3390/ijms23031339>

Academic Editor: Maurizio Battino

Received: 30 December 2021

Accepted: 22 January 2022

Published: 25 January 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Public concern about electromagnetic fields (EMFs) from power systems is increasing along with the electricity demand, wireless technologies, and changes in work systems and social behavior [1–4]. For modern populations, extremely low-frequency (ELF) electric and magnetic fields (MFs) are common exposures and complex biological mechanisms underly the potential effects of externally-applied MFs [5,6]. In 2002, the International Agency for Research on Cancer (IARC) categorized ELF (including the power frequencies of 50 and 60 Hz) MFs as “possibly carcinogenic to humans” [7].

Controversial and often contradictory scientific reports continue to stimulate debates on the biological effects of EMFs, often leading to confusion and distraction which hamper the development of univocal conclusions on the real hazards that are caused by EMFs [8]. In this review the association between MF and cancer will be reviewed by considering the effect of MF in causing cancer as well as the application of MF as a therapeutic and diagnostic (theranostic) tool. Epidemiological studies, including both domestic/residential and occupational data, as well as human and animal cell studies that were published in the last 20 years will be also considered to provide an overview of the state of the art literature.

The strategy that was implemented to carry out this review was based on a deep search in the databases Web of Science (2000–2021), PubMed (2000–2021), and the EMF

Portal (<https://www.EMF-portal.org/en>, accessed on 1 December 2021). By considering as entries the terms “cancer” AND “magnetic field” the total number of Web of Science Core Collection papers in the period from January 2000 to December 2021 was 12,364, whereas, for the same period, the total number of papers in PubMed was 11,539. The selection of papers was done on the terms: diagnostics, therapy, epidemiology, policy, along with a selection of cancer types, and the exclusion criteria was the impossibility to obtain a full text or the lack of specificity with the selected areas of the review.

Despite the narrative nature of this view, quantitative data on MF exposure and methodologies are described in five supplementary Tables (Supplementary Tables S1–S5), whereas a supplementary data set (Supplementary Data Set S1, EndNote file) contains all references that were cited in this article in addition to many other references.

1.1. Definition and Natural/Anthropogenic Sources of Magnetic Fields

EMFs are present everywhere in our environment. Electric fields are produced by the local build-up of electric charges in the atmosphere that are associated with thunderstorms. The Earth’s MF, or geomagnetic field (GMF), is the principal source of static fields (SFs) [9]. It interacts with the geosphere and the biosphere and plays a major role in shielding the harmful effects of cosmic radiation. Different areas inside our planet are responsible for the GMF which can be represented as the sum of MFs of several sources:

$$FT = F_0 + F_m + F_a + F_e + \delta F$$

where F_0 is the dipolar component of the GMF, F_m is the field of world anomalies that are associated with the heterogeneity of the planet interior (non-dipolar field), F_a is the magnetization of rocks in the Earth’s crust (anomalous field), F_e is the external sources field, and δF is the field variation that is also associated to external causes. The main GMF is also represented by the sum of the dipolar and non-dipolar fields ($F = F_0 + F_m$).

The GMF is composed of three orthogonal vectors: X, Y, and Z. The combination of the two horizontal vectors yields the horizontal component H , which is aligned in the direction of the compass needle and that can be expressed as:

$$H = \sqrt{X^2 + Y^2}$$

Whereas the total field intensity, which at the poles is directed towards the center of the planet, can be expressed as:

$$H = \sqrt{X^2 + Y^2 + Z^2}$$

The angle that is formed between H and the geographic north is the declination, D , whereas the inclination, I , is the angle between the horizontal plane and the vector of total field intensity F . In the international SI system the magnetic induction or magnetic flux density (B) is measured in Tesla (T) and its subunits ($\mu T = 10^{-6} T$; $nT = 10^{-9} T$). One tesla equals one Weber per square meter, corresponding to 10^4 gauss (G), which is the unit of magnetic field in the centimeter-gram-second system. Thus, $1 G = 100 \mu T$.

The magnetic flux density, B , is linked to the magnetic field strength, H , by a material constant, the magnetic permeability μ (also called magnetic conductivity).

$$B = \mu \times H$$

The magnetic permeability, μ , is a measure of the permeability of materials for MFs.

The power flux density, S , of the EMF consists of energy fractions of the electric and MF components and is measured in Watts per square meter ($W m^{-2}$). The field strength decreases with increasing distance from the field source.

The strength of the GMF at the surface of the Earth ranges from over $60 \mu T$ around the magnetic poles in northern Canada, the south of Australia, and in parts of Siberia to less

than 30 μT in an area that includes most of South America and South Africa (the so-called South Atlantic anomaly) [10]. In the Earth's history, the GMF has changed with the so-called geomagnetic reversals, where the GMF was characterized by periods (more-or-less extended) with the same polarity. These reversals occurred some hundred times since the Earth's formation, with intervals between the polarity phases estimated around 300,000 years. The present normal polarity started around 780,000 years ago; therefore, an imminent geomagnetic reversal would not be so unexpected. The South Atlantic anomaly, a zone with significant reduction of the GMF intensity that is located in front of Brazil/Argentina, could be the initial symptom of a future change of polarity [2]. Changes in GMF intensity imply a reduction of the GMF shield against cosmic radiation, with possible consequences for all living organisms, which cannot avoid the effects of the GMF [11].

It is not clear whether the GMF can contribute to potential health risks, being present in our planet before the evolution of living organisms; however, one question remains whether different values of GMF in different countries in which epidemiological studies of anthropogenic sources of MF have been performed might affect the results of those studies [12].

Besides natural sources, the EM spectrum also includes fields that are generated by human-made sources. For instance, X-rays are generated and used for diagnosis, power sockets are associated with low frequency electromagnetic fields (LF EMFs), and various kinds of higher frequency radio waves are used to transmit information—whether via TV antennas, radio stations, or mobile phone base stations.

Static magnetic fields (SMF) (with direct current, DC) or alternating magnetic fields (AMF) (with alternate current, AC) are formed, depending on the current feed. The polarity of AMF changes according to the cyclic changes in the direction of the current flow (e.g., 100 polarity changes per second with 50 Hz AC), whereas in SMFs, the polarity is unchanged.

The scientifically documented interaction with the organism allows the classification of non-ionizing electromagnetic fields (NI EMF) into low frequency (LF) and radio frequency (RF). The stimulation or excitation of nerves, muscles, and sensory receptors may occur below a threshold of 1 MHz; however, values that are higher than 1 MHz generate only thermal effects. EMFs and radiation cover a wide frequency range. The NI radiation range of the EM spectrum up to 300 GHz comprises SMF (0 Hz) and LF fields (≤ 300 Hz), the intermediate-frequency range between approximately 300 Hz and 10 MHz, and the RF range from 10 MHz to 300 GHz.

Table 1 summarizes the classification of MFs based on type of radiation, field, frequency, and wavelength along with some examples and general effects. Static, non-ionizing electric, and MFs that occur as a by-product. SF (0 Hz) occurs in batteries, at high voltage direct current transmission lines (HVDC lines), where underground cables are present, with permanent magnets, between objects with different electrical charges, and in the GMF. In general, the kind of the MF and the level of the magnetic flux density correspond to those of the GMF. Inside of the converter stations, SMFs occur, and their strengths depend on the voltage and the amount of flowing current. In medicine, strong SMF are used in magnetic resonance imaging (MRI, see also Section 4.1). During an MRI procedure the patient is exposed a strong SMF normally from 1.5–3 T. Only in research facilities, magnetic fields from 7 T up to 9.4 T are used. A huge diversity of products with magnets that are in close proximity to their surface in pillows, belts, bracelets, blankets, pendants, patches, or insoles exhibit SMF in the range between 0.03 and 0.3 T. However, the levels are reduced to a tenth at a 3–4 mm distance from the surface. Therefore, at a distance of several centimeters, the magnetic flux density lowers down to that of the natural GMF.

Table 1. Types of magnetic fields.

Type of Radiation	Type of Field	Frequency	Wavelength	Use	Examples	Effect
NI	SMF	0 Hz	N.A.		GMF, permanent magnets, transmission lines, HVDC lines, batteries, between objects with different electrical charges, MRI	Action of force
NI	AMF	0.3 Hz 3 Hz 16 2/3 Hz 50 Hz 300 Hz 3 kHz 30 kHz	10^6 km 100,000 km 18,000 km 6000 km 1000 km 100 km 10 km	Low Frequency traction current and three phase alternating current	Technical appliances such as power lines, wiring and household appliances such as appliances for heating (e.g., electric cooker, electric heating, washing machine, electric water heater, iron), appliances with a transformer or magnetic coils (e.g., radio clock, low-voltage halogen lamps, television set, WiFi) and appliances with an electric motor (e.g., vacuum cleaner, drill, hand blender, hair dryer, electric cars)	Stimulation/irritation
NI	AMF	300 kHz 3 MHz 30 MHz 300 MHz 3 GHz 30 GHz 300 GHz 3 THz	1 km 100 m 10 m 1 m 10 mm 10 mm 1 mm 100 μ m	Radio frequency. Ra- dio/television, broadcasting-terrestrial, DVBT) and digital mi- crowaves, terahertz waves	Induction cookers and electronic article surveillance systems in stores, as well as many industrial and medical applications, PC monitors, mobile phone, microwave ovens, radar stations. Broadcasting frequencies (short wave, AM, and FM radio), digital television (digital video radio (digital audio broadcasting, DAB). Wireless local area networks (WiFi, WLAN), cordless telephones, Bluetooth devices, baby monitors, electronic article surveillance systems and RFID (radio frequency range), radar systems, radio relay systems, satellite TV and satellite Internet, radio solutions for stationary Internet	Thermal effect
NI	AMF	30 THz 300 THz	10 μ m 1 μ m	Infrared	Bulb lamps, heaters, body scanners for security control	Thermal effect
NI	AMF	380 THz 789 THz	780 nm 380 nm	Visible Light		
Ionizing	AMF	3×10^{15} Hz 3×10^{16} Hz 3×10^{17} Hz 3×10^{18} Hz 3×10^{19} Hz 3×10^{20} Hz	100 nm 10 nm 1 nm 100 pm 10 pm 1 pm	UV-light, X-rays, gamma rays	Nuclear power plants, X-ray machines, radioactive material.	Ionization

LF refers to the frequency range 0–100 kHz. The energy of the EMF that is absorbed in biological tissue and is converted into heat defines the specific absorption rate (SAR) that is obtained by exposure to a frequency that is between 100 kHz and 10 GHz. The SAR is expressed in Watts per kilogram of tissue ($W \text{ kg}^{-1}$) based on an average exposure time of six minute intervals, during which a balance between the energy input and the heat dissipation in the tissue is reached. It is possible to distinguish between the exposure of the whole body or parts of the body by averaging over different body masses. All electrical applications that are run on power supply (railways, electrical appliances in the home, and at working places) lie in the range of LF AMF up to 1 kHz (wavelengths larger than 300 km) (low frequency (0.1 Hz–1 kHz)). No extremely LF AF occur in nature. ELF AFs are generated by technical appliances such as power lines, wiring, and household appliances. ELF Electric and MFs are generated by the power lines and their strength and distribution

in the area surrounding the power lines depend on several parameters (including voltage, amperage, tower shape, as well as alignment, and number, and slackness of the lines). The strength of electric field is mainly found beneath the power lines; however, this effect rapidly diminishes with increasing distance from the power line [13]. Electric cars are a significant source of very high MFs due to the electric motor and large batteries, especially during starting and stopping. In electric cars ELF MF dominate, but intermediate frequency fields can occur also [14].

In the natural environment EMFs with intermediate frequencies (1 kHz–10 MHz) can be generated during the so-called sferics, which are broadband EM impulses that occur in the Earth's atmosphere as a consequence of lightning discharges. Sferics may extend from a few kHz to several tens of kHz (3–100 kHz) [15]. Intermediate frequency includes the lower range of the radiofrequency band with its corresponding applications, but also applications that are working with specific frequencies, such as induction cookers and electronic article surveillance systems in stores, as well as many industrial and medical applications.

Radio frequency (30 kHz–300 GHz) includes a range of "broadcasting frequencies" (between 30 kHz and 300 MHz; wavelengths from 10 km to 1 m) covering long wave radio broadcasting, amplitude modulation (AM) radio broadcasting, shortwave radio broadcasting, and frequency modulation (FM) radio broadcasting (authorized in the very high frequency range). Terahertz waves are also in the non-ionizing radiation spectral range, between 300 GHz and 10 THz (wavelengths from 1 mm to 30 μm). For example, they are used for quality control of industrial products, at some airports in body scanners for security control, or in skin cancer scanning systems [16].

The following range of 384 THz to 789 THz (780 nm to 380 nm) is referred to as visible light. This is succeeded by the ranges of ultraviolet radiation and the ionizing radiation with even shorter wavelengths.

As noted above, the magnetic field strength around a conductor increases with rising electric current strength and decreases with growing distance from the field source. It is dependent on the type of source how fast the field decreases (Figure 1).

1.2. Public Health Initiatives and Concern

In 1996 the World Health Organization (WHO) launched a large, multidisciplinary research effort to respond to growing public health concerns about the possible health effects from exposure to EMF sources. The International EMF Project, open to any WHO Member State government, brings together current knowledge and available resources of key international and national agencies and scientific institutions. Among the aims and scopes of the EMF Project are: (a) develop and publish a health risk assessment on EM RF fields; (b) develop and disseminate information materials on risk management policies of EMF; (c) provide technical support to national authorities and international organizations regarding NI radiation; (d) establish an inter-agency committee on NI radiation safety to exchange information and harmonize activities; and (e) develop international standards for protection against NI radiation [17].

Worldwide, many countries set their own national standards for exposure based on the guidelines that are set by the International Commission on Non-Ionizing Radiation Protection (ICNIRP), a non-governmental organization that was formally recognized by WHO. Risk assessment analyses that are based on publicly available data are used to help formulate government guidance on occupational MF by also considering the cancer cases that were prevented and the monetary benefits accruing to society by reducing workplace exposures [18]. An overview of the current knowledge regarding EMF-related health risks including recommendations for the diagnosis, treatment, and accessibility measures of electromagnetic hypersensitivity (EHS) to improve and restore individual health outcomes as well as for the development of strategies for prevention has been recently published [19]. The International Radiation Protection Association (IRPA) represents national radiation protection societies [20]. An updated and reliable source of information is provided by the EMF Portal (<https://www.EMF-portal.org/en>, accessed on 30 December 2021).

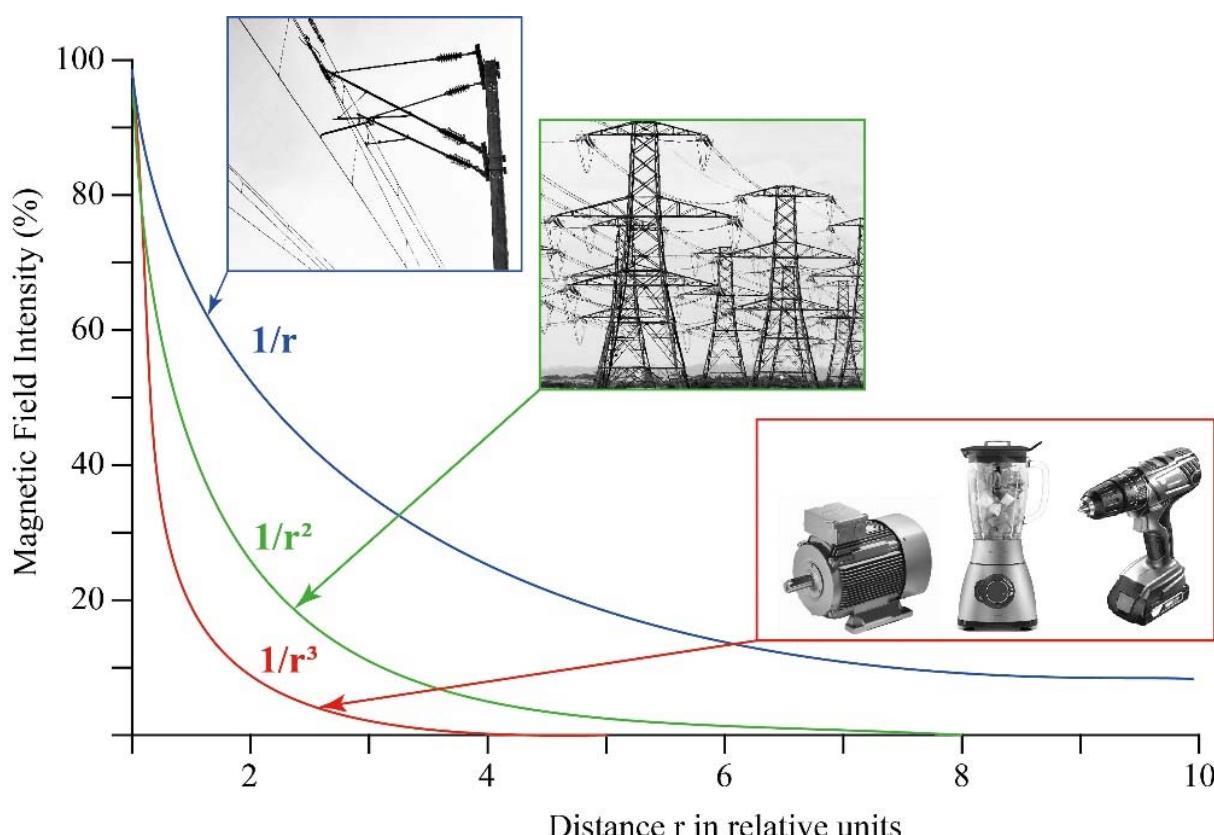


Figure 1. The magnetic field intensity decreases with growing distance from the field source.

In general, the type and extent of the cautionary policy that is chosen critically depends on the strength of the evidence for a health risk and the scale and nature of the potential consequences. In many countries, the adoption of a principle of caution or prudent avoidance implies the low-cost avoidance of unnecessary exposure as long as there is scientific uncertainty about its health effects [21,22]. However, still some policies are effective in preventing new situations with long-term exposure of children to MFs from overhead power lines, but these generally do not include underground cables and other sources of MFs [23]. Preventive measures and precautionary principles are necessary to warrant the reduction of exposure to children because of their greater sensitivity to ELF EMF [24–26]. The American Academy of Pediatrics set out new recommendations to decrease the adverse effects of exposure on children also to mobile phones [27].

2. Epidemiological Studies Evaluating MF and Cancer Relationships

It is known that epidemiological studies alone cannot be used to determine a clear cause and effect relationship when considering MF and cancer. This is mainly because epidemiological studies evaluate only the statistical associations between exposure and disease, which may not be necessarily caused by the exposure. Only the presence of a consistent and strong association between the exposure and the effect, a clear dose-response relationship, support that is provided by relevant animal studies, a credible biological explanation, and above all, if there is consistency between the studies can support cause and effect conclusions. In studies involving EMF and cancer most of these factors are generally missing. Studies of the potential health effects of EMF have concentrated on the MF because it is generally assumed to be the component that is most likely to have biological effects [28]. Since the first evidence determining the relationship between the ELF EMF and leukemia in children [29], epidemiological studies in this context increased and the IARC classified the ELF EMF in group 2B, a “possible carcinogen” to humans, whereas static electric and MFs are not classifiable as carcinogenic to humans (Group 3) [30]. Although

it is generally accepted that EMFs can exert biological effects, in general, epidemiological studies show a weak and sometimes inconsistent association between exposure to power-frequency fields (PFF) and cancer. In most cases, the studies fail to show a dose-response relationship [31,32]. The opposite happens in laboratory studies where PFF points towards causing or contributing to cancer (see below). The application of “Hill’s criteria” (i.e., strength, plausibility, specificity, biologic gradient, consistency, coherence, experimental evidence, temporality, and analogy) to laboratory and epidemiological studies shows a weak evidence for a causal association between cancer and the exposure to PFF [33].

Because cancer is one of the significant problems of global health, epidemiologic studies have faced the question of whether occupational and residential exposure to ELF EMF might be carcinogenic. There are three main explanatory hypotheses that appear in the literature: (i) the EM hypothesis, attributing EHS to EMF exposure; (ii) the cognitive hypothesis, assuming that EHS results from false beliefs in EMF harmfulness; (iii) the attributional hypothesis, considering EHS as a surviving strategy for pre-existing conditions [34]. Most of the epidemiologic studies explored the association between ELF EMFs and the susceptibility to different cancers. In the next section the residential/domestic and the occupational exposure to MF as related to cancer occurrence will be described.

2.1. Epidemiology of Residential/Domestic Exposure to MF

A survey of the literature indicates that residential exposure to EMFs is associated to an increased risk of cancers, particularly breast cancer, brain tumors, and leukemias. However, most of these studies are based on small numbers of high field-exposed cases [16,35,36] and an increasing number of studies does not support an epidemiologic association of adult cancers with residential MFs [35–37]. Many of the studies clearly have shortcomings, which often prevent any firm conclusions [38,39]. Moreover, the indirect measures of EMF exposure used may also correlate with other factors such as social status (e.g., age, race, gender) or environmental pollution. It is possible that these unconsidered and confounding factors may contribute to the cancer rates that are reported and also to the contrasting results that are reported in various EMF studies [40]. Indeed, EMF, which itself is not believed to be genotoxic, could influence carcinogenesis if it exerted either direct or indirect effects on target cells [41].

Another important issue is the exposure assessment. The exposure to MF can vary greatly over time and distance, has multiple sources, and is imperceptible and ubiquitous [42–45]. In exposed schools, children may experience a higher chance of receiving a mean exposure $>0.4 \mu\text{T}$ during school hours [46,47]; whereas those living in big buildings or using electric heating appliances in larger families had a generally higher level of personal indoor exposure [48]. Based on the known location of domestic and service MF sources, apartments can be reliably classified as high and low MF-exposed [49–51]. Methodologies for estimating MF at study residences as well as for characterizing the sources of uncertainty in these estimates have been developed [52]. In residential/domestic epidemiological studies, geographic information that is collected in an exact place of residence at the time of cancer diagnosis can provide several strategic geophysical elements for assessment [53]. The estimation of the overall exposure level from a single address is also informative [54]. In general, the public health impact of residential fields is considered limited, but the available data show the occurrence of both no impact and substantial impact possibilities [55].

2.1.1. Brain Tumor

The incidence of primary brain tumors has increased in many countries worldwide [56,57] and gliomas are the most frequent primary brain tumors in adults [58]. Residential exposure during childhood to EMF produced inconsistent results and a lack of an association when related to brain cancers, regardless of the exposure metrics that were used whether based on wire codes, distance, or the measured or calculated fields [59]. When focusing on the health effects, the most studied sources of ELF MF are power lines. Exposure to ELF MF that is emitted by power lines can be assessed by direct methods that

rely on measurements at a given place over a time range [60] or by individual monitoring through measuring ELF MF exposures throughout the day by wearable dosimeters [61]. Both methods give little or no information on historical exposure to ELF MF. Indirect methods include geographical information system (GIS) which have been used along with declarative data, such as residential history, to assess residential ELF MF exposure in the general population [62–65]. Case-control studies that are based on death certificates revealed an association between adult brain tumor mortality and living less than 50 m (odd ratios, OR 1.10 95%, CI 0.74–1.64) [66] or 100 m (OR 2.99, 95%, CI 0.86–10.40) [67] from power lines. In a recent work, significant associations were found between the cumulated duration living at <50 m to high voltage lines (50/60 Hz, 0.3 μT) and all brain tumors (OR 2.94; 95%CI 1.28–6.75) and glioma (OR 4.96; 95%CI 1.56–15.77) [65], confirming previous studies [35,66–69]. Contrasting results have been reported for brain tumors in children. In childhood brain cancer, with the exception of the possibility of a moderate risk increase in high cut-point analyses (0.3/0.4 μT), no increased risk was evident for different exposure metrics [70,71], whereas children whose MF exposure level was above 0.3 or 0.4 μT, an elevated risk of brain tumor was observed [72–74]. In residential areas, the transient electric and MFs would induce higher current density in the child's body than power frequency fields with similar field strength [75]. In some studies, there is no evidence for a role of ELF cellphone EMFs in childhood brain cancer [27].

2.1.2. Breast Cancer

Breast cancer threatens women with the highest incidence and second highest mortality rate of all cancers and in women aged 65 and older when nearly one half of all new breast cancer is diagnosed [76]. The excessive exposure to MFs increases the risk of female breast cancer, as demonstrated in several pooled or meta-analyses as well as subsequent peer-reviewed studies [69]. It is questionable whether chronic human exposure to MFs might affect melatonin secretion, its circadian rhythm, or both [77–80]. In general, no cumulative effects on melatonin secretion in humans have been found in response to MFs and this rebuts the "melatonin hypothesis" in which a decrease in plasma melatonin concentration (or a disruption in its secretion) would be correlated with the occurrence of breast cancers as a consequence of exposure to MFs [81–88]. Indeed, MF exposure correlates with an increased proliferative activity of the mammary epithelium, which is a likely explanation for the cocarcinogenic or tumor-promoting effects of MF exposure that is observed [89]. However, some authors found a motivation for going back to the melatonin hypothesis in relation to data, suggesting magnetosensory disruption by ELF MF in mammals, and magnetosensitivity in humans, along with the influence of MFs on circadian rhythmicity with a consequent disruption of non-photic sensory stimuli of various nature [90].

In studies that were based on the measurement of an electric bedding device, only premenopausal breast cancer (OR = 1.23; 95% CI: 1.01, 1.49, $p = 0.04$) showed a slight increased risk [91]. Breast cancer was found to increase with the number of years and seasons of bedding device use during sleep. Similar trends in dose response were shown in both premenopausal and postmenopausal women and for both estrogen receptor-positive and estrogen receptor-negative tumors. Therefore, there is a growing body of evidence that the use of electric bedding devices may increase breast cancer risk [92]. In terms of geographic variation of breast cancer rates, the results are inconclusive and do not support a major role of MF risk factors in the etiology of breast cancer [93].

2.1.3. Leukemias

Leukemia is the most common cancer in children [94]. The analysis of reports on childhood leukemia as related to exposure to MFs shows that a statistically significant association between MF exposure and childhood leukemia is found in almost all government or independent studies with an elevated risk of at least OR = 1.5, while a not significant or even suggestive association is reported in many industry supported studies [69].

Several meta-analyses showed a statistical association between childhood leukemia and a range of exposure 0.1–2.36 μT MF intensity [74,95–99]. Children that are exposed to elevated ELF-MF show relative risks of leukemia between 1.3 and 2 [100–102] and the highest exposure was associated with an increased risk of B-lineage acute lymphoblastic leukemia (B-ALL) when compared with lower exposures [103]. A significant association was observed during the night (OR = 3.21, 95% CI 1.33–7.80) between childhood leukemia and MF exposure [104].

Several epidemiologic case-control studies examined the association between childhood cancer risk and distance to high-voltage overhead transmission lines (HVOTL). Statewide, record-based case-control studies of childhood leukemia evidenced the occurrence of risk that was associated with greater exposure to MF that was generated in areas that were close to power lines [66,105–108]. Living in polluted regions and pre- and post-natal exposure to high voltage power lines has been described as risk factors of acute lymphoblastic leukemia (ALL) in people of low socioeconomic status Iranian population [109]. In children with ALL, ELF MF-exposure was found to have no impact on the survival probability or risk of relapse [110].

The occurrence of childhood acute leukemia (AL) has been studied around nuclear power plants (NPP). The results suggest a possible excess risk of AL in the close vicinity to NPP [111]. The small, but statistically significant increased incidence of AL in the surrounding of some NPP have motivated governments to work toward a better understanding of the main causes of AL long-term strategic research agendas through interdisciplinary and international efforts [112].

MFs are not the only factor that varies in the vicinity of MF sources, complicating interpretation of any associations. Several reports demonstrate that MFs that are generated by different sources are not an important cause of leukemia both in adults [37,113,114] and children in many geographical areas [12,113–120]. Moreover, exposure levels in some big cities are always significantly far lower than 0.3–0.4 μT [121].

The associations that were observed between power lines exposure and childhood leukemia appears to be not related to mobility [122,123]. No indications were also found of an association of risk for people that are exposed to magnetic fields from underground [124] and above ground lines [106,107] or of a trend in risk with increasing MF for leukemia. Residential proximity to transformer stations has been associated with a borderline risk of childhood cancer [125].

Distance from HVOTLs during the year of birth is unlikely to be associated with an increased risk of leukemia [73] and little evidence was found between exposure to MF inside infant incubators and increased risk of childhood leukemia [126]. No statistically significant association was observed between wire codes and childhood leukemia [127]. Recently, a synoptic analysis provided evidence that the risk of childhood leukemia is not increased by exposure to ELF EMFs, suggesting that IARC's classification of ELF EMF needs revision [128].

By considering both significant and not significant correlations between MF and leukemia, no major environmental risk factors (including MFs) have been established as major contributors to the global leukemia burden, although distinct incidence patterns by geography, age, and sex suggest a role of the environment in its etiology [129]. The results may be affected by several sources of bias: analyses that are based on continuous exposure show no exposure-disease association, while incoherent exposure-outcome relationships characterize analyses that are based on categorical variables [130].

2.1.4. Other Cancers

EMF in the home-environment (color tv, computer monitor, microwave-oven, cellular phone, etc.) might act as potential contributing factors for the development of cancer, as well as exert indirect effects on humans. Microwave exposure induces L-amino acids change to D-amino acids, and exposure of the human body to microwaves over a long period of time may contribute to induction of cancer [131]. Exposure to EMF that is

generated by electric blankets has been suggested to increase the risk of hormone-dependent cancers such as testicular [132] and prostatic [133], but was not significantly associated with endometrial cancer risk [134]. MFs of industrial frequency have been shown to be a risk factor for occurrence of oncological diseases in the population and an increased incidence of malignant tumors has been noted as the induction of the magnetic field that is produced by HVOTL increases [135]. The incidence of melanoma has been linked to the distance to FM broadcasting towers. A correlation between melanoma incidence and the number of locally receivable FM transmitters was found when geographic differences in melanoma incidences were compared with the magnitude of this environmental stress. Therefore, melanoma might be associated with exposure to FM broadcasting [136]. By considering pregnancy duration, neonatal birth weight, length, head circumference, gender, and con-genital malformations, no significant effects of ELF EMFs were observed; however, precautionary measures are necessary to reduce exposure to EMFs by pregnant women [137].

The Supplementary Table S1 reports the relationship between MFs and cancer in epidemiologic studies that are related to domestic/residential exposure to MFs. The type of cancer, study design, source of MF, range of MF exposure, location of the epidemiologic study, and the main conclusions are reported. Figure 2 summarizes the epidemiology of residential/domestic exposure to MF.

2.2. Epidemiology of Occupational Exposure to MF

During the last few decades, the intensity level of the EM occupational environment has dramatically increased. By job category, the most highly exposed jobs ($>0.23 \mu\text{T}$) included electronics workers, cooks and kitchen workers, cashiers, bakery workers, textile machine operators, and residential and industrial sewing machine operators [138]. The main components of EM pollution are in the ELF (10–300 Hz) and in ultra-low (ULF: 0–10 Hz) frequency bands [139]. Occupational epidemiology reveals that exposure to ELF EMF is generally greater than that in the general population and concerns a large number of workers in a variety of industries (see [140,141] for a historical overview of the occupational EMF epidemiology).

MF exposure limits are more than a thousand times higher than the magnitudes that are associated with the cancer risks that are observed in epidemiologic studies, leaving millions of workers exposed to MF in this large gray area where the public health consequences are unclear [140,141]. The International Labor Organization defines occupational exposure as “exposure of a worker received or committed during a period of work” [142] while the ICNIRP defines occupational exposure as “all exposure to EMF experienced by individuals as a result of performing their regular or assigned job activities” [143].

As discussed for residential/domestic exposure to EMF, in the case of occupational exposure, contrasting results have also been presented in relation to the co-occurrence of different cancers including brain and breast cancer and hematological malignancies.

2.2.1. Brain Cancer

Occupational exposure to ELF EMF is a suspected risk factor for brain tumors; however, the literature reports contrasting results. In adults, some meta-analyses of occupational studies indicate a slightly higher risk for electrical workers, suggesting a small increase in brain cancer risk [61,144,145], including childhood brain tumors [146], while others found no evidence to support the hypothesis that exposure to MFs is a risk factor for gliomas or meningiomas [147–150].

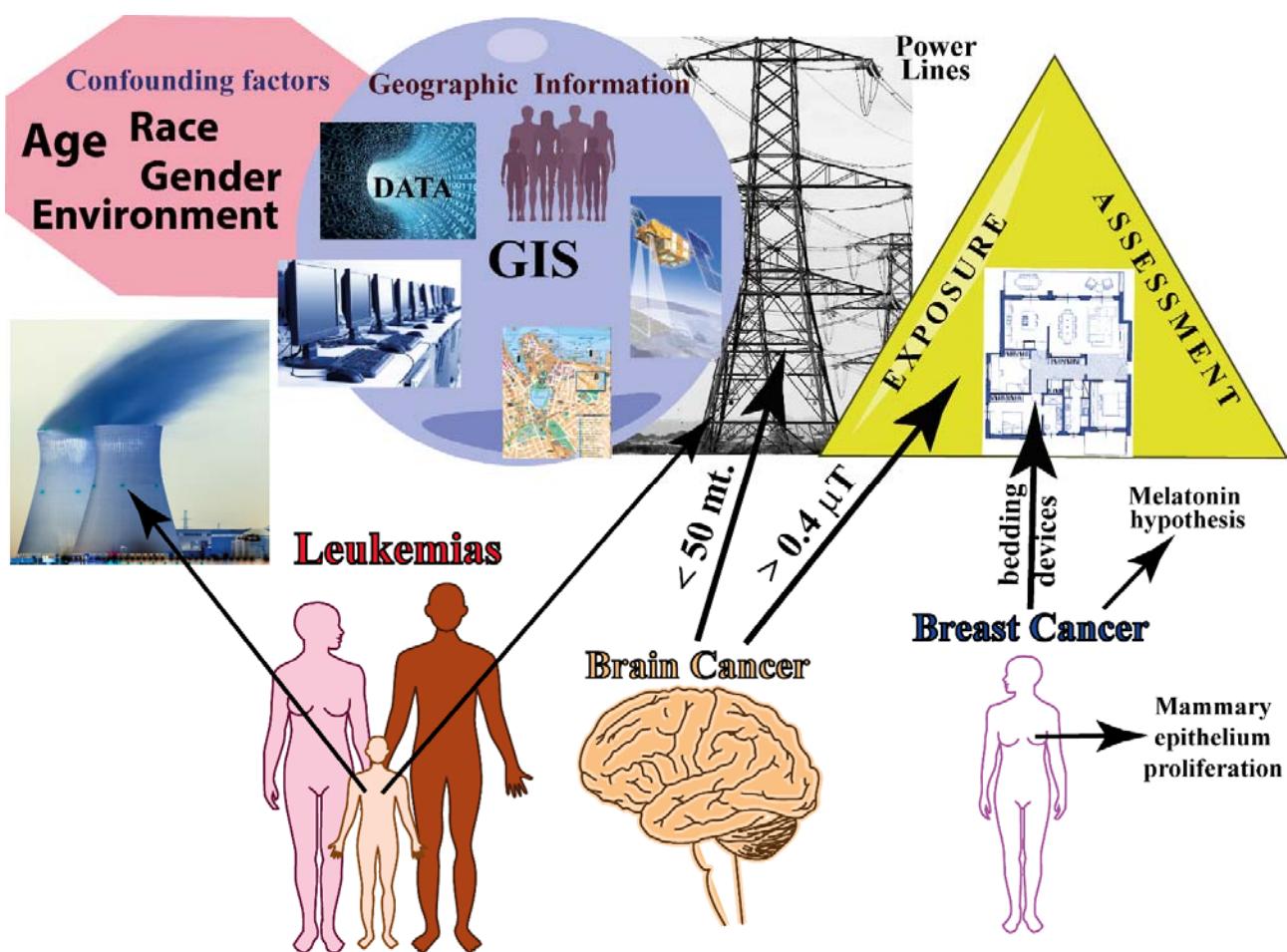


Figure 2. Summary of epidemiology of residential/domestic exposure to MF. The major confounding factors in epidemiological studies are shown along with the main geographical information that is based on GIS (data management, hardware/software, topography, remote detection, and population demographics). Exposure assessment needs to be evaluated both outside and inside the residence. The three major cancers are represented: leukemia affects mainly childhood; brain cancer increases with decreasing distances from EMF sources; and breast cancer is associated with mammary epithelium proliferation.

2.2.2. Breast Cancer

Several studies have evaluated the evidence linking women's occupation and workplace exposures to breast cancer. Overall, the data do not suggest that occupational exposures to EMF increases the risk of breast cancer [144,145,151,152] with some exceptions [153]. Some studies have found an effect when looking at overall risk elevations in the women studied [154] with increased risk among postmenopausal women but not premenopausal women [155]. The hypothesis that daytime occupational exposure to MF enhances the effects of nighttime light exposure on melatonin production (see above the "melatonin hypothesis") has been provided [153]. Occupational MF exposure has also been suggested as a risk factor for breast cancer in men. An elevated risk of breast cancer was found in men who were exposed to 0.6 T when compared to those with exposures <0.3 T; however, large case-control studies of breast cancer in men that have been conducted to date provide limited support for the hypothesis that exposure to MF increases the risk breast cancer in men [156].

2.2.3. Leukemias

Epidemiological studies addressing occupational ELF MF exposure and risk of leukemia in adults have yielded slightly increased risks in exposed workers. Some studies show a positive correlation between occupational MF and leukemias [157–160] and have suggested that stronger effects may be observed for acute myeloid leukemia (AML) [161,162], chronic myeloid leukemia (CML), ALL [138], lymphatic leukemia (LL) [163], and for chronic lymphocytic leukemia (CLL) [162]. In general, however, no clear exposure-response pattern has emerged from the studies that evaluated exposure levels and some results do not support an association between occupational ELF MF or electric shock exposure and AML [164].

Differences between the study designs or the populations that were studied might be the cause of lack of consistency regarding the type of leukemia that is associated with MF exposure, and still no firm conclusions can generally be drawn based on the existing evidence [165–167]. No association was found between childhood leukemia risk and parental occupational exposure to ELF EMF [168–170].

2.2.4. Other Cancers

In men, exposure to EMF showed an increased incidence of tumors of the liver, biliary passages, kidney, and pituitary gland; for these cancer sites an exposure-response relation was indicated [171]. There was very limited evidence for associations between occupational ionizing radiation and testicular cancer, while there were some positive associations for EMF [172].

Women that were exposed to MF showed an increased incidence of astrocytoma I–IV, for cancer of the corpus uteri and multiple myeloma, with a clear exposure-response pattern [171].

For both men and women, there was weak support for the hypothesis that occupational MFs exposure increased the risk of non-Hodgkin lymphoma [173], acoustic neuroma [174,175], and thyroid cancer [176], while sources that produce ELF fields were not associated with neuroblastoma in offspring [177].

MF has previously been associated with mortality from acute myocardial infarction (AMI) and arrhythmia but not from chronic coronary heart disease (CCHD) or atherosclerosis in electric utility workers. For cumulative exposure, no association was observed with mortality from AMI or CCHD, indicating no exposure-related risk increase for AMI mortality, which does not confirm previous results [178].

Supplementary Table S2 reports the relationship between MFs and cancer in epidemiologic studies that are related to the occupational exposure to MFs. The type of cancer, study design, source of MF and occupation, range of MF exposure, location of the epidemiological study, and the main conclusions are reported. Figure 3 summarizes the epidemiology of occupational exposure to MF.

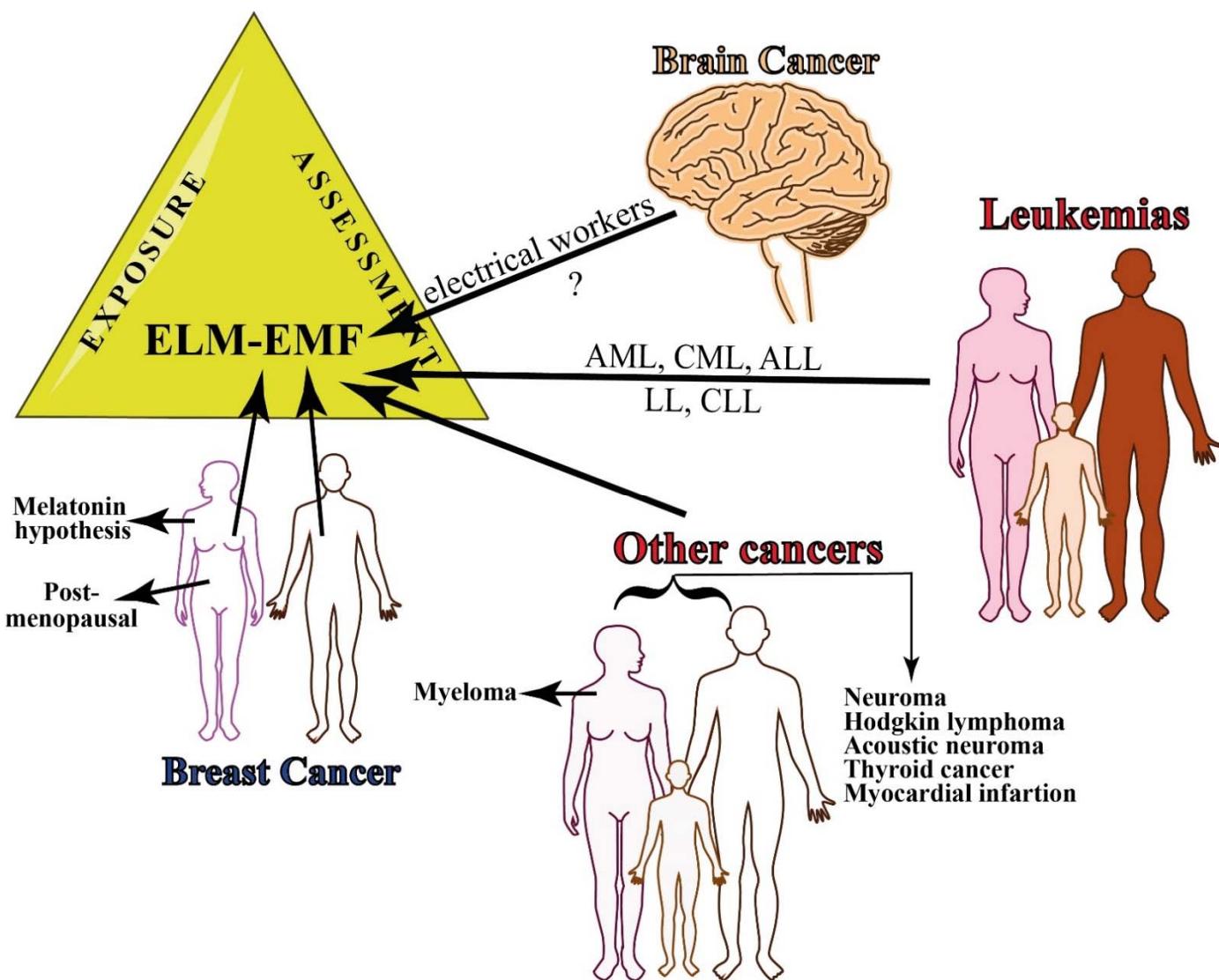


Figure 3. Summary of epidemiology of occupational exposure to MF. There is still little evidence on the relationship between occupational exposure to EMF and brain cancer, whereas several leukemias have been associated with continuous exposure to ELF-EMFs. Breast cancer occurs both in women and men and the risk increases in men that are exposed to 0.6 T. Other cancers that are associated with MF exposure include myeloma in women and several other types of cancer in both women and men.

3. In Vivo and In Vitro Effects of Magnetic Fields on Cancer

In many studies ELF exposure causes significant changes in cell survival, cell cycle progression, DNA integrity, and proliferation [40,179]. The cellular response to ELF MF may depend on many parameters including osmolarity, frequency, waveform, the strength and the exposure duration of the electromagnetic field, genetic/biological characteristics of the cells [180], specific metabolic state, or the specific stage in the cell cycle [180,181]. On the other hand, a common effect of exposure to SMF is the promotion of apoptosis and mitosis, but not of necrosis or modifications of the cell shape [182,183]. The unbalance of the apoptotic process could be linked to Ca^{2+} fluxes that are, in turn, dependent on the effect on the plasma membrane that is exerted by SMF [184–187]. Other possible effects of SMFs that may lead to perturbation of the apoptotic rate, such as an alteration of the gene pattern expression or the increase of oxygen free-radicals, could be, in turn, a co-carcinogenic

factor leading normal cells, most likely with other sub-lethal changes, to contribute to the development of diseases [183].

The reduction of cell proliferation due to MF has been attributed to the interference of signal transduction processes due to the tangential currents that are induced around the cells [188]. The poor reproducibility might be caused by the period-dependent converse cell growth due to the MF and might explain the adverse effects that are observed in several experimental investigations [189].

Quantitative analyses of protein kinases C (PKC) expression patterns demonstrated the translocation of PKC from the cytosolic to the membrane fraction was not affected by MFs [190]. The phosphorylation of extracellular signal-regulated kinases 1/2 (ERK1/2) is increased in response to ELF MF; however, the small increase in ERK1/2 phosphorylation is probably insufficient to affect proliferation and oncogenic transformation [191]. Furthermore, repeated ELF EMF exposures did not show consistent response profiles to time courses of immediate early genes, apoptotic genes, cell proliferation, and stress response [192].

Small changes in transcription may occur in response to MFs [193,194]. Exposure to 900 MHz identified a differential expression of functional pathways genes [195]. Indeed, epigenetic changes, including modifications of histones and microRNA expression and DNA methylation, can be associated to ELF MF exposure [196–198]. However, in HeLa cells, RNA polymerase-catalyzed RNA synthesis as well as DNA polymerase-catalyzed DNA synthesis were found to not be statistically significantly affected by 60 Hz 0.25–0.5 T exposure for 0–60 min [199].

Aberrantly-expressed serum exosomal miRNAs upon MF exposure suggests a series of informative markers to identify the exact dose of MF exposure [200]. ELF MF exposure stabilizes active chromatin, particularly during the transition from a repressive to an active state during cell differentiation [201]. Membrane receptors could be one of the most important targets where ELF MF interacts with the cells [202] and RAS proteins, a member of a large family of GTP-binding proteins that are involved in intracellular signal pathways, may participate in the signal transduction process of 50 Hz MF [203]. EMF was also found to affect the proteasome functionality, inducing an increase in its proteolytic activity [204].

To answer how MF may cause cancer, the action of MF as mutagenic agents and MF involvement on chemical reactions that generate free-radicals have been considered. MF does not seem to exert mutagenic effects and the generation of free-radicals that might be linked to several other factors, beside the variability of MF exposure [40]. In the next part of this section, the current data that are related to studies on human in vitro and cell-free systems as well as in animal models will be discussed. This section will also discuss the role of reactive oxygen species (ROS) and reactive nitrogen species (RNS) and the role of radical pair (RP) formation in MF-cancer relationship.

3.1. Studies on Human Cells (In Vitro Cellular Studies and Cell-Free Systems)

Exposure to ELF MFs combined with ionizing radiation do not suggest any synergistic or antagonistic effects on human blood cells [205], whereas, by using sister chromatid exchanges and comet assays, a slight but significant decrease of cell proliferation was evident in blood cells that were exposed to 980–1020 μT [206].

Dermal fibroblasts that were exposed to 1 mT from one to five hours showed a reduction of their viability [207], while the rePETitive exposure to MF with ELF induced DNA double-strand breaks and apoptosis in lung fibroblasts [208]. However, a longer exposure (48–72 h) to a reduced MF intensity (20–500 μT) did not result in any appreciable effect in the structural morphology and proliferation of human fibroblasts [209].

Exposure of glioma cells to MF of 1.2 μT intensity for three hours revealed changes in both gene and protein expression. Microarray results showed an up-regulation of five genes, whereas 25 genes were down-regulated upon MF exposure, suggesting a response in the glioma cells to the MF treatment [210]. Proteomic studies on glioma SF767 cells show a cytoskeletal intermediate filament protein increased following a low-level MF [211]. On

the other hand no mechanisms that would explain the reported association between MF and carcinogenesis were observed in H4 glioma cells [212].

In human keratinocytes, ELF EMF induces a slight oxidative stress that does not overwhelm the metabolic capacity of the cells or have a cytotoxic effect when the cells are exposed to 25–200 μ T MF intensity [213] but it does not affect melanin synthesis or skin pigmentation [214].

The adherence of leucocytes and T-lymphocytes that were taken from cancer patients strongly increased following exposure to sinusoidal MF at 50 Hz and 0.5–10 mT MF intensity for one hour [215,216], whereas a low-frequency pulsing electromagnetic field of 45 mT for three hours induces cell death in native peripheral blood mononuclear proliferating cells that were isolated from AML patients [217].

ELF MF increases the rate of cell death in normal human lymphoblastoid cell lines and is ineffective on genetic instability syndromes cell lines, i.e., Fanconi anemia group C and ataxia telangiectasia, suggesting that the response of cells to ELF MF is modulated by genes that are implicated in genetic instability syndromes [218]. Exposure to a pulsating MF 50 Hz, 45 mT three times for three hours was found to protect U937 human lymphoid cell lines against puromycin-induced cell death in a cell density-dependent manner (an increased density induced cells death and prevented puromycin-induced cell death) [219].

The effects of 6- and 10-T SMFs was studied on HL-60 cells to evaluate the expression of protooncogenes. It was found that exposure to a strong MF gradient induced c-Jun expression, whereas no alteration of the expression of the *c-fos* and *c-myc* protooncogenes was observed [220]. In the same cell line, exposure to LF EMF to lower intensity (5, 300, and 500 μ T) did not affect calcium signaling [221].

An ELF magnetic field was also found to influence the early development of mesenchymal stem cells that were exposed for 23 days to 50 Hz and 20 mT intensity [222], whereas no variations in surface morphology and cell death occurred between the control and the exposed osteosarcoma human cells that were exposed to 50 Hz and 0.5 mT, although significant changes were noted in cell growth [223].

Supplementary Table S3 reports the effects of MFs on human cell lines. The type of cell, response to MF, range and duration of MF exposure, materials and methods that were used, and the main conclusions are reported.

3.2. Studies on Animals

Exposure to MF was found to be a cancer promoting factor in animal experiments; animal studies are often used in the evaluation of suspected human carcinogens [224]. However, discrepancies in results were found to be associated with the use of different sub-strains, different sources for diet, environmental conditions, and MF exposure metrics [225–227]. The continuous monitoring of both MF and other environmental parameters is, therefore, an important part in the overall quality of the obtained results [228]. Experiments with animals were also important to determine which genetic and environmental factors are critical for potential carcinogenic or cocarcinogenic effects of ELF EMF exposure [229]. Furthermore, the genetic background was found to play a pivotal role in effects of MF exposure [230]. To study the effects of MFs on cancer cell growth, multiple exposure levels have been performed using mice and rats as experimental models.

3.2.1. Studies on Mice

Several studies revealed that there were not significant effects of MF on cancer in mouse studies, failing to support the hypothesis that acute MF exposure causes DNA damage. The experimental mice were injected with mammary murine adenocarcinoma to investigate the interaction between a 50 Hz, 2 mT MF exposure and cell growth.

Neither the time of tumor cell injection nor the time of exposure produced differences between the unexposed, sham, and exposed mice [231] and no association between exposure and the incidence of benign or malignant tumors was found in squamous cell carcinomas of mice that were exposed to 60 Hz, 2 mT [232]. Also, the results do not sup-

port the hypothesis that acute MF exposure causes DNA damage and apoptosis in the cerebellums of immature mice that were exposed to 60 Hz and 1 mT for 2 h [233,234].

Exposure to 50 Hz and 500 μ T MF did not alter responses of inflammatory genes and the activation of splenic lymphocytes in mice [235] and was not a significant risk factor for hematopoietic diseases, even when high exposure levels were used (50 Hz, 1 mT for 7 days) [236]. In mice, as revealed in humans (see above), long-term and continuous exposure to simulated powerline MFs (0.5–77 μ T) did not result in a decreased nocturnal melatonin secretion [85].

The use of NI MFs has shown early promise in a number of animal studies as an effective tool against many types of cancer (see also below). The effect of varying durations of MF exposure on tumor growth and viability has been studied in mice that were injected with breast cancer cells by using an *in vivo* imaging system. The results showed the potential of MF in cancer therapeutics, either as adjunct or primary therapy [237].

Long-term exposure showed a significant effect of MF on mice. When a parental generation of six week-old OF1 mice was exposed to an artificial ELF MF (50 Hz, 15 μ T) for 14 weeks, activated partial thromboplastin time and reptilase time values significantly increased in the female mice, which also showed a very significant shortening of the prothrombin time that was associated with ELF MF exposure [238]. A chronic exposure of mice to a 60 Hz and 110 μ T intensity was found to influence some hematologic parameters and the weight of the liver and also caused spleen hyperfunction [239]. Long-term exposure to MF (50 Hz, 50 μ T) was found to be a significant risk factor for neoplastic development and fertility in C57BL/6NJ female mice and C3H/HeNJ male mice [240].

3.2.2. Studies on Rat

Both positive and negative effects of MF have been demonstrated in studies using rats. Artificial MFs of 1 mT that was administered for seven weeks was not carcinogenic nor cancer-promoting for colon carcinogenesis in male Sprague–Dawley rats [241]. No overall effects of 60 Hz and 1 mT MF on splenomegaly or survival were found in the exposed Fischer/344 rats. In addition, no significant and/or consistent differences were detected in the hematological parameters between the exposed and control rats [242]. MF exposure for 14 weeks to 6.45 μ T did not appear to be a strong co-tumorigenic agent in Sprague–Dawley female rats mammary, lung, and skin models [243]. On the other hand, EMFs resulted in significant alterations in cell adhesion mechanisms when histological, immunohistochemical, and histopathological analyses were performed on Wistar albino rats that were exposed for six months to 5 mT MFs [244]. MF exposure (50 Hz, 100 μ T for 2–26 weeks) of female Sprague–Dawley rats resulted in an increased proliferative activity of the mammary epithelium which was associated with the cocarcinogenic or tumor-promoting effects of MF exposure that was observed in the 7,12-dimethylbenz(a)anthracene model of breast cancer [89], and mammary tumorigenesis [245].

The Supplementary Table S4 reports the effects of MFs on both mice and rats. The type of cell, response to MF, range and duration of MF exposure, materials and methods used, and the main conclusions are reported. Figure 4 summarizes the *in vivo* and *in vitro* effects of magnetic fields on cancer from studies on human cells (including cell-free systems) and in animals (mice and rat).

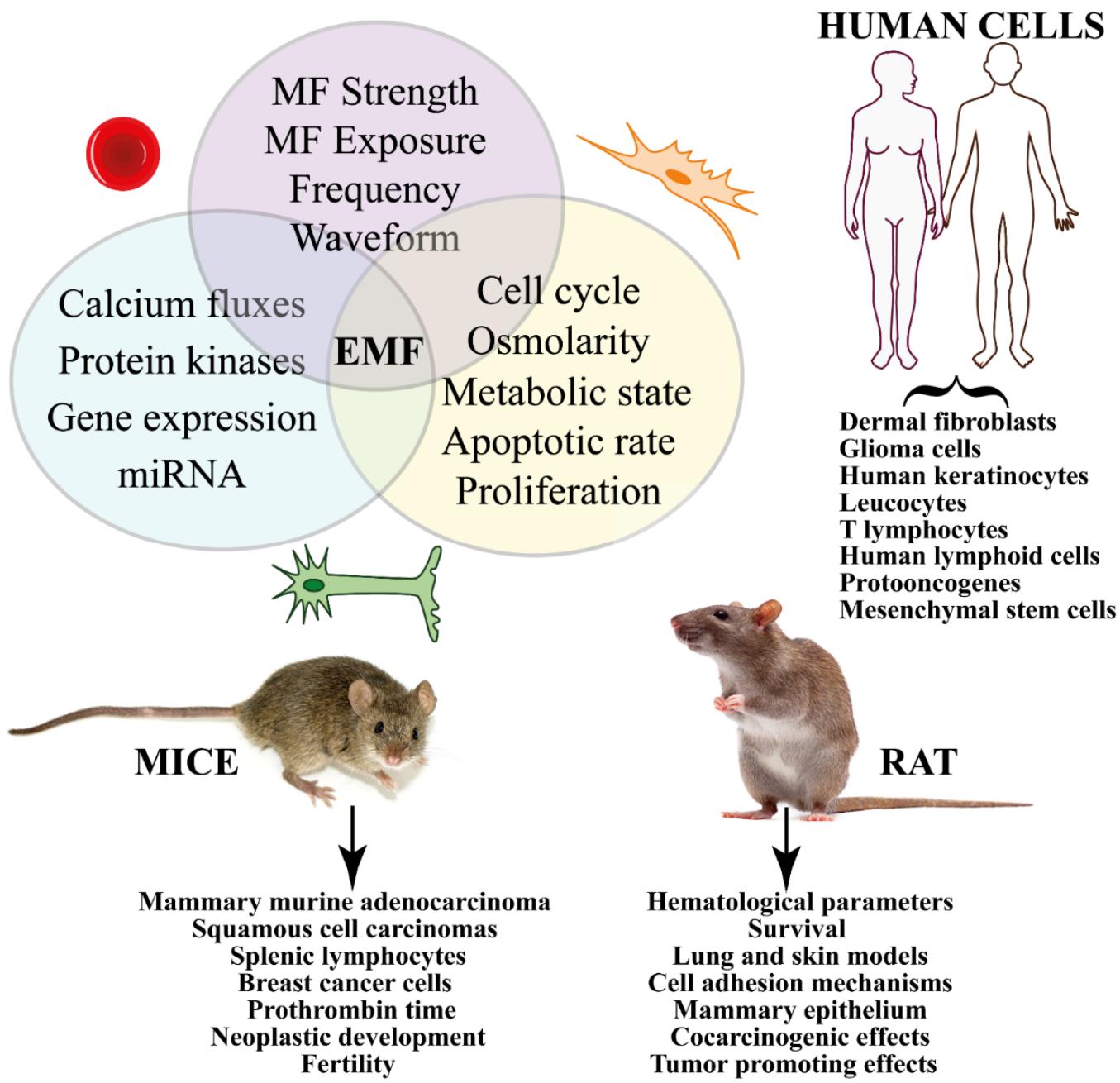


Figure 4. Summary of the in vivo and in vitro effects of MFs on cancer from studies on human and animal cells. Different treatments (e.g., strength, duration, frequency, etc.) induce signal transduction pathways that eventually trigger gene expression. In vitro studies show significant effects on cell cycle, proliferation, and apoptosis. Human cells have been used to evaluate the effect of MF on several cancer types, whereas animal experimentation has been focused on mice and rats.

3.3. Involvement of Reactive Oxygen Species (ROS) and Reactive Nitrogen Species (RNS)

ROS (such as superoxide [$O_2^{\bullet-}$] and hydrogen peroxide [H_2O_2]) and RNS (e.g., nitric oxide [NO^{\bullet}]) are generated during the oxidative cell metabolism. The cellular oxidative stress depends on the balance between the production of ROS and RNS and the activity of the antioxidant system. Excessive ROS/RNS, which is caused by the deregulated redox homeostasis, is a hallmark of disease [246]. Free-radical-scavenging enzymes, such as catalase (CAT), glutathione peroxidase (GPX), and superoxide dismutase (SOD), are the first line of defense against oxidative injury [247]. After EMF exposure, significant

variations in the total antioxidant activity; vitamin E and vitamin A concentrations; increase of malondialdehyde (MDA, a product of polyunsaturated fatty acid peroxidation which is used as an indicator of oxidative stress in cells and tissues); and plasma selenium concentration in erythrocytes have been observed [248]. A recent review concluded that most animal and many cell studies show increased oxidative stress that is caused by MF [249]. Free-radical formation and the consequences of their effects on living systems explains the increased cancer risks that are associated with mobile phone use, occupational exposure to MF, and residential exposure to power lines [250].

ROS may be involved in RP reactions that have been considered as one of the mechanisms of transduction that is able to initiate EMF-induced oxidative stress [19]. It is known that applied MFs and magnetic isotope substitution can alter the rates and product yields of free-radical reactions with the formation of transient paramagnetic intermediates in non-equilibrium electron spin states. The most common source of spin-chemical effects are organic RPs. Typically formed in a singlet (S) or a triplet (T) state by a reaction that conserves electron spin, RPs interconvert coherently between their S and T states as a result of the Zeeman, hyperfine, exchange, and dipolar interactions of the electrons and the nuclear spins to which they are coupled [251]. Applied MFs alter the extent and timing of the $S \rightleftharpoons T$ interchange and hence the yields of products that are formed spin-selectively from the S and T states [252]. MFs and spin effects have proven to be useful mechanistic tools for radical mechanism in biology [252] and the RP mechanism (RPM) has been associated with increased levels of ROS [253]. Moreover, the role of cryptochromes as the putative magnetosensitive molecules in magnetoreception has been considered in the RPM and discussed as related to cancer-relevant biological processes [254]. The results are also consistent with MF effects on light-independent radical reactions [255].

Significant increases in ROS levels have been found to influence the hepatic redox state [256] and were observed in several cell lines just after the end of ELF EMF exposure [213,257–265]. ELF EMF exposure also elevates the expression of RNS and $O_2^{\bullet-}$, which are countered by compensatory changes in antioxidant CAT activity and enzymatic kinetic parameters that are related to cytochrome P450 (CYP-450) and CAT activity [266]. Moreover, the modulation in kinetic parameters of CAT, CYP-450, SOD, and MDA concentration and iNOS enzymes in response to ELF EMF [267–269] and the negligible effects on GPX [270] indicates an interaction between the ELF EMF and the enzymological system. SMF promoted the release of ferrous iron (Fe^{2+}) and induced the production of ROS in osteosarcoma stem cells [271]. Superoxide increased in the micronucleus and mitochondria with an exposure-response relationship and cytosolic superoxide increased at 10 μ T in SH-SY5Y and C6 cells [272]. These results confirm that the threshold for biological effects of ELF MFs may be as low as 10 μ T.

In liver tissue of female rats, long-term ELF-MF exposure enhanced the oxidative/nitrosative stress and might have a deteriorative effect on cellular proteins rather than lipids by enhancing 3-nitrotyrosine formation [273]. MF appears to induce apoptosis effects through oxidative stress [274] and mitochondrial depolarization [275], whereas the influence of correlations between ELF EMF and vitamin E supplementation have been shown on antioxidant enzyme activity in AT478 malignant cells in vitro [276]. In embryonic stem cell-derived embryoid bodies, exposure to 10 mT MFs increased ROS [277].

Supplementary Table S5 reports the effects of MFs on ROS and RNS. The type of cell/tissue/organ, response to MF, range and duration of MF exposure, materials and methods used, and the main conclusions are reported. Figure 5 summarizes the involvement of ROS and RNS in the cellular and organismic responses to MFs.

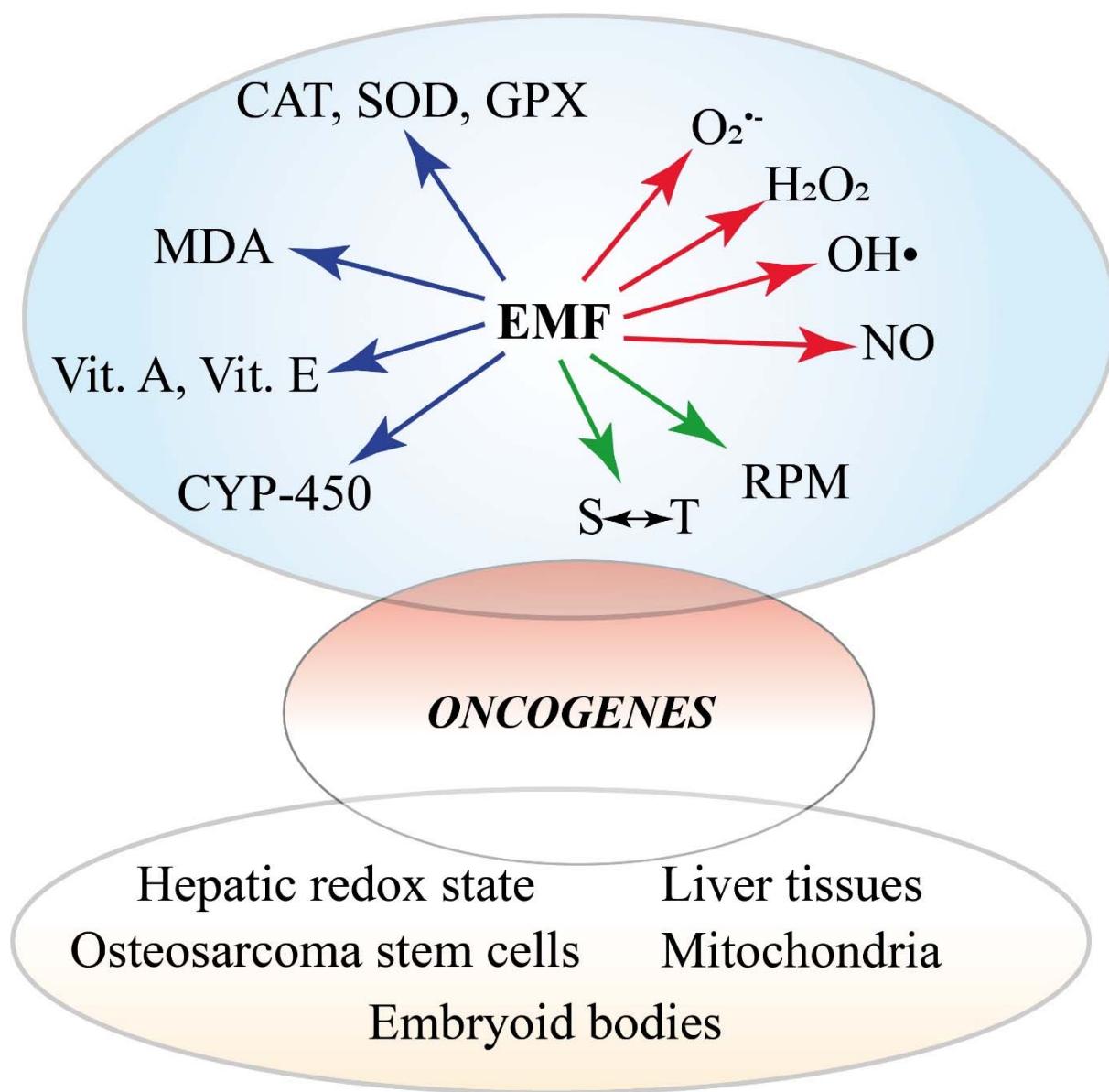


Figure 5. Summary of the involvement of ROS and RNS in the cellular and organismic responses to MFs. There are three major effects of varying MFs that are reported. MFs alter the redox status of the cell by affecting the activity and gene expression of ROS-scavenging systems, including CAT, SOD, GPX, vitamins, and monooxygenases. Membrane degradation is evidenced by MDA detection. On the other hand, MFs trigger the production of ROS and NOS and early events involving the radical pair mechanism and spin-chemical effects. The altered oxidative status eventually induces the expression of oncogenes. The alteration of the oxidative status is also evident at the subcellular, cellular, tissue, and organ level.

4. Magnetic Fields and Cancer Theranostics

So far, we have discussed the possibility that exposure to MFs may be correlated with cancer. However, MFs have been widely used for cancer diagnostic and therapeutic applications. In this section, the use of MRI as a cancer diagnostic and therapeutic method and the use of magnetic nanoparticle for cancer treatment will be discussed.

4.1. Magnetic Fields and Cancer Diagnosis

MRI, a medical application of nuclear magnetic resonance (NMR), uses strong MFs, MF gradients, and radio waves to generate images of the organs in the body [278]. There

is a trade-off between MF dose effects and the image quality of MR-guided radiotherapy systems and the MF strength may affect the severity of MF dose effects [279], such as the electron return effect [280]. MRI has also been used in combination with positron emission tomography (PET) and has a strong potential clinical use for the imaging of several forms of cancer [281]. Using ultrasmall superparamagnetic particles of iron oxide (USPIO), it is possible to enhance the power of MRI for noninvasive diagnostics of different types of cancer [282,283] (see also 4.2). Since alterations in the Na^+ ion concentration may potentially be used as a biomarker for malignant tumor diagnosis and prognosis, Na-23-magnetic resonance imaging (Na-23-MRI) was found to have potential as a direct noninvasive *in vivo* diagnostic and prognostic biomarker for cancer therapy, particularly in cancer immunotherapy [284]. The potential role of MRI in the detection of several cancers will be discussed in the next paragraphs.

4.1.1. Brain and Glioma Cancer

MRI has allowed the characterization and diagnosis of human brain cancers in spatial and volumetric analysis [285], as a substitute for biopsies [286], in glioma genotyping [287], brain cancer classification [288,289], as a non-invasive tool for simultaneous and automated tumor segmentation [290], and to investigate the early stages of slow-growing invasive tumors [291–294]. MRI is used before treatment and at the end of treatment or disease progression [295,296] and to assess neurological complications of cancer treatment [297]. PET/MRI is used in brain tumor grading and staging [298,299] as a diagnostic and therapeutic strategy for glioma [300] and for improved diagnostic and therapeutic assessment in pediatric, teenage, and young adult brain tumors [301]. ^{11}C -methyl-L-methionine (C-11-MET) PET/MRI was found to improve the diagnostic accuracy to differentiate treatment-related changes from true progression in recurrent glioma [302] and was useful for the assessment of isocitrate dehydrogenase (IDH) status [303]. Perfusion MRI is used to differentiate glioma from brain metastasis [304], whereas dynamic glucose-enhanced (DGE) MRI is a feasible technique for studying brain tumor enhancement reflecting differences in tumor blood volume and permeability with respect to a normal brain [305]. MRI-coupled fluorescence molecular tomography (MRI-FMT) determines epidermal growth factor receptor status in brain cancer [306] while advanced MRI techniques contribute to biological and imaging features of glioma and immune system interactions [307] and in the clinical management of adult gliomas [308]. The application of gadofluorine-M (GfM) results in superior delineation of experimental glioma compared with conventional MRI techniques [309], whereas the labeling tumor cells with superparamagnetic iron oxide (SPIO) and performing an MRI scan dynamically monitors the development and biological behavior of glioma at a very early stage [310]. Vascular, extracellular, and restricted diffusion for cytometry in tumors MRI has been used as a potential test for diagnostic stratification to investigate the tissue microstructure in glioma [311], whereas dynamic contrast-enhanced (DCE) MRI detects increases in gadolinium (Gd)-enhancement of brain tumors [312] and provides an unambiguous indication of the brain tumor photodynamic therapy outcome [313]. The enhancement on MRI may assist in identifying HER2 overexpression in breast cancer brain metastases [314].

4.1.2. Head and Neck Cancer

According to the National Cancer Data Base, head and neck cancer accounted for 6.6% of all new cancers [315]. The use of PET and MRI, separately or combined, has been successful for assessing the metastatic lymph nodes in patients with head and neck cancer and offers advantages in staging with regard to increased anatomical details and radiation dose reduction [316–318]. Diffusion-weighted imaging (DWI) and intravenous (IV) contrast T1 dynamic perfusion imaging are a valid support for the functional MRI of tumors of the head and neck [319–322] and algorithms for automatic head and neck 3D tumor segmentation from MRI have been developed [323]. For head and neck cancer, MRI-guided radiotherapy achieves clinical outcomes that are comparable to contemporary series

reporting on intensity-modulated radiotherapy (IMRT) [324] and the use of targeted 3 T MRI was found to be useful for defining the presence and extent of large nerve perineural spread in head and neck cancers [325]. Retrospective image fusion of PET/MRI for the assessment of the extent of the primary tumor (T stage) and metastasis to regional lymph nodes (N stage) has been evaluated [326], whereas USPIO-enhanced MRI in patients with a clinical neck cancer was able to differentiate borderline-sized lymph nodes [327].

4.1.3. Thyroid Cancer

Thyroid cancer is one of the fastest growing cancer diagnoses worldwide [328]; it is the most common endocrine malignancy in children [329], it is three times more frequent in females [330], and poorly differentiated and anaplastic thyroid carcinomas represent a challenge to physicians on the basis of the current cancer treatment modalities [331]. MRI shows sensitivity and specificity to diagnose recurrent thyroid carcinomas [332], thyroid cancer cervical lymph node metastases [333], papillary carcinomas [334], and is more sensitive than ultrasonography in detecting central compartment metastases in papillary thyroid carcinoma [335]. However, MRI is inadequate for the detection of metastases in small lymph nodes [336]. Esophageal invasion by thyroid carcinoma was accurately predicted with MRI [337], whereas MRI, DWI-MRI, and MRI-based computer-aided diagnosis (CAD) allow the differentiation of thyroid nodules whether benign or malignant [338–340] and for the detection of inner and outer thyroid lamina invasion [341]. PET/MRI of the neck is superior to PET-computed tomography (CT) in detecting iodine-positive lesions [342] and provided further information in an overwhelming majority of thyroid cancer patients [343].

4.1.4. Breast Cancer

Among current clinical imaging modalities, MRI of the breast has the highest sensitivity for breast cancer detection and is becoming an indispensable tool for breast-imaging procedures [344,345]. Multi-parametric (Mp) MRI is the most sensitive imaging modality for breast cancer detection [346] and has been successfully used in combination with PET/MRI [347] and other imaging modalities. Abbreviated breast MRI uses only a select number of sequences and postcontrast imaging reduces the table time and reading time to maximize availability, patient tolerance, and accessibility [348], and may enable the more widespread use of breast-MRI for screening [349].

4.1.5. Lung Cancer

Among all cancer, lung cancer has the highest rate of mortality in the western world [350]. MRI is suitable for lung cancer screening with an excellent sensitivity and specificity of malignant as well as calcified and subsolid nodules [351–355] and for radiation treatment planning in lung cancer [356]. DWI-MRI protocol have been designed for imaging malignant lung tumors, achieving satisfactory within-patient repeatability [357], while recent advances in PET/MRI for lung cancer staging have been reviewed [358,359].

4.1.6. Gastric Cancer

Gastric cancer is an important healthcare problem from a global perspective, being the fifth most common malignancy and the third leading cause of cancer-related death [360]. Most of the cases are diagnosed at late stages when the treatment is largely ineffective, and MRI is of great value in patients with gastric cancer [361,362]. MRI is useful for distant metastasis assessment with particular reference to peritoneal and liver metastases assessment [361]. DWI-MRI and apparent diffusion coefficient (ADC) values showed to be useful in preoperative MRI staging of gastric cancer [363,364], but has a low accuracy to detect or to differentiate metastatic and non-metastatic lymph nodes based on their ADC values in gastric cancer [365]. MRI is useful for the diagnosis of serosal invasion of gastric cancer [366], in the diagnosis of regional lymph node metastases [367], and is the best method in the assessment of gastric wall infiltration in gastric cancer [368]. MRI is more accurate in achieving adequate staging results [369,370] and in the evaluation of

T-stage than CT [371,372]. DCE-MRI parameters of gastric cancers may provide prognostic information [373,374], whereas multiparametric fully integrated ¹⁸fluorodesoxyglucose ([F-18]-FDG)-PET/MRI may improve the diagnostic accuracy for translational gastric cancer research [375], for preoperative M staging as well as the resectability of gastric cancers compared to multi detector computed tomography (MDCT) [376]. Compared with PET/CT, PET/MRI performs more accurately in TNM staging and is optimal for accurate N staging [377], whereas high-resolution MRI (HR MRI) has good diagnostic performance and may serve as an alternative technique in the T staging of patients with esophagogastric junction cancer in clinical practice [378]. The preoperative prediction results of MRI are consistent with postoperative pathological results [379], although the clinical use of MRI for gastric cancer is still under discussion [380].

4.1.7. Pancreatic Cancer

Pancreatic cancer is a deadly disease, mainly because it is very resistant to chemotherapy and radiation therapy and is generally discovered very late [381]. MRI provides relevant information for the diagnostic evaluation of malignant pancreatic tumors [382,383] by predicting the survival in advanced pancreatic cancer patients [384]. The use of MRI of the liver for the initial staging of pancreatic cancer results in lower total costs and higher effectiveness [385]. MRI and CT show similar performance in the presurgical evaluation of pancreatic cancer [386,387]. Preoperative MRI is instrumental to detect the stage and resectability of pancreatic cancer [388] and preoperative gadoxetic acid-enhanced liver MRI has a high diagnostic performance in detecting liver metastasis from pancreatic ductal adenocarcinoma [389,390]. Gadolinium-enhanced MRI with DWI detected synchronous liver metastases [391], whereas [F-18]-FDG PET/MRI provides an imaging tool to improve the staging of pancreatic cancer and for the identification of Sister Mary Joseph nodules [392]. However, the addition of DWI to conventional MRI does not facilitate the differentiation of pancreatic cancer from chronic pancreatitis [393], whereas MRI can differentiate pancreatic carcinoma from chronic pancreatitis successfully when including Gd-enhanced T1-weighted 3D-GE sequences [394]. MRI-guided celiac plexus neurolysis is an effective and minimally invasive procedure for the palliative pain management of pancreatic cancer [395], whereas dynamic susceptibility contrast MRI (DSC-MRI) may predict early progression in patients with advanced pancreatic cancer that are undergoing chemotherapy [396]. MRI has been used to monitor radiofrequency heat (RFH)-enhanced chemotherapy in pancreatic cancers for the efficient treatment of pancreatic malignancies using MRI/RFH-integrated local chemotherapy [397].

4.1.8. Hepatocellular Carcinoma

Primary liver cancer is the second most common cause of cancer mortality worldwide and the sixth most common cancer overall [398]. MRI is superior to CT in sensitivity, specificity, and accuracy [399] and can be used to determine the differential diagnosis [400–402], variant analysis [403], arterial phase hyperenhancement [404], small precursor and recurrent lesions [405,406], liver perfusion [407], histological grade, microvascular invasion status, local and systemic therapeutic responses, prognosis [408,409], and as a preoperative marker [410] in hepatocellular carcinoma patients. PET/MRI imaging is also used for the diagnosis of patients with hepatocellular carcinoma [411–413], whereas multi-phasic MRI staging was found to be more accurate than the straight hepatocellular carcinoma-grading approach [414]. Texture analysis that was based on gadolinium-ethoxybenzyl-diethylenetriamine penta-acetic acid (GdEOB-DTPA)-enhanced MRI is used for early prediction of therapeutic outcome in intermediate hepatocellular carcinoma [415], identification of vessels encapsulating tumor clusters-positive hepatocellular carcinoma [416], has a higher diagnostic rate and a better diagnostic value in small hepatocellular carcinoma [417], and for the detection of capsule appearance in patients with hepatocellular carcinoma [404] and liver cirrhosis [418,419]. DCE-MRI is used in the prediction of staging B or C hepatocellular carcinoma [415] and for the quantification of perfusion metrics [420] with a superior modal-

ity for diagnosis compared with dynamic contrast-enhanced CT-scan [421]. DWI-MRI is the gold standard in detecting recurrent lesions [405], monitoring response to therapy, predicting response, assessing prognosis, and distinguishing tumor recurrence from the treatment effect [422]; however, DWI adds little value to MRI in target delineation [423].

4.1.9. Gallbladder Carcinoma

The most common cancer of the biliary system is gallbladder carcinoma [424]. MRI is a useful imaging tool for the staging, diagnosis, and evaluation of the treatment response and provides superior soft-tissue characterization of the gallbladder and biliary tree [425,426]. DWI is the preferred imaging technique for discriminating benign from malignant disease in gallbladder cancer [427,428].

4.1.10. Renal Cancer

Renal cancer (that is neoplasia of the kidney, renal pelvis, or ureter (ICD-9 189 and ICD-10 C64-C66)) is the seventh most common malignancy in men [429]. There are three main risk factors for cancer of the kidney: age, smoking, and obesity [430]. Renal carcinoma is often first detected and characterized with imaging, with CT and MRI being the most common modalities that are used for diagnosis, staging, and surveillance [431–433]. MRI differentiates papillary renal cell carcinoma from other renal masses [434], whereas MRI and normalized ADC has utility in differentiating central renal cell carcinoma from renal pelvic urothelial carcinoma [435]. DCE MRI allows an estimation of the grading of renal cell carcinoma [436] and along with DWI MRI and multiphase contrast-enhanced MRI (MCE-MRI) contributes with prognostic information, even at baseline scans, by predicting the tumor response early after initiating therapy [437].

4.1.11. Bladder Cancer

Bladder cancer is the fourth most common cancer worldwide [438]. MRI is effective in bladder cancer staging as well as differentiating superficial from invasive tumors and organ-confined from non-organ-confined tumors [439–447]. MRI has shown potential for the detection of muscle invasion [448]. Mp-MRI has been a useful modality for the T staging of bladder cancer for clinical and research applications [449,450], whereas DCE-MRI provides response biomarkers in clinical trials in subjects with primary bladder cancer [451]. For bladder cancer patients, diagnostics that are based on the use of hybrid systems incorporating both MRI scanning capabilities with the linear accelerator offers a number of potential advantages [452], whereas in bladder cancer patients that are undergoing cystectomy, DWI is used in the detection of metastatic pelvic lymph nodes [453] and in the preoperative T staging of urinary bladder cancer [454].

4.1.12. Ovarian Cancer

Ovarian cancer is the most lethal gynecologic malignancy; it accounts for 2.5% of all malignancies among females but 5% of female cancer deaths because of low survival rates that are largely driven by late-stage diagnoses [455]. An MRI of ovarian cancer has been instrumental to differentiate metastatic ovarian tumors from primary epithelial ovarian cancers [456]. Functional MRI techniques such as tumor-selective molecular imaging (TSMI), DW-MRI, and DCE-MRI are under evaluation as possible predictive and prognostic biomarkers in the context of ovarian malignancy and in routine clinical practice [457–461]. Contrasting results are reported about the role of PET/MRI in ovarian cancer [462]. MRI was found to be more sensitive than PET/CT for detecting local pelvic recurrence and peritoneal lesions of recurrent ovarian tumors [463,464], although PET/CT had a higher specificity than pelvic MRI for diagnosis of malignant ovarian tumors [465–467]. A consensus process in the creation of a standardized lexicon for ovarian and adnexal lesions for MRI and the resultant lexicon has been recently published [468].

4.1.13. Cervical Uterine Cancer

Cervical uterine cancer is the leading cause of morbidity and mortality in women in developing countries and is known to be related to human papillomavirus [469]. Pelvic MRI is the reference examination for the evaluation of cervical cancers, allowing the accurate evaluation of tumor size, parametrial extension, and lymph node metastasis, which are essential factors for therapeutic management [470–473]. MRI, CT/MRI, and PET/MRI have been used for cervical cancer staging and lymph node metastasis [474,475], and PET/MRI was found to possess a higher diagnostic sensitivity [476], specificity [477], and accuracy [478,479], also during pregnancy [480], being helpful in clinical diagnosis [481], prediction [482], and treatment [483,484]. MRI diagnosis is an auxiliary method for cervical cancer treatment when used in combination with tumor markers (e.g., squamous cell carcinoma antigen) [485] and for the management of women with early cervical cancer [486]. DWI-MRI and ADC are used as a non-invasive imaging methods for characterizing the fraction of collagen I-positive tissue across different tumor models of uterine cervical cancer [487], for the pathological grade of tumor [488], for the differentiation between metastatic and non-metastatic pelvic lymph nodes [489], and between normal and cancerous tissue in the uterine cervix [490]. Another noninvasive technique that is used to assess tumor vascular oxygenation at 3 T in cervical cancer staging is blood oxygenation level-dependent contrast MRI [491].

4.1.14. Endometrial Cancer

Endometrial cancer is the fourth most common malignancy in women and the most common gynecological malignancy in the developed world after lung, colorectal, and breast cancer [492]. MRI is recommended for the initial staging and report of endometrial cancer [493–496] and preoperative pelvic MRI is a moderately sensitive and specific method of identifying invasion to the outer half of myometrium in endometrial cancer [497–499]. MRI has a high specificity and negative predictive value in endometrial cancer staging [500]; however, its accuracy in detecting myoinvasion is limited [501]. MRI with DWI and DCE sequences can help establish a correct diagnosis [502,503], while 3.0 T multimodal MRI is an important imaging tool for preoperative endometrial cancer staging [504]. MRI quantitative assessments such as tumor area ratio (TAR), tumor volume ratio (TVR), MRI-based whole-tumor radiomic signatures, and ADC were found to improve the accuracy of preoperative staging, helping in the risk stratification of endometrial cancer [505–507]. The combination of MRI and immunohistochemical examination is a powerful tool for preoperative risk stratification to assist in clinical decision-making for endometrial cancer patients [508]. [F-18]-FDG PET/MRI is a valid imaging technique in patients with endometrial cancer, both in staging and restaging as an alternative diagnostic strategy to conventional imaging modalities, also considering the limited radiation exposure [509–511], whereas integrated PET/MRI successfully assesses the lymph node metastasis and myometrial invasion in patients with endometrial cancer [512,513]. MRI-guided intensity-modulated radiation therapy has been used for locally recurrent endometrial cancer after resection [514].

4.1.15. Prostate Cancer

Prostate cancer is the most common cancer for males, and it is estimated that 15% men are predicted to develop prostate cancer over their lifetime [515]. The application of MRI has been successfully used for its sensitivity in detecting clinically significant cancer and the ability to localize the tumor within the prostate gland [516,517] by using Mp-MRI [518–522] and in the hybrid PET/MRI [523–525]. In addition to the fusion strategy, biopsies with MRI targets play an important role in the assessment of patients with a previous negative prostate biopsy [518].

4.1.16. Testicular Cancer

In men between 15 and 49 years-old, the most common nonhematologic malignancy is testicular cancer [526]. It has excellent cure rates; however, poor guideline adherence

can lead to inappropriate management with a detrimental effect on outcomes [527]. MRI is successful in the diagnosis of testicular germ cell cancer [528,529], particularly when directed towards the retroperitoneum and pelvis only [530]. Functional information that is based on DWI and DCE MRI data improve testicular mass lesion characterization [531,532] and can be used to characterize small, solid testicular tumors [533] and for the follow-up of testicular cancer patients [534], independently of the examiner [535]. Mp-MRI can potentially differentiate benign stromal tumors from malignant testicular neoplasms, which can help to avoid radical orchectomy [536,537]. MRI can be used as an alternative to CT to reduce radiation exposure [538,539] and is a valuable diagnostic aid in the preoperative localization of ectopic testes in cryptorchidism or if findings are equivocal [540].

4.1.17. Colorectal Cancer

Colorectal cancer is a common cancer and a common cause of death. There is evidence that an important proportion of colorectal cancer patients remain untreated [541]. Pelvic MRI is used for the local of T and N staging of rectal cancer and has the advantage of improved patient comfort [542], improved reproducibility and accuracy [543], reduced care costs [544], and for completeness and better understanding of related pelvic anatomy [545–547]. The use of DWI with ADC value in addition to conventional MRI yields better diagnostic accuracy than using conventional MRI alone in detection, correlation with tumor histologic grade, and the initial staging in patients with locally advanced colorectal cancer [548,549]; however, DW-MRI is inferior to [F-18]-FDG-PET for the detection of primary lesions but superior for the detection of lymph node metastases [550]. MRI shows moderate sensitivity and good specificity for the detection of extramural venous invasion (EMVI) in colorectal cancer [551,552], while 3D colorectal MRI gives better and accurate segmentation results than 3D fully convolutional neural networks alone [553]. For lymph node metastasis of colorectal cancer, the sensitivity and specificity of preoperative diagnosis by diffusion (D-MRI) is higher if the node is hyperintense and more than 9 mm in diameter [554], whereas high-b-value DWI-MRI has a high sensitivity and specificity to detect colorectal adenocarcinoma [555]. Recent developments and emerging technologies in CT and MRI are changing the management of colorectal cancer patients in many clinical scenarios [556]. MRI is more accurate than CT [557] and MDCT for the evaluation of liver metastases [558], whereas for patients with colorectal cancer, PET/MRI may aid in the selection of more appropriate treatment strategies [559]. Whole-body MRI (WB-MRI) is a radiation-free alternative to standard sequential algorithms of staging and follow-up of colorectal cancer [560].

4.2. Magnetic Fields and Cancer Treatment

Radiation therapy, chemotherapy, and immunotherapy, alone or in combination with therapies such as photothermal therapy, photodynamic therapy, hyperthermia, and radiotherapy have been proposed in the recent literature [561]. In the next section, the use of magnetic nanoparticles for the delivery of anticancer agents and magnetomechanical tools and in hyperthermia will be discussed.

4.2.1. Delivery of Anticancer Agents via Magnetic Carrier Particles

An exciting new prospect in treating cancer is the delivery of anticancer agents via magnetic carrier particles, which are used as a "carrier system" for a variety of anticancer agents [562–565]. For instance, by using an external MF, it is possible to guide magnetic iron oxide nanoparticles (MIONs) to their target. This is the principle behind the development of superparamagnetic iron oxide nanoparticles (SPIONs) as novel drug delivery vehicles [566]. Palmitoyl chitosan that is co-encapsulated with SPIONs and the anticancer drug paclitaxel via the nanoprecipitation process increased the amount of drug in cancer cells [567], whereas doxorubicin (Dox)-conjugated heparin was used with the SPION technology for targeted anticancer drug delivery [568]. In MCF-7 breast cancer cells, Dox was rapidly internalized and exhibited higher toxicity than treatments with Dox alone when it was

assembled in magnetic nanoparticle-supported lipid bilayers [569] and with Dox-loaded polymer@Au/Fe₃O₄ core/shell nanoparticles in simulated cancerous environments [570]. Bioactive molecules such as curcumin can be loaded in magnetic silk fibroin core-shell nanoparticles to enhance cytotoxicity and cellular uptake in the human Caucasian breast adenocarcinoma cell lines with superior biomedical applications due to their size range, which is particularly desired for cell internalization [571].

It has to be considered that since the magnetic force decreases rapidly with the distance from the magnets, the targeting of tumors that are situated at large distances from the surface of the human body might be difficult. Therefore, the delivery of anticancer agents via magnetic carrier particles appears to be more suitable for treating sub-surface cancers within the human body [572]. Nevertheless, several new magnetic nanoparticles have been designed and evaluated for cancer treatment, offering the ability to deliver drugs efficiently [573–575].

Despite the increasing body of evidence supporting promising results, there are some drawbacks that are related to magnetic nanoparticles (MNP) use in drug delivery, such as the difficulty in maintaining the therapeutic action in three dimensions inside the human body, the limitation to maintain efficacy in the target organ once the MF is removed from outside, and the limited effective incorporation of magnetic iron oxide nanoparticles into biomedical systems [576,577].

4.2.2. Magnetomechanical Methods for Cancer Therapy

Magnetomechanical therapy is one of the most prospective directions in tumor microsurgery based on a physical nanostructure that is able to transform the magnetic moment to mechanical torque and a ligand molecule that allows the scalpel to precisely target tumor cells [578]. Nano-magnetomechanical activation (NMMA) of the MNPs is used to localize and apply force to target biomolecular structures as transport vesicles, cell organelles, enzyme molecules, etc., without significant heating [579]. Nanospinners can exert mechanical forces under a rotating magnetic field at 15 Hz and 40 mT to target the mitochondria of cancer cells [580]. Iron nanowires that are functionalized with anti-CD44 antibodies have been used in a combination therapy that included magnetomechanical and photothermal treatments on colon cancer cells [581]. Hedgehog-like microspheres that were composed of needle-like MNP with carbon and gold double shells seriously damaged cancer cells and strongly inhibited tumor growth through mechanical force [582]. Magnetic disks are a new generation of MNP with outstanding properties to face biomedical challenges in cancer treatment microsurgery [578] and the investigation toward the most efficient magnetomechanical actuator to destroy cancer cells has been recently reviewed [583].

4.2.3. Magnetic Hyperthermia Ad Cancer Treatment

Magnetic hyperthermia treatment (MHT) utilizes heat that is generated by MNPs under an alternating MF to selectively kill tumor cells [584–586]. When exposed to an alternating magnetic field (AMF), MNPs can generate heat via hysteresis loss (large multi-domain MNPs) or through Neel- and Brownian relaxation losses (typically small, single-core MNPs) [587]. The efficiency of MHT depends on the size, concentration, solution viscosity, and composition of MNPs as well as the strength and frequency of the MF [588].

Several materials are used to prepare MNP for MHT. Ferrimagnetic glass-ceramic have been successfully used as thermo-seeds for a hyperthermic treatment of carcinoma cells in Sprague–Dawley rats [589], whereas magnetite cationic liposomes where used to generate hyperthermia on local tumors and lung metastases in a mouse model of osteosarcoma [590]. Spinel ferrite nanoparticles were successfully synthesized and used for MHT [591], whereas MIONs (such as crack-free ferrimagnetic maghemite, γ-Fe₂O₃) may be useful for the in situ hyperthermic treatment of cancer [592–595]. SPIONs have been increasingly studied for their excellent MHT applications [596,597], whereas lanthanum-strontium manganite particles that were embellished with gold nanoparticles were found to be suitable for the treatment of deeper tumors [598].

The limitations and advantages to more effective clinical use of MNP-based thermometry to achieve greater impact on clinical translation of MNH have been recently reported [599]. Despite the wide use of MHT in clinical application, the technology suffers from inadequate and uneven heating due to low and heterogeneous concentrations of MNPs within the target tumor [600,601]. It has been calculated that, to achieve sufficient hyperthermia in targeted tumors, a high concentration of MNP is required [602] and often the particle concentration is below that which is needed to induce sufficient heating of tissue, thus lowering the therapeutic effects of MHT [603].

The use of MNPs in “traditional” biomedical applications that are related to cancer theranostics, such as drug delivery, hyperthermia, MRI, micro nuclear magnetic resonance, and surface-enhanced Raman spectroscopic detection technology has been demonstrated [604–609], including the development of next-generation high-performance theranostic agents that are based on MNP assemblies [610]. Figure 6 summarizes the use of MFs in cancer theranostics.

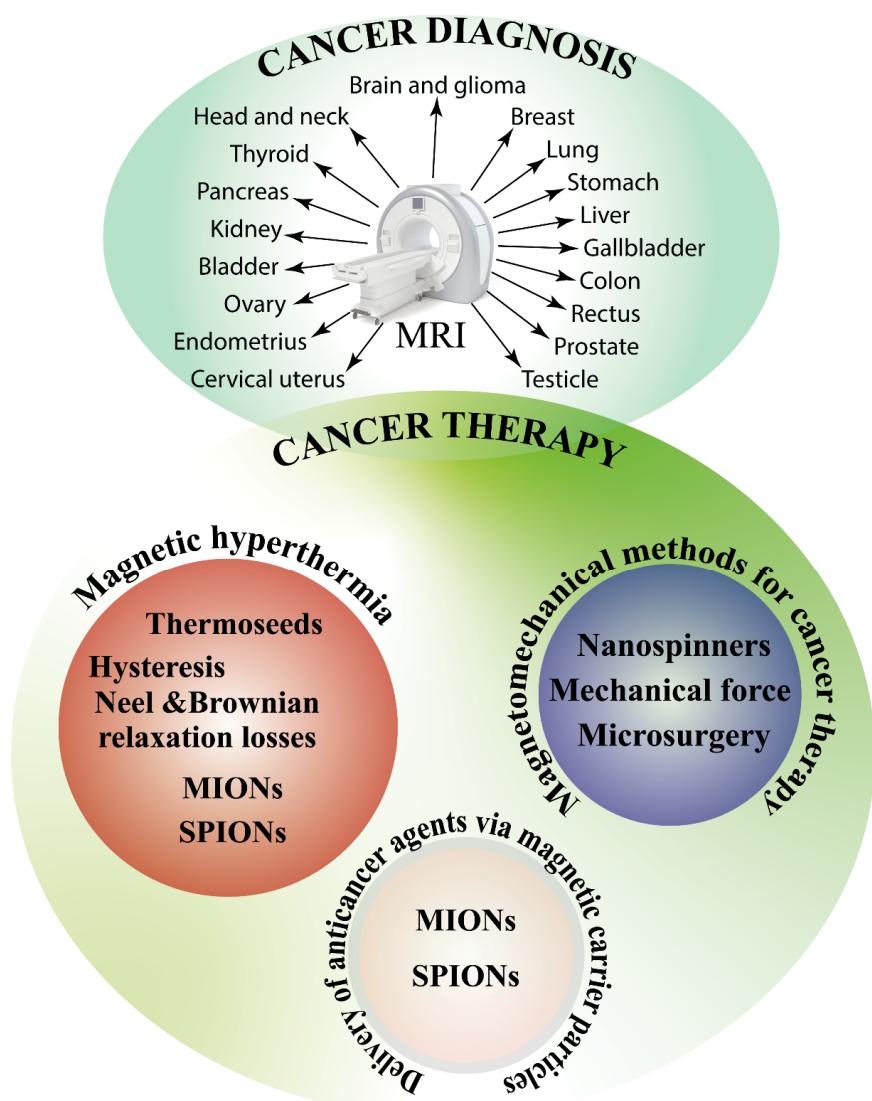


Figure 6. MFs and cancer theranostics. Theranostics combines the terms diagnostics and therapy. MF are used in MRI for the diagnosis of several cancers, whereas the use of magnetic nanoparticles for cancer therapy encompasses three major areas: magnetic hyperthermia that is aimed to kill cancer cells with heat; drug delivery by the use of SPIONs and MIONs; and the exploitation of MNP mechanical forces for application in nano and microsurgery.

5. Conclusions

The potential health effects of man-made EMF have been a topic of scientific interest since the late 1800s and have received particular attention during the last 40 years. Since the first studies suggesting a relationship between MF and childhood cancer [29], the scientific community has evaluated the possible mechanisms for the effects of MFs on biological systems. Epidemiological studies are often controversial and sometimes misleading. Nevertheless, there is a consensus on the positive relationship between residential/domestic exposure to ELF EMF and the occurrence of brain cancer, whereas contrasting results require more experimentation to assess the influence of occupational exposure to MFs on brain cancer. The epidemiology of leukemia as related to ELF EMF in adults is controversial in both residential/domestic and occupational exposure. For children, leukemia is not associated to occupational exposure, whereas a growing body of evidence indicates a correlation between residential/domestic exposure to ELF EMF and childhood leukemia. Breast cancer has been related to ELF EMF exposure more in residential/domestic epidemiological studies than in occupational, but the melatonin hypothesis, although recently revisited, finds little consensus. When studied at the cellular and in vitro level, MFs exert their effect on both human and animal (rat and mice, mainly) cells when used at a high intensity and for a long time. The common response is the production of ROS, which trigger a cascade of other cellular responses which might be the direct consequence of MF exposure. The use of MF is instrumental for the diagnosis and therapy (theranostic) of cancer. MRI is instrumental for the precise diagnosis of different cancers, whereas MNPs open the new era of nanomedicine, allowing (i) the smart delivery of anticancer drugs, (ii) nanosurgery through their magnetomechanic properties, and (iii) fighting the cancer cells *in situ* by exploiting their capability to generate heat (hyperthermia) via hysteresis loss or through Neel- and Brownian relaxation losses.

Figure 7 summarizes the effects of MFs on cancer that were discussed in this review.

Although humans do not perceive the presence or changes of MFs, variations in MF intensity and inclination exert biological effects, with the greatest effects observed at the cellular and subcellular level. The basic response to MF relies on ROS-production with RPM playing a potential role in magneto-perception. Scientists do not generally agree that there is a cause-effect relationship between exposure to MF and cancer, also because of the difficulty in obtaining reproducible effects that are consistent with the hypothesis that MF may cause or promote cancer. MFs might not be the direct cause of cancer but may contribute to ROS-production and generate oxidative stress through RPM [611], which could trigger or enhance the expression of oncogenes [612]. Large-scale epidemiological studies are needed to help resolve these issues along with in depth studies on the relationship between magnetoreception, ROS-generation, and cancer.

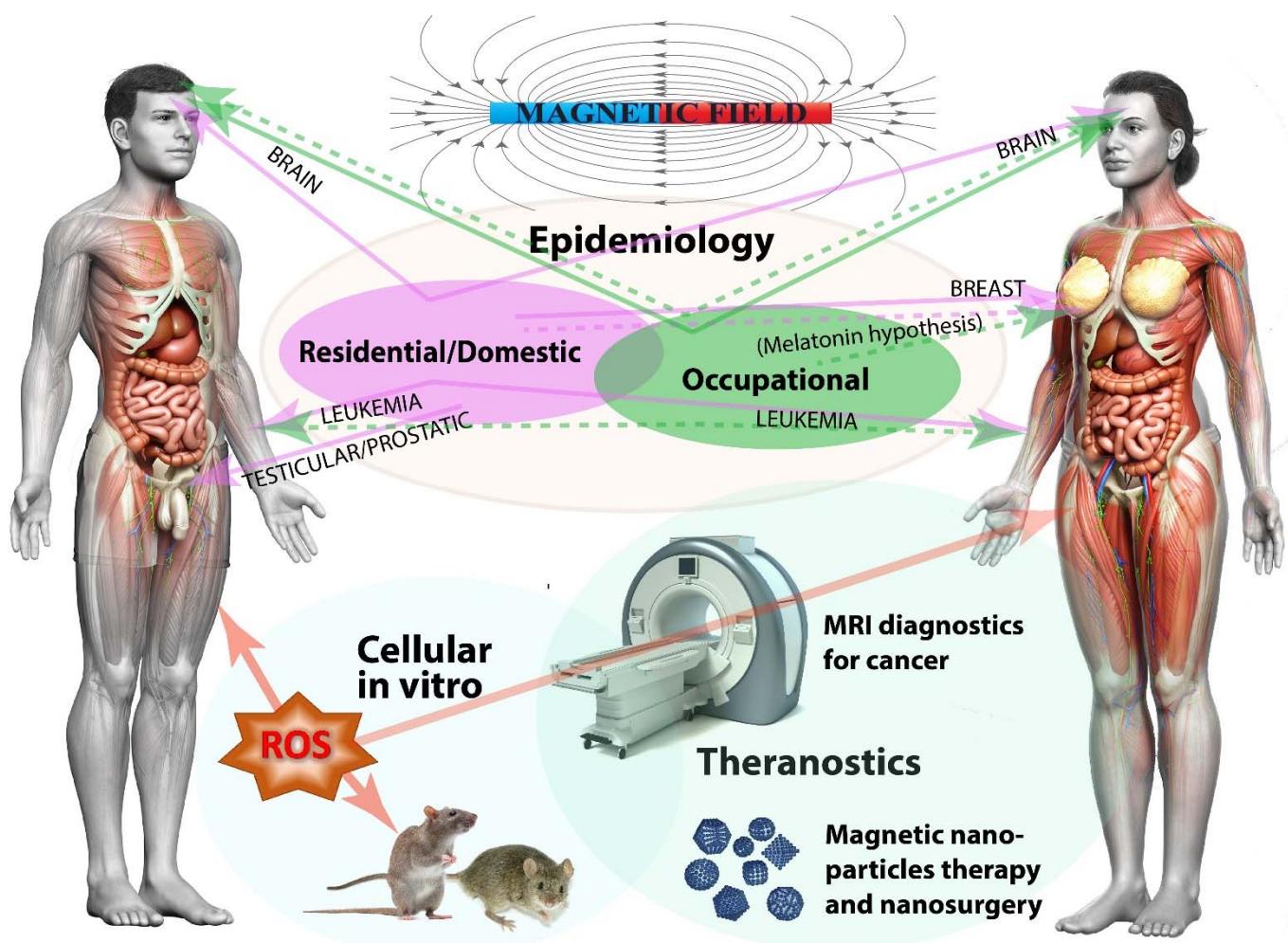


Figure 7. The effects of MFs on cancer. Humans are exposed to a complex mix of man-made electric and magnetic fields at many different frequencies both at home and at work. Epidemiological studies indicate that there is a positive relationship (solid lines) between residential/domestic and occupational exposure to ELF EMF and brain cancer, although some other studies indicate that there is no relationship (dotted lines). Breast cancer appears to be more related to residential/domestic exposure than occupational and in both epidemiological surveys, the so-called “melatonin hypothesis” finds weak evidence. Testicular/prostatic cancer is associated with residential/domestic exposure, as is leukemia, which is mostly associated (particularly in children) with the close proximity to ELF EMF. The cellular and in vitro studies on both animal (mainly rat and mice) and human cells indicate the role of ROS-generation as a consequence of exposure to different MF intensity and timing, suggesting also a magnetoreception mechanism that is based on RPs. Finally, MFs can be used for theranostic applications; MRI is instrumental for the diagnosis of several cancers, whereas the use of MNP allows the treatment of cancer by nanomedical applications for the precise delivery of anticancer drugs, nanosurgery by magnetomechanic methods, and the selective killing of cancer cells by magnetic hyperthermia.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms23031339/s1>.

Funding: This work was supported by the University of Turin local research funds to M.E.M.

Data Availability Statement: Supporting Data File S1 contains the EndNote Library used for this review.

Conflicts of Interest: The author declares no conflict of interest.

Abbreviations

[F-18]-FDG	¹⁸ Fluor Desoxy Glucose
ADC	Apparent Diffusion Coefficient
AF	Alternating Field
AL	Acute Leukemia
ALL	Acute Lymphoblastic/lymphocytic Leukemia
AM	Amplitude Modulation
AMF	Alternating Magnetic Field
AMI	Acute Myocardial Infarction
AML	Acute Myeloid Leukemia
B-ALL	B-Lineage Acute Lymphoblastic Leukemia
C-11-MET	¹¹ C-methyl-L-Methionine
CAD	Computer-Aided Diagnosis
CAT	Catalase
CCHD	Chronic Coronary Heart Disease
CLL	Chronic Lymphocytic Leukemia
CML	Chronic Myeloid Leukemia
CT	Computed Tomography
CYP-450	Cytochrome P450
DAB	Digital Audio Broadcasting
DCE	Dynamic Contrast-Enhanced
DGE	Dynamic Glucose-Enhanced
D-MRI	Diffusion Magnetic Resonance Imaging
Dox	Doxorubicin
DSC-MRI	Dynamic Susceptibility Contrast Magnetic Resonance Imaging
DVBT	Digital Video Broadcasting Terrestrial
DWI	Diffusion-Weighted Imaging
EHS	Electromagnetic Hypersensitivity
ELF	Extremely Low Frequency
EM	Electromagnetic
EMF	Electromagnetic Field
EMVI	Extramural Venous Invasion
ERK1/2	Extracellular Signal-Regulated Kinases 1/2
FM	Frequency Modulation
FMT	Fluorescence Molecular Tomography
GdEOB-DTPA	Gadolinium-Ethoxybenzyl-Diethylenetriamine Pentaacetic Acid
GfM	Gadofluorine-M
GIS	Geographical Information System
GMF	Geomagnetic Field
GPX	Glutathione Peroxidase
HR MRI	High-Resolution Magnetic Resonance Imaging
HVDC	High Voltage Direct Current
HVOTL	High-Voltage Overhead Transmission Line
IARC	International Agency for Research on Cancer
ICNIRP	International Commission on Non-Ionizing Radiation Protection
IDH	Isocitrate Dehydrogenase
IRPA	International Radiation Protection Association
LF	Low-Frequency
LF EMF	Low Frequency Electromagnetic Field
LL	Lymphatic Leukemia
MCE-MRI	Multiphase Contrast-Enhanced Magnetic Resonance Imaging
MDA	Malondialdehyde
MDCT	Multi Detector Computed Tomography
MF	Magnetic Field
MHT	Magnetic Hyperthermia Treatment
MION	Magnetic Iron Oxide Nanoparticles
MNP	Magnetic NanoParticle

Mp-MRI	Multi-Parametric Magnetic Resonance Imaging
MRI	Magnetic Resonance Imaging
Na-23-MRI	Na-23-Magnetic Resonance Imaging
NI	Non Ionizing
NI EMF	Non-Ionizing Electromagnetic Field
NMMA	Nano-Magnetomechanical Activation
NMR	Nuclear Magnetic Resonance
NPP	Nuclear Power Plants
OR	Odd Ratio
PET	Positron Emission Tomography
PFF	Power-Frequency Field
PKC	Protein Kinases C
RF	Radio Frequency
RFH	Radiofrequency Heat
RFID	Radio Frequency Range
RNS	Reactive Nitrogen Species
ROS	Reactive Oxygen Species
RP	Radical Pair
RPM	Radical Pair Mechanism
S	Singlet State
SAR	Specific Absorption Rate
SF	Static Field
SMF	Static Magnetic Field
SOD	Superoxide Dismutase
SPIO	Superparamagnetic Iron Oxide
SPION	Superparamagnetic Iron Oxide Nanoparticle
T	Triplet State
TAR	Tumor Area Ratio
TSMI	Tumor-Selective Molecular Imaging
TVR	Tumor Volume Ratio
ULF	Ultra-Low Frequency
USPIO	Ultrasmall Superparamagnetic Particles of Iron Oxide
WB-MRI	Whole Body Magnetic Resonance Imaging
WHO	World Health Organization
WLAN	Wireless Local Area Networks

References

- Monadizadeh, S.; Kibert, C.J.; Li, J.X.; Woo, J.; Asutosh, A.; Roostaie, S.; Kouhirostami, M. A review of protocols and guidelines addressing the exposure of occupants to electromagnetic field radiation (EMFr) in buildings. *J. Green Build.* **2021**, *16*, 55–81. [[CrossRef](#)]
- Rathebe, P.C.; Modisane, D.S.; Rampedi, M.B.; Biddesay-Manilal, S.; Mbonane, T.P. A review on residential exposure to electromagnetic fields from overhead power lines: Electrification as a health burden in rural communities. In Proceedings of the 2019 Open Innovations (OI), Cape Town, South Africa, 2–4 October 2019; pp. 219–221.
- Gupta, S.; Sharma, R.S.; Singh, R. Non-ionizing radiation as possible carcinogen. *Int. J. Environ. Health Res.* **2020**, *1*–25. [[CrossRef](#)] [[PubMed](#)]
- Mazzanti, G. Evaluation of continuous exposure to magnetic field from ac overhead transmission lines via historical load databases: Common procedures and innovative heuristic formulas. *IEEE Trans. Power Deliv.* **2010**, *25*, 238–247. [[CrossRef](#)]
- Landler, L.; Keays, D.A. Cryptochrome: The magnetosensor with a sinister side? *PLoS Biol.* **2018**, *16*, e3000018. [[CrossRef](#)] [[PubMed](#)]
- Jacobson, J.I. Speculations on the influence of electromagnetism on genomic and associated structures. *J. Int. Med. Res.* **1996**, *24*, 1–11. [[CrossRef](#)]
- Repacholi, M. Concern that “EMF” magnetic fields from power lines cause cancer. *Sci. Total Environ.* **2012**, *426*, 454–458. [[CrossRef](#)] [[PubMed](#)]
- Saliev, T.; Begimbetova, D.; Masoud, A.R.; Matkarimov, B. Biological effects of non-ionizing electromagnetic fields: Two sides of a coin. *Prog. Biophys. Mol. Biol.* **2019**, *141*, 25–36. [[CrossRef](#)] [[PubMed](#)]
- Lednev, V.V. Possible mechanism for the influence of weak magnetic fields on biological systems. *Bioelectromagnetics* **1991**, *12*, 71–75. [[CrossRef](#)] [[PubMed](#)]

10. Occhipinti, A.; De Santis, A.; Maffei, M.E. Magnetoreception: An unavoidable step for plant evolution? *Trends Plant Sci.* **2014**, *19*, 1–4. [[CrossRef](#)] [[PubMed](#)]
11. Maffei, M.E. Magnetic field effects on plant growth, development, and evolution. *Front. Plant Sci.* **2014**, *5*, 445. [[CrossRef](#)]
12. Swanson, J.; Kheifets, L. Could the geomagnetic field be an effect modifier for studies of power-frequency magnetic fields and childhood leukaemia? *J. Radiol. Prot.* **2012**, *32*, 413–418. [[CrossRef](#)] [[PubMed](#)]
13. Amoon, A.T.; Swanson, J.; Vergara, X.; Kheifets, L. Relationship between distance to overhead power lines and calculated fields in two studies. *J. Radiol. Prot.* **2020**, *40*, 431–443. [[CrossRef](#)] [[PubMed](#)]
14. Wyszkowska, J.; Szczygiel, M.; Trawinski, T. Static magnetic field and extremely low-frequency magnetic field in hybrid and electric vehicles. *Prz. Elektrotech.* **2020**, *96*, 60–62. [[CrossRef](#)]
15. Vaitl, D.; Propson, N.; Stark, R.; Schienle, A. Natural very-low-frequency sferics and headache. *Int. J. Biometeorol.* **2001**, *45*, 115–123. [[CrossRef](#)] [[PubMed](#)]
16. Feychtig, M.; Ahlbom, A.; Kheifets, L. EMF and health. *Annu. Rev. Public Health* **2005**, *26*, 165–189. [[CrossRef](#)] [[PubMed](#)]
17. Repacholi, M.H. Who's international EMF project. *Radiat. Prot. Dosim.* **1999**, *83*, 1–4. [[CrossRef](#)]
18. Bowman, J.D.; Ray, T.K.; Park, R.M. Possible health benefits from reducing occupational magnetic fields. *Am. J. Ind. Med.* **2013**, *56*, 791–805. [[CrossRef](#)] [[PubMed](#)]
19. Belyaev, I.; Dean, A.; Eger, H.; Hubmann, G.; Jandrisovits, R.; Kern, M.; Kundi, M.; Moshammer, H.; Lercher, P.; Muller, K.; et al. Europaem EMF guideline 2016 for the prevention, diagnosis and treatment of EMF-related health problems and illnesses. *Rev. Environ. Health* **2016**, *31*, 363–397. [[CrossRef](#)]
20. Repacholi, M.H. A history of the international commission on non-ionizing radiation protection. *Health Phys.* **2017**, *113*, 282–300. [[CrossRef](#)] [[PubMed](#)]
21. Soto Sumuano, J.L.; Abundis Gutierrez, E.; Tlacuilo-Parra, J.A.; Garibaldi Covarrubias, R.F.; Romo Rubio, H. Electromagnetic radiation, childhood leukemia and regulation. *Rev. Int. Contam. Ambient.* **2020**, *36*, 229–240.
22. Knave, B. Electromagnetic fields and health outcomes. *Ann. Acad. Med. Singap.* **2001**, *30*, 489–493. [[PubMed](#)]
23. Kelfkens, G.; Prupperts, M. Magnetic fields and childhood leukemia; science and policy in the Netherlands. In Proceedings of the Conference of the European Medical and Biological Engineering Conference (EMBEC) and the Nordic-Baltic Conference on Biomedical Engineering and Medical Physics (NBC), Tampere, Finland, 11–15 June 2017; Eskola, H., Vaisanen, O., Viik, J., Hyttinen, J., Eds.; Springer: Singapore, 2018; Volume 65, pp. 498–501.
24. Teepen, J.C.; van Dijck, J. Impact of high electromagnetic field levels on childhood leukemia incidence. *Int. J. Cancer* **2012**, *131*, 769–778. [[CrossRef](#)] [[PubMed](#)]
25. Calvente, I.; Fernandez, M.F.; Villalba, J.; Olea, N.; Nunez, M.I. Exposure to electromagnetic fields (non-ionizing radiation) and its relationship with childhood leukemia: A systematic review. *Sci. Total Environ.* **2010**, *408*, 3062–3069. [[CrossRef](#)] [[PubMed](#)]
26. Calvente, I.; Davila-Arias, C.; Ocon-Hernandez, O.; Perez-Lobato, R.; Ramos, R.; Artacho-Cordon, F.; Olea, N.; Nunez, M.I.; Fernandez, M.F. Characterization of indoor extremely low frequency and low frequency electromagnetic fields in the inmigranada cohort. *PLoS ONE* **2014**, *9*, e106666. [[CrossRef](#)]
27. Miah, T.; Kamat, D. Current understanding of the health effects of electromagnetic fields. *Pediatric Ann.* **2017**, *46*, E172–E174. [[CrossRef](#)] [[PubMed](#)]
28. Bates, M.N. Extremely low-frequency electromagnetic-fields and cancer—The epidemiologic evidence. *Environ. Health Perspect.* **1991**, *95*, 147–156. [[CrossRef](#)]
29. Wertheimer, N.; Leeper, E. Electrical wiring configurations and childhood cancer. *Am. J. Epidemiol.* **1979**, *109*, 273–284. [[CrossRef](#)] [[PubMed](#)]
30. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Non-ionizing radiation, part 2: Radiofrequency electromagnetic fields. *IARC Monogr. Eval. Carcinog. Risks Hum.* **2013**, *102 Pt 2*, 1–460.
31. Deutsch, S.; Wilkening, G.M. Electromagnetic field cancer scares. *Health Phys.* **1997**, *73*, 301–309. [[CrossRef](#)] [[PubMed](#)]
32. Karimi, A.; Moghaddam, F.G.; Valipour, M. Insights in the biology of extremely low-frequency magnetic fields exposure on human health. *Mol. Biol. Rep.* **2020**, *47*, 5621–5633. [[CrossRef](#)] [[PubMed](#)]
33. Moulder, J.E.; Foster, K.R. Biological effects of power-frequency fields as they relate to carcinogenesis. *Proc. Soc. Exp. Biol. Med.* **1995**, *209*, 309–324. [[CrossRef](#)] [[PubMed](#)]
34. Dieudonne, M. Electromagnetic hypersensitivity: A critical review of explanatory hypotheses. *Environ. Health* **2020**, *19*, 48. [[CrossRef](#)] [[PubMed](#)]
35. Elliott, P.; Shaddick, G.; Douglass, M.; de Hoogh, K.; Briggs, D.J.; Toledano, M.B. Adult cancers near high-voltage overhead power lines. *Epidemiology* **2013**, *24*, 184–190. [[CrossRef](#)]
36. Lambrozo, J. Electric and magnetic fields with a frequency of 50–60 hz: Assessment of 20 years of research. *Indoor Built Environ.* **2001**, *10*, 299–305.
37. Souques, M.; Lambrozo, J. 50–60 hz magnetic fields and health: What's new? *Radioprotection* **2015**, *50*, 95–99. [[CrossRef](#)]
38. Sheppard, A.R.; Kavet, R.; Renew, D.C. Exposure guidelines for low-frequency electric and magnetic fields: Report from the brussels workshop. *Health Phys.* **2002**, *83*, 324–332. [[CrossRef](#)] [[PubMed](#)]
39. Maes, A.; Verschaeve, L. Genetic damage in humans exposed to extremely low-frequency electromagnetic fields. *Arch. Toxicol.* **2016**, *90*, 2337–2348. [[CrossRef](#)] [[PubMed](#)]

40. Lacy-Hulbert, A.; Metcalfe, J.C.; Hesketh, R. Biological responses to electromagnetic fields. *FASEB J.* **1998**, *12*, 395–420. [CrossRef] [PubMed]
41. Kavet, R. EMF and current cancer concepts. *Bioelectromagnetics* **1996**, *17*, 339–357. [CrossRef]
42. Rathebe, P.; Weyers, C.; Raphela, F. Exposure levels of elc magnetic fields in the residential areas of mangaung metropolitan municipality. *Environ. Monit. Assess.* **2018**, *190*, 544. [CrossRef] [PubMed]
43. Navarro-Camba, E.A.; Segura-Garcia, J.; Gomez-Perretta, C. Exposure to 50 hz magnetic fields in homes and areas surrounding urban transformer stations in silla (spain): Environmental impact assessment. *Sustainability* **2018**, *10*, 2641. [CrossRef]
44. Sadeghi, T.; Ahmadi, A.; Javadian, M.; Gholamian, S.A.; Delavar, M.A.; Esmailzadeh, S.; Ahmadi, B.; Hadighi, M.S.H. Preterm birth among women living within 600 meters of high voltage overhead power lines: A case-control study. *Rom. J. Intern. Med.* **2017**, *55*, 145–150. [CrossRef] [PubMed]
45. Ahlbom, A.; Cardis, E.; Green, A.; Linet, M.; Savitz, D.; Swerdlow, A.; Epidemiology, I.S.C. Review of the epidemiologic literature on EMF and health. *Environ. Health Perspect.* **2001**, *109*, 911–933. [PubMed]
46. Alonso, A.; Bahillo, A.; de la Rosa, R.; Carrera, A.; Duran, R.J.; Fernandez, P. Measurement procedure to assess exposure to extremely low-frequency fields: A primary school case study. *Radiat. Prot. Dosim.* **2012**, *151*, 426–436. [CrossRef] [PubMed]
47. Li, C.Y.; Sung, F.C.; Chen, F.L.; Lee, P.C.; Silva, M.; Mezei, G. Extremely-low-frequency magnetic field exposure of children at schools near high voltage transmission lines. *Sci. Total Environ.* **2007**, *376*, 151–159. [CrossRef] [PubMed]
48. Tognola, G.; Chiaramello, E.; Bonato, M.; Magne, I.; Souques, M.; Fiocchi, S.; Parazzini, M.; Ravazzani, P. Cluster analysis of residential personal exposure to elc magnetic field in children: Effect of environmental variables. *Int. J. Environ. Res. Public Health* **2019**, *16*, 4363. [CrossRef] [PubMed]
49. Huss, A.; Goris, K.; Vermeulen, R.; Kromhout, H. Does apartment's distance to an in-built transformer room predict magnetic field exposure levels? *J. Expo. Sci. Environ. Epidemiol.* **2013**, *23*, 554–558. [CrossRef] [PubMed]
50. Roosli, M.; Jenni, D.; Kheifets, L.; Mezei, G. Extremely low frequency magnetic field measurements in buildings with transformer stations in switzerland. *Sci. Total Environ.* **2011**, *409*, 3364–3369. [CrossRef] [PubMed]
51. Ilonen, K.; Markkanen, A.; Mezei, G.; Juutilainen, J. Indoor transformer stations as predictors of residential elc magnetic field exposure. *Bioelectromagnetics* **2008**, *29*, 213–218. [CrossRef]
52. Vergara, X.P.; Kavet, R.; Crespi, C.M.; Hooper, C.; Silva, J.M.; Kheifets, L. Estimating magnetic fields of homes near transmission lines in the california power line study. *Environ. Res.* **2015**, *140*, 514–523. [CrossRef]
53. Aldrich, T.E.; Andrews, K.W.; Liboff, A.R. Brain cancer risk and electromagnetic fields (EMFs): Assessing the geomagnetic component. *Arch. Environ. Health* **2001**, *56*, 314–319. [CrossRef]
54. Nikkila, A.; Kendall, G.; Raitanen, J.; Spycher, B.; Lohi, O.; Auvinen, A. Effects of incomplete residential histories on studies of environmental exposure with application to childhood leukaemia and background radiation. *Environ. Res.* **2018**, *166*, 466–472. [CrossRef] [PubMed]
55. Greenland, S.; Kheifets, L.; Zafanella, L.E.; Kalton, G.W. Leukemia attributable to residential magnetic fields: Results from analyses allowing for study biases. *Risk Anal.* **2006**, *26*, 471–482. [CrossRef]
56. Miller, K.D.; Ostrom, Q.T.; Kruchko, C.; Patil, N.; Tihan, T.; Cioffi, G.; Fuchs, H.E.; Waite, K.A.; Jemal, A.; Siegel, R.L.; et al. Brain and other central nervous system tumor statistics, 2021. *CA Cancer J. Clin.* **2021**, *71*, 381–406. [CrossRef]
57. Farmanfarma, K.K.; Mohammadian, M.; Shahabinia, Z.; Hassanipour, S.; Salehiniya, H. Brain cancer in the world: An epidemiological review. *World Cancer Res. J.* **2019**, *6*, 5.
58. Pourreza, R.; Zhuge, Y.; Ning, H.; Miller, R. Brain tumor segmentation in MRI scans using deeply-supervised neural networks. In Proceedings of the 3rd International Workshop on Brain-Lesion (BrainLes) Held Jointly at the Conference on Medical Image Computing for Computer Assisted Intervention (MICCAI), Quebec City, QC, Canada, 14 September 2017; Springer International Publishing Ag: Quebec City, QC, Canada, 2018; pp. 320–331.
59. Kheifets, L.I. Electric and magnetic field exposure and brain cancer: A review. *Bioelectromagnetics* **2001**, *22*, S120–S131. [CrossRef]
60. Schoenfeld, E.R.; Henderson, K.; O'Leary, E.; Grimson, R.; Kaune, W.; Leske, M.C. Magnetic field exposure assessment: A comparison of various methods. *Bioelectromagnetics* **1999**, *20*, 487–496. [CrossRef]
61. Eskelinen, T.; Keinänen, J.; Salonen, H.; Juutilainen, J. Use of spot measurements for assessing residential elc magnetic field exposure: A validity study. *Bioelectromagnetics* **2002**, *23*, 173–176. [CrossRef] [PubMed]
62. Ahmadi, H.; Mohseni, S.; Akmal, A.A.S. Electromagnetic fields near transmission lines—Problems and solutions. *Iran. J. Environ. Health Sci. Eng.* **2010**, *7*, 181–188.
63. Kheifets, L.; Crespi, C.M.; Hooper, C.; Oksuzyan, S.; Cockburn, M.; Ly, T.; Mezei, G. Epidemiologic study of residential proximity to transmission lines and childhood cancer in california: Description of design, epidemiologic methods and study population. *J. Expo. Sci. Environ. Epidemiol.* **2015**, *25*, 45–52. [CrossRef] [PubMed]
64. Blaasaas, K.G.; Tynes, T. Comparison of three different ways of measuring distances between residences and high voltage power lines. *Bioelectromagnetics* **2002**, *23*, 288–291. [CrossRef] [PubMed]
65. Carles, C.; Esquirol, Y.; Turuban, M.; Piel, C.; Migault, L.; Pouchieu, C.; Bouvier, G.; Fabbro-Peray, P.; Lebailly, P.; Baldi, I. Residential proximity to power lines and risk of brain tumor in the general population. *Environ. Res.* **2020**, *185*, 109473. [CrossRef]
66. Marcilio, I.; Gouveia, N.; Pereira Filho, M.L.; Kheifets, L. Adult mortality from leukemia, brain cancer, amyotrophic lateral sclerosis and magnetic fields from power lines: A case-control study in Brazil. *Rev. Bras. Epidemiol.* **2011**, *14*, 580–588. [CrossRef] [PubMed]

67. Baldi, I.; Coureau, G.; Jaffré, A.; Gruber, A.; Ducamp, S.; Provost, D.; Lebailly, P.; Vital, A.; Loiseau, H.; Salamon, R. Occupational and residential exposure to electromagnetic fields and risk of brain tumors in adults: A case–control study in Gironde, France. *Int. J. Cancer* **2011**, *129*, 1477–1484. [CrossRef] [PubMed]
68. Li, C.Y.; Lin, R.S.; Sung, F.C. Elevated residential exposure to power frequency magnetic field associated with greater average age at diagnosis for patients with brain tumors. *Bioelectromagnetics* **2003**, *24*, 218–221. [CrossRef] [PubMed]
69. Carpenter, D.O. Extremely low frequency electromagnetic fields and cancer: How source of funding affects results. *Environ. Res.* **2019**, *178*, 108688. [CrossRef] [PubMed]
70. Mezei, G.; Gadallah, M.; Kheifets, L. Residential magnetic field exposure and childhood brain cancer—A meta-analysis. *Epidemiology* **2008**, *19*, 424–430. [CrossRef] [PubMed]
71. Kheifets, L.; Ahlbom, A.; Crespi, C.M.; Feychtig, M.; Johansen, C.; Monroe, J.; Murphy, M.F.G.; Oksuzyan, S.; Preston-Martin, S.; Roman, E.; et al. A pooled analysis of extremely low-frequency magnetic fields and childhood brain tumors. *Am. J. Epidemiol.* **2010**, *172*, 752–761. [CrossRef] [PubMed]
72. Saito, T.; Nitta, H.; Kubo, O.; Yamamoto, S.; Yamaguchi, N.; Akiba, S.; Honda, Y.; Hagihara, J.; Isaka, K.; Ojima, T.; et al. Power-frequency magnetic fields and childhood brain tumors: A case-control study in Japan. *J. Epidemiol.* **2010**, *20*, 54–61. [CrossRef] [PubMed]
73. Kroll, M.E.; Swanson, J.; Vincent, T.J.; Draper, G.J. Childhood cancer and magnetic fields from high-voltage power lines in england and wales: A case-control study. *Br. J. Cancer* **2010**, *103*, 1122–1127. [CrossRef]
74. Seomun, G.; Lee, J.; Park, J. Exposure to extremely low-frequency magnetic fields and childhood cancer: A systematic review and meta-analysis. *PLoS ONE* **2021**, *16*, e0251628. [CrossRef]
75. Ozen, S. Low-frequency transient electric and magnetic fields coupling to child body. *Radiat. Prot. Dosim.* **2008**, *128*, 62–67. [CrossRef]
76. Miller, K.D.; Nogueira, L.; Mariotto, A.B.; Rowland, J.H.; Yabroff, K.R.; Alfano, C.M.; Jemal, A.; Kramer, J.L.; Siegel, R.L. Cancer treatment and survivorship statistics, 2019. *CA Cancer J. Clin.* **2019**, *69*, 363–385. [CrossRef]
77. Davis, S.; Mirick, D.K.; Chen, C.; Stanczyk, F.Z. Effects of 60-hz magnetic field exposure on nocturnal 6-sulfatoxymelatonin, estrogens, luteinizing hormone, and follicle-stimulating hormone in healthy reproductive-age women: Results of a crossover trial. *Ann. Epidemiol.* **2006**, *16*, 622–631. [CrossRef] [PubMed]
78. Graham, C.; Cook, M.R.; Gerkovich, M.M.; Sastre, A. Melatonin and 6-ohms in high-intensity magnetic fields. *J. Pineal Res.* **2001**, *31*, 85–88. [CrossRef] [PubMed]
79. Youngstedt, S.D.; Kripke, D.F.; Elliott, J.A.; Assmus, J.D. No association of 6-sulfatoxymelatonin with in-bed 60-hz magnetic field exposure or illumination level among older adults. *Environ. Res.* **2002**, *89*, 201–209. [CrossRef]
80. Levallois, P.; Dumont, M.; Touitou, Y.; Gingras, S.; Masse, B.; Gauvin, D.; Kroger, E.; Bourdages, M.; Douville, P. Effects of electric and magnetic fields from high-power lines on female urinary excretion of 6-sulfatoxymelatonin. *Am. J. Epidemiol.* **2001**, *154*, 601–609. [CrossRef] [PubMed]
81. Feychtig, M. Invited commentary: Extremely low-frequency magnetic fields and breast cancer now it is enough! *Am. J. Epidemiol.* **2013**, *178*, 1046–1050. [CrossRef] [PubMed]
82. Touitou, Y.; Lambrozo, J.; Camus, F.O.; Charbuy, H. Magnetic fields and the melatonin hypothesis: A study of workers chronically exposed to 50-hz magnetic fields. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2003**, *284*, R1529–R1535. [CrossRef] [PubMed]
83. Touitou, Y.; Selmaoui, B.; Lambrozo, J.; Auzeby, A. Assessment of the effects of magnetic fields (50 hz) on melatonin secretion in humans and rats. A circadian study. *Bull. Acad. Natl. Med.* **2002**, *186*, 1625–1639. [PubMed]
84. Davis, S.; Mirick, D.K. Residential magnetic fields, medication use, and the risk of breast cancer. *Epidemiology* **2007**, *18*, 266–269. [CrossRef] [PubMed]
85. de Bruyn, L.; de Jager, L.; Kuyl, J.M. The influence of long-term exposure of mice to randomly varied power frequency magnetic fields on their nocturnal melatonin secretion patterns. *Environ. Res.* **2001**, *85*, 115–121. [CrossRef] [PubMed]
86. O’Leary, E.S.; Schoenfeld, E.R.; Henderson, K.; Grimson, R.; Kabat, G.C.; Kaune, W.T.; Gammon, M.D.; Leske, C.; Grp, E. Wire coding in the EMF and breast cancer on long island study: Relationship to magnetic fields. *J. Expo. Anal. Environ. Epidemiol.* **2003**, *13*, 283–293. [CrossRef]
87. London, S.J.; Pogoda, J.M.; Hwang, K.L.; Langholz, B.; Monroe, K.R.; Kolonel, L.N.; Kaune, W.T.; Peters, J.M.; Henderson, B.E. Residential magnetic field exposure and breast cancer risk: A nested case-control study from a multiethnic cohort in los angeles county, california. *Am. J. Epidemiol.* **2003**, *158*, 969–980. [CrossRef] [PubMed]
88. Haugsdal, B.; Tynes, T.; Rotnes, J.S.; Griffiths, D. A single nocturnal exposure to 2–7 millitesla static magnetic fields does not inhibit the excretion of 6-sulfatoxymelatonin in healthy young men. *Bioelectromagnetics* **2001**, *22*, 1–6. [CrossRef]
89. Fedrowitz, M.; Westermann, J.; Loscher, W. Magnetic field exposure increases cell proliferation but does not affect melatonin levels in the mammary gland of female sprague dawley rats. *Cancer Res.* **2002**, *62*, 1356–1363. [PubMed]
90. Vanderstraeten, J.; Verschaeve, L.; Burda, H.; Boulard, C.; Brouwer, C. Health effects of extremely low-frequency magnetic fields: Reconsidering the melatonin hypothesis in the light of current data on magnetoreception. *J. Appl. Toxicol.* **2012**, *32*, 952–958. [CrossRef]
91. Zhang, Y.M.; Lai, J.S.; Ruan, G.R.; Chen, C.; Wang, D.W. Meta-analysis of extremely low frequency electromagnetic fields and cancer risk: A pooled analysis of epidemiologic studies. *Environ. Int.* **2016**, *88*, 36–43. [CrossRef]

92. Zhu, K.M.; Hunter, S.; Payne-Wilks, K.; Roland, C.L.; Forbes, D.S. Use of electric bedding devices and risk of breast cancer in african-american women. *Am. J. Epidemiol.* **2003**, *158*, 798–806. [CrossRef] [PubMed]
93. Laden, F.; Hunter, D.J. Environmental risk factors and female breast cancer. *Annu. Rev. Public Health* **1998**, *19*, 101–123. [CrossRef] [PubMed]
94. Amoon, A.T.; Swanson, J.; Magnani, C.; Johansen, C.; Kheifets, L. Pooled analysis of recent studies of magnetic fields and childhood leukemia. *Environ. Res.* **2022**, *204*, 7. [CrossRef] [PubMed]
95. Elwood, J.M. Childhood leukemia and residential magnetic fields: Are pooled analyses more valid than the original studies? *Bioelectromagnetics* **2006**, *27*, 112–118. [CrossRef] [PubMed]
96. Schuz, J. Exposure to extremely low-frequency magnetic fields and the risk of childhood cancer: Update of the epidemiological evidence. *Prog. Biophys. Mol. Biol.* **2011**, *107*, 339–342. [CrossRef]
97. Ghahremani, S.; Shiroudbakhti, K.; Kordasiabi, A.H.S.; FiroozBakht, M.; Hosseinzadegan, M.; Ashrafinia, F.; Rahafard, S. Exposure to magnetic fields and childhood leukemia: An overview of meta-analysis. *Int. J. Pediatr.-Mashhad* **2020**, *8*, 11361–11365.
98. Tafrishi, R.; Seyfari, B.; Rahimi, R.; Chaichi, Z.; Tarazjani, A.D.; Marvi, N.; Maazallah, M.; Dolatian, Z.; Ashrafinia, F. Modifiable and non-modifiable factors affecting the risk of childhood leukemia: An overview of meta-analysis. *Int. J. Pediatr.* **2021**, *9*, 13243–13248.
99. Zhao, L.Y.; Liu, X.D.; Wang, C.P.; Yan, K.K.; Lin, X.J.; Li, S.; Bao, H.H.; Liu, X. Magnetic fields exposure and childhood leukemia risk: A meta-analysis based on 11,699 cases and 13,194 controls. *Leuk. Res.* **2014**, *38*, 269–274. [CrossRef] [PubMed]
100. Amoon, A.T.; Crespi, C.M.; Ahlbom, A.; Bhatnagar, M.; Bray, I.; Bunch, K.J.; Clavel, J.; Feychtig, M.; Hemon, D.; Johansen, C.; et al. Proximity to overhead power lines and childhood leukaemia: An international pooled analysis. *Br. J. Cancer* **2018**, *119*, 364–373. [CrossRef] [PubMed]
101. Pedersen, C.; Brauner, E.V.; Rod, N.H.; Albieri, V.; Andersen, C.E.; Ulbak, K.; Hertel, O.; Johansen, C.; Schuz, J.; Raaschou-Nielsen, O. Distance to high-voltage power lines and risk of childhood leukemia—An analysis of confounding by and interaction with other potential risk factors. *PLoS ONE* **2014**, *9*, e107096. [CrossRef] [PubMed]
102. Pedersen, C.; Johansen, C.; Schuz, J.; Olsen, J.H.; Raaschou-Nielsen, O. Residential exposure to extremely low-frequency magnetic fields and risk of childhood leukaemia, cns tumour and lymphoma in denmark. *Br. J. Cancer* **2015**, *113*, 1370–1374. [CrossRef]
103. Nunez-Enriquez, J.C.; Correa-Correa, V.; Flores-Lujano, J.; Perez-Saldivar, M.L.; Jimenez-Hernandez, E.; Martin-Trejo, J.A.; Espinoza-Hernandez, L.E.; Medina-Sanson, A.; Cardenas-Cardos, R.; Flores-Villegas, L.V.; et al. Extremely low-frequency magnetic fields and the risk of childhood b-lineage acute lymphoblastic leukemia in a city with high incidence of leukemia and elevated exposure to elff magnetic fields. *Bioelectromagnetics* **2020**, *41*, 581–597. [CrossRef] [PubMed]
104. Schuz, J.; Grigat, J.P.; Brinkmann, K.; Michaelis, J. Residential magnetic fields as a risk factor for childhood acute leukaemia: Results from a german population-based case-control study. *Int. J. Cancer* **2001**, *91*, 728–735. [CrossRef]
105. Kheifets, L.; Crespi, C.M.; Hooper, C.; Cockburn, M.; Amoon, A.T.; Vergara, X.P. Residential magnetic fields exposure and childhood leukemia: A population-based case-control study in california. *Cancer Causes Control* **2017**, *28*, 1117–1123. [CrossRef]
106. Crespi, C.M.; Swanson, J.; Vergara, X.P.; Kheifets, L. Childhood leukemia risk in the california power line study: Magnetic fields versus distance from power lines. *Environ. Res.* **2019**, *171*, 530–535. [CrossRef] [PubMed]
107. Crespi, C.M.; Vergara, X.P.; Hooper, C.; Oksuzyan, S.; Wu, S.; Cockburn, M.; Kheifets, L. Childhood leukaemia and distance from power lines in california: A population-based case-control study. *Br. J. Cancer* **2016**, *115*, 122–128. [CrossRef]
108. Sermage-Faure, C.; Demoury, C.; Rudant, J.; Goujon-Bellec, S.; Guyot-Goubin, A.; Deschamps, F.; Hemon, D.; Clavel, J. Childhood leukaemia close to high-voltage power lines—The geocap study, 2002–2007. *Br. J. Cancer* **2013**, *108*, 1899–1906. [CrossRef]
109. Tabrizi, M.M.; Bidgoli, S.-A. Increased risk of childhood acute lymphoblastic leukemia (all) by prenatal and postnatal exposure to high voltage power lines: A case control study in isfahan, iran. *Asian Pac. J. Cancer Prev.* **2015**, *16*, 2347–2350. [CrossRef] [PubMed]
110. Schuz, J.; Dasenbrock, C.; Ravazzani, P.; Roosli, M.; Schar, P.; Bounds, P.L.; Erdmann, F.; Borkhardt, A.; Cobaleda, C.; Fedrowitz, M.; et al. Extremely low-frequency magnetic fields and risk of childhood leukemia: A risk assessment by the arimmora consortium. *Bioelectromagnetics* **2016**, *37*, 183–189. [CrossRef] [PubMed]
111. Sermage-Faure, C.; Laurier, D.; Goujon-Bellec, S.; Chartier, M.; Guyot-Goubin, A.; Rudant, J.; Hemon, D.; Clavel, J. Childhood leukemia around french nuclear power plantsuthe geocap study, 2002–2007. *Int. J. Cancer* **2012**, *131*, E769–E780. [CrossRef] [PubMed]
112. Ziegelberger, G.; Dehos, A.; Grosche, B.; Hornhardt, S.; Jung, T.; Weiss, W. Childhood leukemia—Risk factors and the need for an interdisciplinary research agenda. *Prog. Biophys. Mol. Biol.* **2011**, *107*, 312–314. [CrossRef]
113. Wünsch, V.; Pelissari, D.M.; Barbieri, F.E.; Sant'Anna, L.; de Oliveira, C.T.; de Mata, J.F.; Tone, L.G.; Lee, M.L.D.; de Andrea, M.L.M.; Bruniera, P.; et al. Exposure to magnetic fields and childhood acute lymphocytic leukemia in Sao Paulo, Brazil. *Cancer Epidemiol.* **2011**, *35*, 534–539. [CrossRef]
114. Jirik, V.; Pekarek, L.; Janout, V.; Tomaskova, H. Association between childhood leukaemia and exposure to power-frequency magnetic fields in Middle Europe. *Biomed. Environ. Sci.* **2012**, *25*, 597–601. [PubMed]
115. Hakim, A.S.B.; Abd Rahman, N.B.; Mokhtar, M.Z.; Bin Said, I.; Hussain, H. ELF-EMF correlation study on distance from overhead transmission lines and acute leukemia among children in Klang Valley, Malaysia. In Proceedings of the 2014 IEEE Conference on Biomedical Engineering and Sciences (IECBES), Kuala Lumpur, Malaysia, 8–10 December 2014; pp. 710–714.
116. Karipidis, K.K. Survey of residential power-frequency magnetic fields in Melbourne, Australia. *Radiat. Prot. Dosim.* **2015**, *163*, 81–91. [CrossRef] [PubMed]

117. Jirík, V.; Pekarek, L.; Janout, V. Assessment of population exposure to extremely low frequency magnetic fields and its possible childhood health risk in the Czech Republic. *Indoor Built Environ.* **2011**, *20*, 362–368. [[CrossRef](#)]
118. Kokate, P.A.; Mishra, A.K.; Lokhande, S.K.; Bodhe, G.L. Extremely low frequency electromagnetic field (ELF-EMF) and childhood leukemia (cl) near transmission lines: A review. *Adv. Electromagn.* **2016**, *5*, 30–40. [[CrossRef](#)]
119. Zaki, A.M.; Abd Rahim, M.A.; Zaidun, Z.; Ramdzan, A.R.; Isa, Z.M. Exposure to non-ionizing radiation and childhood cancer: A meta-analysis. *Middle East J. Cancer* **2020**, *11*, 1–11.
120. Swanson, J.; Bunch, K.J. Reanalysis of risks of childhood leukaemia with distance from overhead power lines in the UK. *J. Radiol. Prot.* **2018**, *38*, N30–N35. [[CrossRef](#)] [[PubMed](#)]
121. Liorni, I.; Parazzini, M.; Struchen, B.; Fiocchi, S.; Roosli, M.; Ravazzani, P. Children's personal exposure measurements to extremely low frequency magnetic fields in Italy. *Int. J. Environ. Res. Public Health* **2016**, *13*, 549. [[CrossRef](#)]
122. Amoon, A.T.; Arah, O.A.; Kheifets, L. The sensitivity of reported effects of EMF on childhood leukemia to uncontrolled confounding by residential mobility: A hybrid simulation study and an empirical analysis using caps data. *Cancer Causes Control* **2019**, *30*, 901–908. [[CrossRef](#)] [[PubMed](#)]
123. Amoon, A.T.; Oksuzyan, S.; Crespi, C.M.; Arah, O.A.; Cockburn, M.; Vergara, X.; Kheifets, L. Residential mobility and childhood leukemia. *Environ. Res.* **2018**, *164*, 459–466. [[CrossRef](#)] [[PubMed](#)]
124. Bunch, K.J.; Swanson, J.; Vincent, T.J.; Murphy, M.F.G. Magnetic fields and childhood cancer: An epidemiological investigation of the effects of high-voltage underground cables. *J. Radiol. Prot.* **2015**, *35*, 695–705. [[CrossRef](#)] [[PubMed](#)]
125. Auger, N.; Bilodeau-Bertrand, M.; Marcoux, S.; Kosatsky, T. Residential exposure to electromagnetic fields during pregnancy and risk of child cancer: A longitudinal cohort study. *Environ. Res.* **2019**, *176*, 108524. [[CrossRef](#)] [[PubMed](#)]
126. Soderberg, K.C.; Naumburg, E.; Anger, G.; Cnattingius, S.; Ekbom, A.; Feychtig, M. Childhood leukemia and magnetic fields in infant incubators. *Epidemiology* **2002**, *13*, 45–49. [[CrossRef](#)] [[PubMed](#)]
127. Slusky, D.A.; Does, M.; Metayer, C.; Mezei, G.; Selvin, S.; Buffler, P.A. Potential role of selection bias in the association between childhood leukemia and residential magnetic fields exposure: A population-based assessment. *Cancer Epidemiol.* **2014**, *38*, 307–313. [[CrossRef](#)] [[PubMed](#)]
128. Leitgeb, N. Synoptic analysis clarifies childhood leukemia risk from elc magnetic field exposure. *J. Electromagn. Anal. Appl.* **2015**, *7*, 10. [[CrossRef](#)]
129. Schuz, J.; Erdmann, F. Environmental exposure and risk of childhood leukemia: An overview. *Arch. Med. Res.* **2016**, *47*, 607–614. [[CrossRef](#)]
130. Salvan, A.; Ranucci, A.; Lagorio, S.; Magnani, C.; Grp, S.R. Childhood leukemia and 50 hz magnetic fields: Findings from the Italian setil case-control study. *Int. J. Environ. Res. Public Health* **2015**, *12*, 2184–2204. [[CrossRef](#)]
131. Omura, Y.; Losco, M. Electromagnetic-fields in the home-environment (color tv, computer monitor, microwave-oven, cellular phone, etc) as potential contributing factors for the induction of oncogen c-fos ab1, oncogen c-fos ab2, integrin alpha-5-beta-1 and development of cancer, as well as effects of microwave on amino-acid-composition of food and living human brain. *Acupunct. Electro-Ther. Res.* **1993**, *18*, 33–73.
132. Verreault, R.; Weiss, N.S.; Hollenbach, K.A.; Strader, C.H.; Daling, J.R. Use of electric blankets and risk of testicular cancer. *Am. J. Epidemiol.* **1990**, *131*, 759–762. [[CrossRef](#)]
133. Zhu, K.; Weiss, N.S.; Stanford, J.L.; Daling, J.R.; Stergachis, A.; McKnight, B.; Brawer, M.K.; Levine, R.S. Prostate cancer in relation to the use of electric blanket or heated water bed. *Epidemiology* **1999**, *10*, 83–85. [[CrossRef](#)]
134. McElroy, J.A.; Newcomb, P.A.; Trentham-Dietz, A.; Hampton, J.M.; Kanarek, M.S.; Remington, P.L. Endometrial cancer incidence in relation to electric blanket use. *Am. J. Epidemiol.* **2002**, *156*, 262–267. [[CrossRef](#)]
135. Gudina, M.V.; Borodin, A.S.; Tuzhilkin, D.A.; Pikalova, L.V. Malignant neoplasms on territories with different levels of magnetic fields of industrial frequency. In Proceedings of the 24th International Symposium on Atmospheric and Ocean Optics: Atmospheric Physics, Tomsk, Russia, 2–5 July 2018; Matvienko, G.G., Romanovskii, O.A., Eds.; SPIE—International Society for Optics and Photonics: Bellingham, WA, USA, 2018; Volume 10833.
136. Hallberg, O.; Johansson, O. Melanoma incidence and frequency modulation (FM) broadcasting. *Arch. Environ. Health* **2002**, *57*, 32–40. [[CrossRef](#)]
137. Mahram, M.; Ghazavi, M. The effect of extremely low frequency electromagnetic fields on pregnancy and fetal growth, and development. *Arch. Iran. Med.* **2013**, *16*, 221–224.
138. Deadman, J.E.; Infante-Rivard, C. Individual estimation of exposures to extremely low frequency magnetic fields in jobs commonly held by women. *Am. J. Epidemiol.* **2002**, *155*, 368–378. [[CrossRef](#)]
139. Ptitsyna, N.G.; Villoresi, G.; Kopytenko, Y.A. Railway-generated magnetic field: Environmental aspects. *Railw. Transp. Policies Technol. Perspect.* **2009**, 87–140.
140. Bowman, J.D.; Methner, M.M. Hazard surveillance for industrial magnetic fields: II. Field characteristics from waveform measurements. *Ann. Occup. Hyg.* **2000**, *44*, 615–633. [[CrossRef](#)]
141. Methner, M.M.; Bowman, J.D. Hazard surveillance for industrial magnetic fields: I. Walkthrough survey of ambient fields and sources. *Ann. Occup. Hyg.* **2000**, *44*, 603–614. [[CrossRef](#)]
142. ILO. *Radiation Protection of Workers—Ionizing Radiation*, ILO Code of Practice; International Labour Office: Geneva, Switzerland, 1987.
143. ICNIRP. Guidelines for limiting exposure to time-varying electric and magnetic fields (1 hz–100 khz). *Health Phys.* **2010**, *99*, 818–836. [[CrossRef](#)]

144. Engel, C.L.; Rasanayagam, M.S.; Gray, J.M.; Rizzo, J. Work and female breast cancer: The state of the evidence, 2002–2017. *New Solut.* **2018**, *28*, 55–78. [[CrossRef](#)]
145. McCurdy, A.L.; Wijnberg, L.; Loomis, D.; Savitz, D.; Nylander-French, L.A. Exposure to extremely low frequency magnetic fields among working women and homemakers. *Ann. Occup. Hyg.* **2001**, *45*, 643–650. [[CrossRef](#)]
146. Li, P.; McLaughlin, J.; Infante-Rivard, C. Maternal occupational exposure to extremely low frequency magnetic fields and the risk of brain cancer in the offspring. *Cancer Causes Control* **2009**, *20*, 945–955. [[CrossRef](#)]
147. Sorahan, T. Magnetic fields and brain tumour risks in uk electricity supply workers. *Occup. Med.* **2014**, *64*, 157–165. [[CrossRef](#)] [[PubMed](#)]
148. Turner, M.C.; Benke, G.; Bowman, J.D.; Figuerola, J.; Fleming, S.; Hours, M.; Kincl, L.; Krewski, D.; McLean, D.; Parent, M.E.; et al. Occupational exposure to extremely low-frequency magnetic fields and brain tumor risks in the interocc study. *Cancer Epidemiol. Biomark. Prev.* **2014**, *23*, 1863–1872. [[CrossRef](#)] [[PubMed](#)]
149. Oraby, T.; Sivaganesan, S.; Bowman, J.D.; Kincl, L.; Richardson, L.; McBride, M.; Siemiatycki, J.; Cardis, E.; Krewski, D.; On behalf of the INTEROCC Study Group. Berkson error adjustment and other exposure surrogates in occupational case-control studies, with application to the canadian interocc study. *J. Expo. Sci. Environ. Epidemiol.* **2018**, *28*, 251–258. [[CrossRef](#)] [[PubMed](#)]
150. Carlberg, M.; Koppel, T.; Ahonen, M.; Hardell, L. Case-control study on occupational exposure to extremely low-frequency electromagnetic fields and the association with meningioma. *BioMed Res. Int.* **2018**, *2018*, 5912394. [[CrossRef](#)]
151. Goodman, M.; Kelsh, M.; Ebi, K.; Iannuzzi, J.; Langholz, B. Evaluation of potential confounders in planning a study of occupational magnetic field exposure and female breast cancer. *Epidemiology* **2002**, *13*, 50–58. [[CrossRef](#)]
152. Kelsh, M.A.; Bracken, T.D.; Sahl, J.D.; Shum, M.; Ebi, K.L. Occupational magnetic field exposures of garment workers: Results of personal and survey measurements. *Bioelectromagnetics* **2003**, *24*, 316–326. [[CrossRef](#)]
153. Juutilainen, J.; Kumlin, T. Occupational magnetic field exposure and melatonin: Interaction with light-at-night. *Bioelectromagnetics* **2006**, *27*, 423–426. [[CrossRef](#)]
154. Hardell, L.; Holmberg, B.; Malker, H.; Paulsson, L.E. Exposure to extremely-low-frequency electromagnetic-fields and the risk of malignant diseases—An evaluation of epidemiologic and experimental findings. *Eur. J. Cancer Prev.* **1995**, *4*, 3–107. [[CrossRef](#)] [[PubMed](#)]
155. McElroy, J.A.; Egan, K.M.; Titus-Ernstoff, L.; Anderson, H.A.; Trentham-Dietz, A.; Hampton, J.M.; Newcomb, P.A. Occupational exposure to electromagnetic field and breast cancer risk in a large, population based, case-control study in the united states. *J. Occup. Environ. Med.* **2007**, *49*, 266–274. [[CrossRef](#)]
156. Grundy, A.; Harris, S.A.; Demers, P.A.; Johnson, K.C.; Agnew, D.A.; Villeneuve, P.J.; Canadian Canc Registries, E. Occupational exposure to magnetic fields and breast cancer among canadian men. *Cancer Med.* **2016**, *5*, 586–596. [[CrossRef](#)]
157. Minder, C.E.; Pfluger, D.H. Leukemia, brain tumors, and exposure to extremely low frequency electromagnetic fields in swiss railway employees. *Am. J. Epidemiol.* **2001**, *153*, 825–835. [[CrossRef](#)] [[PubMed](#)]
158. Cam, S.T.; Firlarler, A.; Ozden, S.; Canseven, A.G.; Seyhan, N. Occupational exposure to magnetic fields from transformer stations and electric enclosures in Turkey. *Electromagn. Biol. Med.* **2011**, *30*, 74–79. [[CrossRef](#)] [[PubMed](#)]
159. Bethwaite, P.; Cook, A.; Kennedy, J.; Pearce, N. Acute leukemia in electrical workers: A new zealand case-control study. *Cancer Causes Control* **2001**, *12*, 683–689. [[CrossRef](#)] [[PubMed](#)]
160. Kheifets, L.; Monroe, J.; Vergara, X.; Mezei, G.; Afifi, A.A. Occupational electromagnetic fields and leukemia and brain cancer: An update to two meta-analyses. *J. Occup. Environ. Med.* **2008**, *50*, 677–688. [[CrossRef](#)]
161. Roosli, M.; Lortscher, M.; Egger, M.; Pfluger, D.; Schreier, N.; Lortscher, E.; Locher, P.; Spoerri, A.; Minder, C. Leukaemia, brain tumours and exposure to extremely low frequency magnetic fields: Cohort study of Swiss railway employees. *Occup. Environ. Med.* **2007**, *64*, 553–559. [[CrossRef](#)]
162. Huss, A.; Spoerri, A.; Egger, M.; Kromhout, H.; Vermeulen, R.; Swiss Natl, C. Occupational extremely low frequency magnetic fields (elf-mf) exposure and hematolymphopoietic cancers—Swiss national cohort analysis and updated meta-analysis. *Environ. Res.* **2018**, *164*, 467–474. [[CrossRef](#)]
163. Nordenson, I.; Mild, K.H.; Jarventaus, H.; Hirvonen, A.; Sandstrom, M.; Wilen, J.; Blix, N.; Norppa, H. Chromosomal aberrations in peripheral lymphocytes of train engine drivers. *Bioelectromagnetics* **2001**, *22*, 306–315. [[CrossRef](#)]
164. Talibov, M.; Guxens, M.; Pukkala, E.; Huss, A.; Kromhout, H.; Slottje, P.; Martinsen, J.I.; Kjaerheim, K.; Spare, P.; Weiderpass, E.; et al. Occupational exposure to extremely low-frequency magnetic fields and electrical shocks and acute myeloid leukemia in four nordic countries. *Cancer Causes Control* **2015**, *26*, 1079–1085. [[CrossRef](#)]
165. Tynes, T.; Haldorsen, T. Residential and occupational exposure to 50 hz magnetic fields and hematological cancers in Norway. *Cancer Causes Control* **2003**, *14*, 715–720. [[CrossRef](#)]
166. Harrington, J.M.; Nichols, L.; Sorahan, T.; van Tongeren, M. Leukaemia mortality in relation to magnetic field exposure: Findings from a study of united kingdom electricity generation and transmission workers, 1973–1997. *Occup. Environ. Med.* **2001**, *58*, 307–314. [[CrossRef](#)]
167. Willett, E.V.; McKinney, P.A.; Fear, N.T.; Cartwright, R.A.; Roman, E. Occupational exposure to electromagnetic fields and acute leukaemia: Analysis of a case-control study. *Occup. Environ. Med.* **2003**, *60*, 577–583. [[CrossRef](#)] [[PubMed](#)]
168. Hug, K.; Grize, L.; Seidler, A.; Kaatsch, P.; Schuz, J. Parental occupational exposure to extremely low frequency magnetic fields and childhood cancer: A german case-control study. *Am. J. Epidemiol.* **2010**, *171*, 27–35. [[CrossRef](#)] [[PubMed](#)]

169. Talibov, M.; Olsson, A.; Bailey, H.; Erdmann, F.; Metayer, C.; Magnani, C.; Petridou, E.; Auvinen, A.; Spector, L.; Clavel, J.; et al. Parental occupational exposure to low-frequency magnetic fields and risk of leukaemia in the offspring: Findings from the childhood leukaemia international consortium (CLIC). *Occup. Environ. Med.* **2019**, *76*, 746–753. [[CrossRef](#)]
170. Su, L.L.; Fei, Y.; Wei, X.X.; Guo, J.; Jiang, X.G.; Lu, L.Q.; Chen, G.D. Associations of parental occupational exposure to extremely low-frequency magnetic fields with childhood leukemia risk. *Leuk. Lymphoma* **2016**, *57*, 2855–2862. [[CrossRef](#)]
171. Hakansson, N.; Floderus, B.; Gustavsson, P.; Johansen, C.; Olsen, J.H. Cancer incidence and magnetic field exposure in industries using resistance welding in Sweden. *Occup. Environ. Med.* **2002**, *59*, 481–486. [[CrossRef](#)]
172. Yousif, L.; Blettner, M.; Hammer, G.P.; Zeeb, H. Testicular cancer risk associated with occupational radiation exposure: A systematic literature review. *J. Radiol. Prot.* **2010**, *30*, 389–406. [[CrossRef](#)]
173. Karipidis, K.; Benke, G.; Sim, M.; Fritschi, L.; Yost, M.; Armstrong, B.; Hughes, A.M.; Grulich, A.; Vajdic, C.M.; Kaldor, J.M.; et al. Occupational exposure to power frequency magnetic fields and risk of non-hodgkin lymphoma. *Occup. Environ. Med.* **2007**, *64*, 25–29. [[CrossRef](#)]
174. Forssen, U.M.; Lonn, S.; Ahlbom, A.; Savitz, D.A.; Feychtig, M. Occupational magnetic field exposure and the risk of acoustic neuroma. *Am. J. Ind. Med.* **2006**, *49*, 112–118. [[CrossRef](#)]
175. Carlberg, M.; Koppel, T.; Ahonen, M.; Hardell, L. Case-control study on occupational exposure to extremely low-frequency electromagnetic fields and the association with acoustic neuroma. *Environ. Res.* **2020**, *187*, 109621. [[CrossRef](#)]
176. Lope, V.; Perez-Gomez, B.; Aragones, N.; Lopez-Abente, G.; Gustavsson, P.; Floderus, B.; Dosemeci, M.; Silva, A.; Pollan, M. Occupational exposure to ionizing radiation and electromagnetic fields in relation to the risk of thyroid cancer in Sweden. *Scand. J. Work Environ. Health* **2006**, *32*, 276–284. [[CrossRef](#)]
177. De Roos, A.J.; Teschke, K.; Savitz, D.A.; Poole, C.; Grufferman, S.; Pollock, B.H.; Olshan, A.F. Parental occupational exposures to electromagnetic fields and radiation and the incidence of neuroblastoma in offspring. *Epidemiology* **2001**, *12*, 508–517. [[CrossRef](#)] [[PubMed](#)]
178. Sahl, J.; Mezei, G.; Kavet, R.; McMillan, A.; Silvers, A.; Sastre, A.; Kheifets, L. Occupational magnetic field exposure and cardiovascular mortality in a cohort of electric utility workers. *Am. J. Epidemiol.* **2002**, *156*, 913–918. [[CrossRef](#)] [[PubMed](#)]
179. Lange, S.; Viergutz, T.; Simko, M. Modifications in cell cycle kinetics and in expression of g(1) phase-regulating proteins in human amniotic cells after exposure to electromagnetic fields and ionizing radiation. *Cell Prolif.* **2004**, *37*, 337–349. [[CrossRef](#)] [[PubMed](#)]
180. Huang, L.Z.; Dong, L.A.; Chen, Y.T.; Qi, H.S.; Xiao, D.M. Effects of sinusoidal magnetic field observed on cell proliferation, ion concentration, and osmolarity in two human cancer cell lines. *Electromagn. Biol. Med.* **2006**, *25*, 113–126. [[CrossRef](#)] [[PubMed](#)]
181. Buemi, M.; Marino, D.; Di Pasquale, G.; Floccari, F.; Senatore, M.; Aloisi, C.; Grasso, F.; Mondio, G.; Perillo, P.; Frisina, N.; et al. Cell proliferation/cell death balance in renal cell cultures after exposure to a static magnetic field. *Nephron* **2001**, *87*, 269–273. [[CrossRef](#)]
182. Sztafrowski, D.; Jazwiec, B.; Gumiela, J.; Kuliczkowski, K. Influence of north and south poles of static magnetic field (SMF) on apoptosis of hl60 cell line. *Prz. Elektrotech.* **2018**, *94*, 182–185.
183. Tenuzzo, B.; Chionna, A.; Panzarini, E.; Lanubile, R.; Tarantino, P.; Di Jeso, B.; Dwikat, M.; Dini, L. Biological effects of 6 mt static magnetic fields: A comparative study in different cell types. *Bioelectromagnetics* **2006**, *27*, 560–577. [[CrossRef](#)]
184. Naziroglu, M.; Clg, B.; Dogan, S.; Uguz, A.C.; Dilek, S.; Faouzi, D. 2.45-gz wireless devices induce oxidative stress and proliferation through cytosolic ca²⁺ influx in human leukemia cancer cells. *Int. J. Radiat. Biol.* **2012**, *88*, 449–456. [[CrossRef](#)]
185. Barati, M.; Javidi, M.A.; Darvishi, B.; Shariatpanahi, S.P.; Moosavi, Z.S.M.; Ghadirian, R.; Khani, T.; Sanati, H.; Simaei, H.; Barough, M.S.; et al. Necroptosis triggered by ros accumulation and Ca²⁺ overload, partly explains the inflammatory responses and anti-cancer effects associated with 1 hz, 100 mt elf-mf in vivo. *Free Radic. Biol. Med.* **2021**, *169*, 84–98. [[CrossRef](#)]
186. Rosen, A.D. Mechanism of action of moderate-intensity static magnetic fields on biological systems. *Cell Biochem. Biophys.* **2003**, *39*, 163–173. [[CrossRef](#)]
187. Wei, J.; Sun, J.; Xu, H.; Shi, L.; Sun, L.; Zhang, J. Effects of extremely low frequency electromagnetic fields on intracellular calcium transients in cardiomyocytes. *Electromagn. Biol. Med.* **2015**, *34*, 77–84. [[CrossRef](#)] [[PubMed](#)]
188. Chen, Y.C.; Chen, C.C.; Tu, W.; Cheng, Y.T.; Tseng, F.G. Design and fabrication of a microplatform for the proximity effect study of localized elf-EMF on the growth of in vitro hela and pc-12 cells. *J. Micromech. Microeng.* **2010**, *20*, 125023. [[CrossRef](#)]
189. Bae, J.E.; Do, J.Y.; Kwon, S.H.; Lee, S.D.; Jung, Y.W.; Kim, S.C.; Chae, K.S. Electromagnetic field-induced converse cell growth during a long-term observation. *Int. J. Radiat. Biol.* **2013**, *89*, 1035–1044. [[CrossRef](#)] [[PubMed](#)]
190. Richard, D.; Lange, S.; Viergutz, T.; Kriehuber, R.; Weiss, D.G.; Simko, M. Influence of 50 hz electromagnetic fields in combination with a tumour promoting phorbol ester on protein kinase c and cell cycle in human cells. *Mol. Cell. Biochem.* **2002**, *232*, 133–141. [[CrossRef](#)] [[PubMed](#)]
191. Kapri-Pardes, E.; Hanoch, T.; Maik-Rachline, G.; Murbach, M.; Bounds, P.L.; Kuster, N.; Seger, R. Activation of signaling cascades by weak extremely low frequency electromagnetic fields. *Cell. Physiol. Biochem.* **2017**, *43*, 1533–1546. [[CrossRef](#)] [[PubMed](#)]
192. Kirschenlohr, H.; Ellis, P.; Hesketh, R.; Metcalfe, J. Gene expression profiles in white blood cells of volunteers exposed to a 50 hz electromagnetic field. *Radiat. Res.* **2012**, *178*, 138–149. [[CrossRef](#)] [[PubMed](#)]
193. Kabacik, S.; Kirschenlohr, H.; Raffy, C.; Whitehill, K.; Coster, M.; Abe, M.; Brindle, K.; Badie, C.; Sienkiewicz, Z.; Bouffler, S. Investigation of transcriptional responses of juvenile mouse bone marrow to power frequency magnetic fields. *Mutat. Res./Fundam. Mol. Mech. Mutagenesis* **2013**, *745*, 40–45. [[CrossRef](#)]

194. Panagopoulos, D.J.; Karabarounis, A.; Lioliosis, C. Elf alternating magnetic field decreases reproduction by DNA damage induction. *Cell Biochem. Biophys.* **2013**, *67*, 703–716. [CrossRef]
195. Pardo, J.C.T.; Grimaldi, S.; Taranta, M.; Naldi, I.; Cinti, C. Microwave electromagnetic field regulates gene expression in t-lymphoblastoid leukemia ccrf-cem cell line exposed to 900 mhz. *Electromagn. Biol. Med.* **2012**, *31*, 1–18. [CrossRef]
196. Zhou, J.L.; Li, C.L.; Yao, G.D.; Chiang, H.A.; Chang, Z.L. Gene expression of cytokine receptors in hl60 cells exposed to a 50 hz magnetic field. *Bioelectromagnetics* **2002**, *23*, 339–346. [CrossRef]
197. Diab, K.A. The impact of the low frequency of the electromagnetic field on human. In *Cell Biology and Translational Medicine, Volume 7: Stem Cells and Therapy: Emerging Approaches*; Turksen, K., Ed.; Springer: Berlin/Heidelberg, Germany, 2020; Volume 1237, pp. 135–149.
198. Giorgi, G.; Del Re, B. Epigenetic dysregulation in various types of cells exposed to extremely low-frequency magnetic fields. *Cell Tissue Res.* **2021**, *386*, 1–15. [CrossRef] [PubMed]
199. Harada, S.; Yamada, S.; Kuramata, O.; Gunji, Y.; Kawasaki, M.; Miyakawa, T.; Yonekura, H.; Sakurai, S.; Bessho, K.; Hosono, R.; et al. Effects of high elf magnetic fields on enzyme-catalyzed DNA and RNA synthesis in vitro and on a cell-free DNA mismatch repair. *Bioelectromagnetics* **2001**, *22*, 260–266. [CrossRef] [PubMed]
200. Li, H.L.; Lin, L.; Li, L.; Zhou, L.; Zhang, Y.; Hao, S.; Ding, Z.H. Exosomal small rna sequencing uncovers the microrna dose markers for power frequency electromagnetic field exposure. *Biomarkers* **2018**, *23*, 315–327. [CrossRef] [PubMed]
201. Manser, M.; Sater, M.R.A.; Schmid, C.D.; Noreen, F.; Murbach, M.; Kuster, N.; Schuermann, D.; Schar, P. ELF-MF exposure affects the robustness of epigenetic programming during granulopoiesis. *Sci. Rep.* **2017**, *7*, 43345. [CrossRef]
202. Sun, W.J.; Gan, Y.P.; Fu, Y.T.; Lu, D.Q.; Chiang, H. An incoherent magnetic field inhibited EGF receptor clustering and phosphorylation induced by a 50-hz magnetic field in cultured FL cells. *Cell. Physiol. Biochem.* **2008**, *22*, 507–514. [CrossRef]
203. Ke, X.Q.; Sun, W.J.; Lu, D.Q.; Fu, Y.T.; Chiang, H. 50-hz magnetic field induces EGF-receptor clustering and activates RAS. *Int. J. Radiat. Biol.* **2008**, *84*, 413–420. [CrossRef] [PubMed]
204. Eleuteri, A.M.; Amici, M.; Bonfili, L.; Cecarini, V.; Cuccioloni, M.; Grimaldi, S.; Giuliani, L.; Angeletti, M.; Fioretti, E. 50 hz extremely low frequency electromagnetic fields enhance protein carbonyl groups content in cancer cells: Effects on proteasomal systems. *J. Biomed. Biotechnol.* **2009**, *2009*, 834239. [CrossRef]
205. Testa, A.; Cordelli, E.; Stronati, L.; Marino, C.; Lovisolo, G.A.; Freseagna, A.M.; Conti, D.; Villani, P. Evaluation of genotoxic effect of low level 50 hz magnetic fields on human blood cells using different cytogenetic assays. *Bioelectromagnetics* **2004**, *25*, 613–619. [CrossRef]
206. Stronati, L.; Testa, A.; Villani, R.; Marino, C.; Lovisolo, G.A.; Conti, D.; Russo, F.; Freseagna, A.M.; Cordelli, E. Absence of genotoxicity in human blood cells exposed to 50 hz magnetic fields as assessed by comet assay, chromosome aberration, micronucleus, and sister chromatid exchange analyses. *Bioelectromagnetics* **2004**, *25*, 41–48. [CrossRef]
207. Kayhan, H.; Erdebilli, B.; Gonen, S.; Esmekaya, M.A.; Ertekin, E.; Canseven, A.G. Effects of extremely low-frequency magnetic field on healthy fibroblasts and breast cancer cells. *J. Istanb. Fac. Med.* **2020**, *83*, 384–389. [CrossRef]
208. Kim, J.; Ha, C.S.; Lee, H.J.; Song, K. RePETitive exposure to a 60-hz time-varying magnetic field induces DNA double-strand breaks and apoptosis in human cells. *Biochem. Biophys. Res. Commun.* **2010**, *400*, 739–744. [CrossRef] [PubMed]
209. Supino, R.; Bottone, M.G.; Pellicciari, C.; Caserini, C.; Bottiroli, G.; Belleri, M.; Veicsteinas, A. Sinusoidal 50 hz magnetic fields do not affect structural morphology and proliferation of human cells in vitro. *Histol. Histopathol.* **2001**, *16*, 719–726.
210. Savage, R.E.; Kanitz, M.H.; Lotz, W.G.; Conover, D.; Hennessey, E.M.; Hanneman, W.H.; Witzmann, F.A. Changes in gene and protein expression in magnetic field-treated human glioma cells. *Toxicol. Mech. Methods* **2005**, *15*, 115–120. [CrossRef] [PubMed]
211. Kanitz, M.H.; Witzmann, F.A.; Lotz, W.G.; Conover, D.; Savage, R.E. Investigation of protein expression in magnetic field-treated human glioma cells. *Bioelectromagnetics* **2007**, *28*, 546–552. [CrossRef] [PubMed]
212. Yoshizawa, H.; Tsuchiya, T.; Mizoe, H.; Ozeki, H.; Kanao, S.; Yomori, H.; Sakane, C.; Hasebe, S.; Motomura, T.; Yamakawa, T.; et al. No effect of extremely low-frequency magnetic field observed on cell growth or initial response of cell proliferation in human cancer cell lines. *Bioelectromagnetics* **2002**, *23*, 355–368. [CrossRef] [PubMed]
213. Calcabrini, C.; Mancini, U.; De Bellis, R.; Diaz, A.R.; Martinelli, M.; Cucchiari, L.; Sestili, P.; Stocchi, V.; Potenza, L. Effect of extremely low-frequency electromagnetic fields on antioxidant activity in the human keratinocyte cell line nctc 2544. *Biotechnol. Appl. Biochem.* **2017**, *64*, 415–422. [CrossRef]
214. Kim, K.; Lee, Y.S.; Kim, N.; Choi, H.D.; Kang, D.J.; Kim, H.R.; Lim, K.M. Effects of electromagnetic waves with LTE and 5g bandwidth on the skin pigmentation in vitro. *Int. J. Mol. Sci.* **2021**, *22*, 170. [CrossRef]
215. Jandova, A.; Pokorny, J.; Cocek, A.; Trojan, S.; Nedbalova, M.; Dohnalova, A. Effects of sinusoidal 0.5 mt magnetic field on leukocyte adherence inhibition. *Electromagn. Biol. Med.* **2004**, *23*, 81–96.
216. Jandova, A.; Hurich, J.; Pokorny, J.; Cocek, A.; Trojan, S.; Nedbalova, M.; Dohnalova, A. Effects of sinusoidal magnetic field on adherence inhibition of leukocytes. *Electro-Magn.* **2001**, *20*, 397–413. [CrossRef]
217. Kaszuba-Zwoinska, J.; Zdzilowska, E.; Chorobik, P.; Slodowska-Hajduk, Z.; Juszczak, K.; Zaraska, W.; Thor, P.J. Pulsing electromagnetic field and death of proliferating peripheral blood mononuclear cells from patients with acute myelogenous leukemia. *Adv. Clin. Exp. Med.* **2011**, *20*, 721–727.
218. Mangiacasale, R.; Tritarelli, A.; Sciamanna, I.; Cannone, M.; Lavia, P.; Barberis, M.C.; Lorenzini, R.; Cundari, E. Normal and cancer-prone human cells respond differently to extremely low frequency magnetic fields. *FEBS Lett.* **2001**, *487*, 397–403. [CrossRef]

219. Kaszuba-Zwoinska, J.; Wojcik, K.; Bereta, M.; Ziomer, A.; Pierzchalski, P.; Rokita, E.; Marcinkiewicz, J.; Zaraska, W.; Thor, P. Pulsating electromagnetic field stimulation prevents cell death of puromycin treated u937 cell line. *J. Physiol. Pharmacol.* **2010**, *61*, 201–205. [[PubMed](#)]
220. Hirose, H.; Nakahara, T.; Zhang, Q.M.; Yonei, S.; Miyakoshi, J. Static magnetic field with a strong magnetic field gradient (41.7 t/m) induces c-jun expression in hl-60 cells. *In Vitro Cell. Dev. Biol.-Anim.* **2003**, *39*, 348–352. [[CrossRef](#)]
221. Golbach, L.A.; Philippi, J.G.M.; Cuppen, J.J.M.; Savelkoul, H.F.J.; Verburg-van Kemenade, B.M.L. Calcium signalling in human neutrophil cell lines is not affected by low-frequency electromagnetic fields. *Bioelectromagnetics* **2015**, *36*, 430–443. [[CrossRef](#)] [[PubMed](#)]
222. Yan, J.H.; Dong, L.A.; Zhang, B.H.; Qi, N.M. Effects of extremely low-frequency magnetic field on growth and differentiation of human mesenchymal stem cells. *Electromagn. Biol. Med.* **2010**, *29*, 165–176. [[CrossRef](#)] [[PubMed](#)]
223. Santini, M.T.; Rainaldi, G.; Ferrante, A.; Indovina, P.L.; Vecchia, P.; Donelli, G. Effects of a 50 hz sinusoidal magnetic field on cell adhesion molecule expression in two human osteosarcoma cell lines (mg-63 and saos-2). *Bioelectromagnetics* **2003**, *24*, 327–338. [[CrossRef](#)]
224. Saunders, R. Static magnetic fields: Animal studies. *Prog. Biophys. Mol. Biol.* **2005**, *87*, 225–239. [[CrossRef](#)]
225. Mild, K.H.; Mattsson, M.O. Elf noise fields: A review. *Electromagn. Biol. Med.* **2010**, *29*, 72–97. [[CrossRef](#)]
226. Marino, A.A.; Wolcott, R.M.; Chervenak, R.; Jourd'heuil, F.; Nilsen, E.; Frilot, C. Nonlinear dynamical law governs magnetic field induced changes in lymphoid phenotype. *Bioelectromagnetics* **2001**, *22*, 529–546. [[CrossRef](#)] [[PubMed](#)]
227. Anderson, L.E.; Morris, J.E.; Sasser, L.B.; Loscher, W. Effects of 50-or 60-hertz, 100 mu t magnetic field exposure in the dmbo mammary cancer model in sprague-dawley rats: Possible explanations for different results from two laboratories. *Environ. Health Perspect.* **2000**, *108*, 797–802.
228. Maruvada, P.S.; Harvey, S.M.; Jutras, P.; Goulet, D.; Mandeville, R. A magnetic field exposure facility for evaluation of animal carcinogenicity. *Bioelectromagnetics* **2000**, *21*, 432–438. [[CrossRef](#)]
229. Loscher, W. Do cocarcinogenic effects of elf electromagnetic fields require repeated long-term interaction with carcinogens? Characteristics of positive studies using the dmbo breast cancer model in rats. *Bioelectromagnetics* **2001**, *22*, 603–614. [[CrossRef](#)] [[PubMed](#)]
230. Fedrowitz, M.; Kamino, K.; Loscher, W. Significant differences in the effects of magnetic field exposure on 7,12-dimethylbenz(a)anthracene-induced mammary carcinogenesis in two substrains of sprague-dawley rats. *Cancer Res.* **2004**, *64*, 243–251. [[CrossRef](#)] [[PubMed](#)]
231. Galloni, P.; Marino, C. Effects of 50 hz magnetic field exposure on tumor experimental models. *Bioelectromagnetics* **2000**, *21*, 608–614. [[CrossRef](#)]
232. McLean, J.R.; Thansandote, A.; McNamee, J.P.; Tryphonas, L.; Lecuyer, D.; Gajda, G. A 60 hz magnetic field does not affect the incidence of squamous cell carcinomas in sencar mice. *Bioelectromagnetics* **2003**, *24*, 75–81. [[CrossRef](#)]
233. McNamee, J.P.; Bellier, P.V.; McLean, J.R.N.; Marro, L.; Gajda, G.B.; Thansandote, A. DNA damage and apoptosis in the immature mouse cerebellum after acute exposure to a 1 mt, 60 hz magnetic field. *Mutat. Res. Genet. Toxicol. Environ. Mutagenesis* **2002**, *513*, 121–133. [[CrossRef](#)]
234. McNamee, J.P.; Bellier, P.V.; Chauhan, V.; Gajda, G.B.; Lemay, E.; Thansandote, A. Evaluating DNA damage in rodent brain after acute 60 hz magnetic-field exposure. *Radiat. Res.* **2005**, *164*, 791–797. [[CrossRef](#)]
235. Luo, X.; Jia, S.J.; Li, R.Y.; Gao, P.; Zhang, Y.W. Occupational exposure to 50 hz magnetic fields does not alter responses of inflammatory genes and activation of splenic lymphocytes in mice. *Int. J. Occup. Med. Environ. Health* **2016**, *29*, 277–291. [[CrossRef](#)]
236. Sommer, A.M.; Lerchl, A. 50 hz magnetic fields of 1 mt do not promote lymphoma development in AKR/J mice. *Radiat. Res.* **2006**, *165*, 343–349. [[CrossRef](#)]
237. Tatarov, I.; Panda, A.; Petkov, D.; Kolappaswamy, K.; Thompson, K.; Kavirayani, A.; Lipsky, M.M.; Elson, E.; Davis, C.C.; Martin, S.S.; et al. Effect of magnetic fields on tumor growth and viability. *Comp. Med.* **2011**, *61*, 339–345.
238. Vallejo, D.; Hidalgo, M.A.; Hernandez, J.M. Effects of long-term exposure to an extremely low frequency magnetic field (15 microt) on selected blood coagulation variables in of1 mice. *Electromagn. Biol. Med.* **2019**, *38*, 279–286. [[CrossRef](#)] [[PubMed](#)]
239. Cabrales, L.B.; Ciria, H.C.; Bruzon, R.P.; Quevedo, M.S.; Cespedes, M.C.; Salas, M.F. Elf magnetic field effects on some hematological and biochemical parameters of peripheral blood in mice. *Electro- Magn.* **2001**, *20*, 185–191. [[CrossRef](#)]
240. Qi, G.Y.; Zuo, X.X.; Zhou, L.H.; Aoki, E.; Okamura, A.; Watanebe, M.; Wang, H.P.; Wu, Q.H.; Lu, H.L.; Tuncel, H.; et al. Effects of extremely low-frequency electromagnetic fields (elf-EMF) exposure on b6c3f1 mice. *Environ. Health Prev. Med.* **2015**, *20*, 287–293. [[CrossRef](#)]
241. Salim, E.I.; Omar, K.M.; Abou-Hattab, H.A.; Abou-Zaid, F.A. Pituitary toxicity but lack of rat colon carcinogenicity of a dc-magnetic field in a medium-term bioassay. *Asian Pac. J. Cancer Prev.* **2008**, *9*, 131–140.
242. Anderson, L.E.; Morris, J.E.; Miller, D.L.; Rafferty, C.N.; Ebi, K.L.; Sasser, L.B. Large granular lymphocytic (lgl) leukemia in rats exposed to intermittent 60 hz magnetic fields. *Bioelectromagnetics* **2001**, *22*, 185–193. [[CrossRef](#)] [[PubMed](#)]
243. Lee, H.J.; Choi, S.Y.; Jang, J.J.; Gimm, Y.M.; Pack, J.K.; Choi, H.D.; Kim, N.; Lee, Y.S. Lack of promotion of mammary, lung and skin tumorigenesis by 20 khz triangular magnetic fields. *Bioelectromagnetics* **2007**, *28*, 446–453. [[CrossRef](#)] [[PubMed](#)]
244. Tuncel, H.; Shimamoto, F.; Cagatay, P.; Kalkan, M.T. Variable e-cadherin expression in a mnu-induced colon tumor model in rats which exposed with 50 hz frequency sinusoidal magnetic field. *Tohoku J. Exp. Med.* **2002**, *198*, 245–249. [[CrossRef](#)]

245. Fedrowitz, M.; Loscher, W. Exposure of fischer 344 rats to a weak power frequency magnetic field facilitates mammary tumorigenesis in the dbma model of breast cancer. *Carcinogenesis* **2008**, *29*, 186–193. [CrossRef]
246. Ushio-Fukai, M.; Ash, D.; Nagarkoti, S.; de Chantemele, E.J.B.; Fulton, D.J.R.; Fukai, T. Interplay between reactive oxygen/reactive nitrogen species and metabolism in vascular biology and disease. *Antioxid. Redox Signal.* **2021**, *34*, 1319–1354. [CrossRef] [PubMed]
247. Sies, H. Oxidative stress: A concept in redox biology and medicine. *Redox Biol.* **2015**, *4*, 180–183. [CrossRef]
248. Azab, A.E.; Ebrahim, S.A. Exposure to electromagnetic fields induces oxidative stress and pathophysiological changes in the cardiovascular system. *J. Appl. Biotechnol. Bioeng.* **2017**, *4*, 96–102. [CrossRef]
249. Schuermann, D.; Mevissen, M. Manmade electromagnetic fields and oxidative stress—biological effects and consequences for health. *Int. J. Mol. Sci.* **2021**, *22*, 3772. [CrossRef] [PubMed]
250. Havas, M. When theory and observation collide: Can non-ionizing radiation cause cancer? *Environ. Pollut.* **2017**, *221*, 501–505. [CrossRef] [PubMed]
251. Hore, P.J.; Ivanov, K.L.; Wasielewski, M.R. Spin chemistry. *J. Chem. Phys.* **2020**, *152*, 120401. [CrossRef]
252. Jones, A.R. Magnetic field effects in proteins. *Mol. Phys.* **2016**, *114*, 1691–1702. [CrossRef]
253. Mattsson, M.O.; Simkó, M. Grouping of experimental conditions as an approach to evaluate effects of extremely low-frequency magnetic fields on oxidative response in *in vitro* studies. *Front. Public Health* **2014**, *2*, 132. [CrossRef]
254. Juutilainen, J.; Herrala, M.; Luukkonen, J.; Naarala, J.; Hore, P.J. Magnetocarcinogenesis: Is there a mechanism for carcinogenic effects of weak magnetic fields? *Proc. Biol. Sci.* **2018**, *285*, 20180590. [CrossRef]
255. Höytö, A.; Herrala, M.; Luukkonen, J.; Juutilainen, J.; Naarala, J. Cellular detection of 50 hz magnetic fields and weak blue light: Effects on superoxide levels and genotoxicity. *Int. J. Radiat. Biol.* **2017**, *93*, 646–652. [CrossRef]
256. Orel, V.E.; Krotevych, M.; Dasyukevich, O.; Rykhalskyi, O.; Syvak, L.; Tsvir, H.; Tsvir, D.; Garmanchuk, L.; Orel Vcapital Ve, C.; Sheina, I.; et al. Effects induced by a 50 hz electromagnetic field and doxorubicin on walker-256 carcinosarcoma growth and hepatic redox state in rats. *Electromagn. Biol. Med.* **2021**, *40*, 475–487. [CrossRef]
257. Mannerling, A.C.; Simko, M.; Mild, K.H.; Mattsson, M.O. Effects of 50-hz magnetic field exposure on superoxide radical anion formation and hsp70 induction in human k562 cells. *Radiat. Environ. Biophys.* **2010**, *49*, 731–741. [CrossRef]
258. Bułdak, R.J.; Polaniak, R.; Bułdak, Ł.; Źwirska-Korczala, K.; Skonieczna, M.; Monsioli, A.; Kukla, M.; Duława-Bułdak, A.; Birkner, E. Short-term exposure to 50 hz emf alters the cisplatin-induced oxidative response in at478 murine squamous cell carcinoma cells. *Bioelectromagnetics* **2012**, *33*, 641–651. [CrossRef] [PubMed]
259. Luukkonen, J.; Liimatainen, A.; Juutilainen, J.; Naarala, J. Induction of genomic instability, oxidative processes, and mitochondrial activity by 50hz magnetic fields in human sh-sy5y neuroblastoma cells. *Mutat. Res./Fundam. Mol. Mech. Mutagenesis* **2014**, *760*, 33–41. [CrossRef]
260. Xu, A.; Wang, Q.; Lin, T. Low-frequency magnetic fields (lf-mfs) inhibit proliferation by triggering apoptosis and altering cell cycle distribution in breast cancer cells. *Int. J. Mol. Sci.* **2020**, *21*, 2952. [CrossRef] [PubMed]
261. Cios, A.; Ciepielak, M.; Stankiewicz, W.; Szymanski, L. The influence of the extremely low frequency electromagnetic field on clear cell renal carcinoma. *Int. J. Mol. Sci.* **2021**, *22*, 1342. [CrossRef] [PubMed]
262. Amara, S.; Abdelmelek, H.; Garrel, C.; Guiraud, P.; Douki, T.; Ravanat, J.L.; Favier, A.; Sakly, M.; Ben Rhouma, K. Zinc supplementation ameliorates static magnetic field-induced oxidative stress in rat tissues. *Environ. Toxicol. Pharmacol.* **2007**, *23*, 193–197. [CrossRef] [PubMed]
263. Amara, S.; Abdelmelek, H.; Garrel, C.; Guiraud, P.; Douki, T.; Ravanat, J.L.; Favier, A.; Sakly, M.; Ben Rhouma, K. Influence of static magnetic field on cadmium toxicity: Study of oxidative stress and DNA damage in rat tissues. *J. Trace Elem. Med. Biol.* **2006**, *20*, 263–269. [CrossRef] [PubMed]
264. Amara, S.; Douki, T.; Garel, C.; Favier, A.; Sakly, M.; Rhouma, K.B.; Abdelmelek, H. Effects of static magnetic field exposure on antioxidative enzymes activity and DNA in rat brain. *Gen. Physiol. Biophys.* **2009**, *28*, 260–265. [CrossRef]
265. Amara, S.; Douki, T.; Ravanat, J.L.; Garrel, C.; Guiraud, P.; Favier, A.; Sakly, M.; Ben Rhouma, K.; Abdelmelek, H. Influence of a static magnetic field (250 mt) on the antioxidant response and DNA integrity in thp1 cells. *Phys. Med. Biol.* **2007**, *52*, 889–898. [CrossRef]
266. Reale, M.; Kamal, M.A.; Patruno, A.; Costantini, E.; D'Angelo, C.; Pesce, M.; Greig, N.H. Neuronal cellular responses to extremely low frequency electromagnetic field exposure: Implications regarding oxidative stress and neurodegeneration. *PLoS ONE* **2014**, *9*, e104973. [CrossRef]
267. Patruno, A.; Tabrez, S.; Pesce, M.; Shakil, S.; Kamal, M.A.; Reale, M. Effects of extremely low frequency electromagnetic field (elf-EMF) on catalase, cytochrome p450 and nitric oxide synthase in erythro-leukemic cells. *Life Sci.* **2015**, *121*, 117–123. [CrossRef]
268. Lee, B.C.; Johng, H.M.; Lim, J.K.; Jeong, J.H.; Baik, K.Y.; Nam, T.J.; Lee, J.H.; Kim, J.; Sohn, U.D.; Yoon, G.; et al. Effects of extremely low frequency magnetic field on the antioxidant defense system in mouse brain: A chemiluminescence study. *J. Photochem. Photobiol. B Biol.* **2004**, *73*, 43–48. [CrossRef] [PubMed]
269. Lewicka, M.; Henrykowska, G.A.; Pacholski, K.; Smigielski, J.; Rutkowski, M.; Dziedziczk-Buczynska, M.; Buczynski, A. The effect of electromagnetic radiation emitted by display screens on cell oxygen metabolism—In vitro studies. *Arch. Med. Sci.* **2015**, *11*, 1330–1339. [CrossRef] [PubMed]
270. Źwirska-Korczala, K.; Adamczyk-Sowa, M.; Polaniak, R.; Sowa, P.; Birkner, E.; Drzazga, Z.; Brzozowski, T.; Konturek, S.J. Influence of extremely-low-frequency magnetic field on antioxidative melatonin properties in at478 murine squamous cell carcinoma culture. *Biol. Trace Elem. Res.* **2004**, *102*, 227–243. [CrossRef]

271. Zhao, B.; Yu, T.; Wang, S.; Che, J.; Zhou, L.; Shang, P. Static magnetic field (0.2–0.4 t) stimulates the self-renewal ability of osteosarcoma stem cells through autophagic degradation of ferritin. *Bioelectromagnetics* **2021**, *42*, 371–383. [CrossRef]
272. Kesari, K.K.; Juutilainen, J.; Luukkonen, J.; Naarala, J. Induction of micronuclei and superoxide production in neuroblastoma and glioma cell lines exposed to weak 50 hz magnetic fields. *J. R. Soc. Interface* **2016**, *13*, 20150995. [CrossRef]
273. Erdal, N.; Gürgül, S.; Tamer, L.; Ayaz, L. Effects of long-term exposure of extremely low frequency magnetic field on oxidative/nitrosative stress in rat liver. *J. Radiat. Res.* **2008**, *49*, 181–187. [CrossRef]
274. Koh, E.K.; Ryu, B.K.; Jeong, D.Y.; Bang, I.S.; Nam, M.H.; Chae, K.S. A 60-hz sinusoidal magnetic field induces apoptosis of prostate cancer cells through reactive oxygen species. *Int. J. Radiat. Biol.* **2008**, *84*, 945–955. [CrossRef]
275. Kahya, M.C.; Naziroğlu, M.; Çığ, B. Selenium reduces mobile phone (900 mhz)-induced oxidative stress, mitochondrial function, and apoptosis in breast cancer cells. *Biol. Trace Elem. Res.* **2014**, *160*, 285–293. [CrossRef]
276. Polaniak, R.; Buldak, R.J.; Karon, M.; Birkner, K.; Kukla, M.; Zwirska-Korczala, K.; Birkner, E. Influence of an extremely low frequency magnetic field (elf-EMF) on antioxidative vitamin e properties in at478 murine squamous cell carcinoma culture in vitro. *Int. J. Toxicol.* **2010**, *29*, 221–230. [CrossRef]
277. Bekhite, M.M.; Finkensieper, A.; Abou-Zaid, F.A.; El-Shourbagy, I.K.; El-Fiky, N.K.; Omar, K.M.; Sauer, H.; Wartenberg, M. Differential effects of high and low strength magnetic fields on mouse embryonic development and vasculogenesis of embryonic stem cells. *Reprod. Toxicol.* **2016**, *65*, 46–58. [CrossRef]
278. Kang, S.K. Measuring the value of MRI: Comparative effectiveness & outcomes research. *J. Magn. Reson. Imaging* **2019**, *49*, e78–e84. [PubMed]
279. van der Heide, U.A.; Frantzen-Steneker, M.; Astreinidou, E.; Nowee, M.E.; van Houdt, P.J. MRI basics for radiation oncologists. *Clin. Transl. Radiat. Oncol.* **2019**, *18*, 74–79. [CrossRef] [PubMed]
280. Raaijmakers, A.J.E.; Raaymakers, B.W.; Lagendijk, J.J.W. Magnetic-field-induced dose effects in mr-guided radiotherapy systems: Dependence on the magnetic field strength. *Phys. Med. Biol.* **2008**, *53*, 909–923. [CrossRef] [PubMed]
281. Morsing, A.; Hildebrandt, M.G.; Vilstrup, M.H.; Wallenius, S.E.; Gerke, O.; Petersen, H.; Johansen, A.; Andersen, T.L.; Hoilund-Carlsen, P.F. Hybrid PET/MRI in major cancers: A scoping review. *Eur. J. Nucl. Med. Mol. Imaging* **2019**, *46*, 2138–2151. [CrossRef]
282. Scheenen, T.W.J.; Zamecnik, P. The role of magnetic resonance imaging in (future) cancer staging note the nodes. *Investig. Radiol.* **2021**, *56*, 42–49. [CrossRef]
283. Avasthi, A.; Caro, C.; Pozo-Torres, E.; Leal, M.P.; Garcia-Martin, M.L. Magnetic nanoparticles as MRI contrast agents. *Top. Curr. Chem.* **2020**, *378*, 43.
284. Poku, L.O.; Cheng, Y.N.; Wang, K.; Sun, X.L. Na-23-MRI as a noninvasive biomarker for cancer diagnosis and prognosis. *J. Magn. Reson. Imaging* **2021**, *53*, 995–1014. [CrossRef]
285. Visser, M.; Muller, D.M.J.; van Duijn, J.M.; Smits, M.; Verburg, N.; Hendriks, E.J.; Nabuurs, R.J.A.; Bot, J.C.J.; Eijgelaar, R.S.; Witte, M.; et al. Inter-rater agreement in glioma segmentations on longitudinal MRI. *NeuroImage Clin.* **2019**, *22*, 101727. [CrossRef]
286. Patil, R.; Ljubimov, A.V.; Gangalum, P.R.; Ding, H.; Portilla-Arias, J.; Wagner, S.; Inoue, S.; Konda, B.; Rekechenetskiy, A.; Chesnokova, A.; et al. MRI virtual biopsy and treatment of brain metastatic tumors with targeted nanobioconjugates: Nanoclinic in the brain. *ACS Nano* **2015**, *9*, 5594–5608. [CrossRef]
287. Lasocki, A.; Anjari, M.; Kokurcan, S.O.; Thust, S.C. Conventional MRI features of adult diffuse glioma molecular subtypes: A systematic review. *Neuroradiology* **2021**, *63*, 353–362. [CrossRef]
288. Machhale, K.; Nandpuru, H.B.; Kapur, V.; Kosta, L. MRI brain cancer classification using hybrid classifier (SVM-KNN). In Proceedings of the 2015 International Conference on Industrial Instrumentation and Control (ICIC), Pune, India, 28–30 May 2015; pp. 60–65.
289. Nandpuru, H.B.; Salankar, S.S.; Bora, V.R. MRI brain cancer classification using support vector machine. In Proceedings of the 2014 IEEE Students' Conference on Electrical, Electronics and Computer Science, Bhopal, India, 1–2 March 2014.
290. Naser, M.A.; Deen, M.J. Brain tumor segmentation and grading of lower-grade glioma using deep learning in MRI images. *Comput. Biol. Med.* **2020**, *121*, 8. [CrossRef] [PubMed]
291. Clement, P.; Booth, T.; Borovecki, F.; Emblem, K.E.; Figueiredo, P.; Hirschler, L.; Jancalek, R.; Keil, V.C.; Maumet, C.; Ozsunar, Y.; et al. Glimr: Cross-border collaborations to promote advanced MRI biomarkers for glioma. *J. Med. Biol. Eng.* **2021**, *41*, 115–125. [CrossRef] [PubMed]
292. Zhuge, Y.; Ning, H.; Mathen, P.; Cheng, J.Y.; Krauze, A.V.; Camphausen, K.; Miller, R.W. Automated glioma grading on conventional MRI images using deep convolutional neural networks. *Med. Phys.* **2020**, *47*, 3044–3053. [CrossRef] [PubMed]
293. Wietelmann, D.; Schumacher, M.; Muendel, J. Brain-stem glioma. *Radiologe* **1998**, *38*, 904–912. [CrossRef] [PubMed]
294. Porcari, P.; Hegi, M.E.; Lei, H.; Hamou, M.F.; Vassallo, I.; Capuani, S.; Gruetter, R.; Mlynarik, V. Early detection of human glioma sphere xenografts in mouse brain using diffusion MRI at 14.1t. *NMR Biomed.* **2016**, *29*, 1577–1589. [CrossRef] [PubMed]
295. Rogers, C.M.; Jones, P.S.; Weinberg, J.S. Intraoperative MRI for brain tumors. *J. Neuro-Oncol.* **2021**, *151*, 479–490. [CrossRef] [PubMed]
296. Poussaint, T.Y.; Kocak, M.; Vajapeyam, S.; Packer, R.I.; Robertson, R.L.; Geyer, R.; Haas-Kogan, D.; Pollack, I.F.; Vezina, G.; Zimmerman, R.; et al. MRI as a central component of clinical trials analysis in brainstem glioma: A report from the pediatric brain tumor consortium (pbtc). *Neuro-Oncology* **2011**, *13*, 417–427. [CrossRef]

297. Cabaj, A.; Bekiesinska-Figatowska, M.; Duczkowska, A.; Duczkowski, M. Brain MRI findings in neurological complications of cancer treatment. *Adv. Clin. Exp. Med.* **2016**, *25*, 789–797. [CrossRef] [PubMed]
298. Kjaer, A.; Loft, A.; Law, I.; Berthelsen, A.K.; Borgwardt, L.; Lofgren, J.; Johnbeck, C.B.; Hansen, A.E.; Keller, S.; Holm, S.; et al. PET/MRI in cancer patients: First experiences and vision from copenhagen. *Magn. Reson. Mat. Phys. Biol. Med.* **2013**, *26*, 37–47. [CrossRef]
299. Tsioris, S.; Bougias, C.; Fotopoulos, A. Principles and current trends in the correlative evaluation of glioma with advanced MRI techniques and PET. *Hell. J. Nucl. Med.* **2019**, *22*, 206–219.
300. Puttick, S.; Bell, C.; Dowson, N.; Rose, S.; Fay, M. PET, MRI, and simultaneous PET/MRI in the development of diagnostic and therapeutic strategies for glioma. *Drug Discov. Today* **2015**, *20*, 306–317. [CrossRef] [PubMed]
301. Shankar, A.; Bomanji, J.; Hyare, H. Hybrid PET-MRI imaging in paediatric and TYA brain tumours: Clinical applications and challenges. *J. Pers. Med.* **2020**, *10*, 218. [CrossRef] [PubMed]
302. Deuschl, C.; Kirchner, J.; Poeppel, T.D.; Schaarschmidt, B.; Kebir, S.; El Hindy, N.; Hense, J.; Quick, H.H.; Glas, M.; Herrmann, K.; et al. C-11-met PET/MRI for detection of recurrent glioma. *Eur. J. Nucl. Med. Mol. Imaging* **2018**, *45*, 593–601. [CrossRef] [PubMed]
303. Kebir, S.; Weber, M.; Lazaridis, L.; Deuschl, C.; Schmidt, T.; Monninghoff, C.; Keyvani, K.; Umutlu, L.; Pierscianek, D.; Forsting, M.; et al. Hybrid c-11-met PET/MRI combined with “machine learning” in glioma diagnosis according to the revised glioma WHO classification 2016. *Clin. Nucl. Med.* **2019**, *44*, 214–220. [CrossRef]
304. Suh, C.H.; Kim, H.S.; Jung, S.C.; Choi, C.G.; Kim, S.J. Perfusion MRI as a diagnostic biomarker for differentiating glioma from brain metastasis: A systematic review and meta-analysis. *Eur. Radiol.* **2018**, *28*, 3819–3831. [CrossRef]
305. Xu, X.; Chan, K.W.Y.; Knutsson, L.; Artemov, D.; Xu, J.D.; Liu, G.; Kato, Y.; Lal, B.; Laterra, J.; McMahon, M.T.; et al. Dynamic glucose enhanced (dge) MRI for combined imaging of blood-brain barrier break down and increased blood volume in brain cancer. *Magn. Reson. Med.* **2015**, *74*, 1556–1563. [CrossRef]
306. Davis, S.C.; Samkoe, K.S.; O’Hara, J.A.; Gibbs-Strauss, S.L.; Payne, H.L.; Hoopes, P.J.; Paulsen, K.D.; Pogue, B.W. MRI-coupled fluorescence tomography quantifies EGFR activity in brain tumors. *Acad. Radiol.* **2010**, *17*, 271–276. [CrossRef]
307. Aquino, D.; Gioppo, A.; Finocchiaro, G.; Bruzzone, M.G.; Cuccarini, V. MRI in glioma immunotherapy: Evidence, pitfalls, and perspectives. *J. Immunol. Res.* **2017**, *2017*, 5813951. [CrossRef]
308. Jenkinson, M.D.; Du Plessis, D.G.; Walker, C.; Smith, T.S. Advanced MRI in the management of adult gliomas. *Br. J. Neurosurg.* **2007**, *21*, 550–561. [CrossRef]
309. Jestaedt, L.; Lemke, D.; Weiler, M.; Pfenning, P.N.; Heiland, S.; Wick, W.; Bendszus, M. Gadofluorine m enhanced MRI in experimental glioma: Superior and persistent intracellular tumor enhancement compared with conventional MRI. *J. Magn. Reson. Imaging* **2012**, *35*, 551–560. [CrossRef]
310. Liao, J.C.; Xia, R.; Liu, T.; Feng, H.; Ai, H.; Song, B.; Gao, F.B. In vivo dynamic monitoring of the biological behavior of labeled c6 glioma by MRI. *Mol. Med. Rep.* **2013**, *7*, 1397–1402. [CrossRef] [PubMed]
311. Zaccagna, F.; Riemer, F.; Priest, A.N.; McLean, M.A.; Allinson, K.; Grist, J.T.; Dragos, C.; Matys, T.; Gillard, J.H.; Watts, C.; et al. Non-invasive assessment of glioma microstructure using verdict MRI: Correlation with histology. *Eur. Radiol.* **2019**, *29*, 5559–5566. [CrossRef] [PubMed]
312. Anka, A.; Thompson, P.; Mott, E.; Sharma, R.; Zhang, R.Z.; Cross, N.; Sun, J.Y.; Flask, C.A.; Oleinick, N.L.; Dean, D. Dynamic contrast enhanced-magnetic resonance imaging (DCE-MRI) for the assessment of pc 4-sensitized photodynamic therapy of a u87-derived glioma model in the athymic nude rat. In Proceedings of the Conference on Photonic Therapeutics and Diagnostics VI, San Francisco, CA, USA, 23–25 January 2010; SPIE—Society of Photo-Optical Instrumentation Engineers: San Francisco, CA, USA, 2010.
313. Belle, V.; Anka, A.; Cross, N.; Thompson, P.; Mott, E.; Sharma, R.; Gray, K.; Zhang, R.Z.; Xu, Y.S.; Sun, J.Y.; et al. Dynamic contrast enhanced-magnetic resonance imaging (DCE-MRI) of photodynamic therapy (PDT) outcome and associated changes in the blood-brain barrier following PC 4-PDT of glioma in an athymic nude rat model. In Proceedings of the Conference on Photonic Therapeutics and Diagnostics VIII, San Francisco, CA, USA, 21–24 January 2012; SPIE—Society of Photo-Optical Instrumentation Engineers: San Francisco, CA, USA, 2012.
314. Young, J.R.; Ressler, J.A.; Mortimer, J.E.; Schmolze, D.; Fitzgibbons, M.; Chen, B.H.T. Performance of enhancement on brain MRI for identifying her2 overexpression in breast cancer brain metastases. *Eur. J. Radiol.* **2021**, *144*, 6. [CrossRef]
315. Papadimitrakopoulou, V.A. Carcinogenesis of head and neck cancer and the role of chemoprevention in its reversal. *Curr. Opin. Oncol.* **2000**, *12*, 240–245. [CrossRef]
316. Loeffelbein, D.J.; Souvatzaglou, M.; Wankerl, V.; Dinges, J.; Ritschl, L.M.; Mucke, T.; Pickhard, A.; Eiber, M.; Schwaiger, M.; Beer, A.J. Diagnostic value of retrospective PET-MRI fusion in head-and-neck cancer. *BMC Cancer* **2014**, *14*, 10. [CrossRef] [PubMed]
317. Wang, K.; Mullins, B.T.; Falchook, A.D.; Lian, J.; He, K.L.; Shen, D.G.; Dance, M.; Lin, W.L.; Sills, T.M.; Das, S.K.; et al. Evaluation of PET/MRI for tumor volume delineation for head and neck cancer. *Front. Oncol.* **2017**, *7*, 8. [CrossRef] [PubMed]
318. Grosse, J.; Hellwig, D. Pet/CT and PET/MRI in head and neck cancer. *Laryngorhinootologie* **2020**, *99*, 12–21.
319. Hermans, R. Diffusion-weighted MRI in head and neck cancer. *Curr. Opin. Otolaryngol. Head Neck Surg.* **2010**, *18*, 72–78. [CrossRef]
320. Thoeny, H.C. Diffusion-weighted MRI in head and neck radiology: Applications in oncology. *Cancer Imaging* **2011**, *10*, 209–214. [CrossRef]

321. Schakel, T.; Hoogduin, J.M.; Terhaard, C.H.J.; Philippens, M.E.P. Diffusion weighted MRI in head-and-neck cancer: Geometrical accuracy. *Radiother. Oncol.* **2013**, *109*, 394–397. [CrossRef]
322. El Beltagi, A.H.; Elsotouhy, A.H.; Own, A.M.; Abdelfattah, W.; Nair, K.; Vattoth, S. Functional magnetic resonance imaging of head and neck cancer: Performance and potential. *Neuradiol. J.* **2019**, *32*, 36–52. [CrossRef] [PubMed]
323. Zhao, B.X.; Soraghan, J.; Di-caterina, G.; Grose, D.; Doshi, T. Automatic 3D detection and segmentation of head and neck cancer from MRI data. In Proceedings of the 2018 7th European Workshop on Visual Information Processing (EUVIP), Tampere, Finland, 26–28 November 2018.
324. Chen, A.M.; Hsu, S.; Lamb, J.; Yang, Y.; Agazaryan, N.; Steinberg, M.L.; Low, D.A.; Cao, M. MRI-guided radiotherapy for head and neck cancer: Initial clinical experience. *Clin. Transl. Oncol.* **2018**, *20*, 160–168. [CrossRef] [PubMed]
325. Baulch, J.; Gandhi, M.; Sommerville, J.; Panizza, B. 3t MRI evaluation of large nerve perineural spread of head and neck cancers. *J. Med. Imaging Radiat. Oncol.* **2015**, *59*, 578–585. [CrossRef] [PubMed]
326. Kanda, T.; Kitajima, K.; Suenaga, Y.; Konishi, J.; Sasaki, R.; Morimoto, K.; Saito, M.; Otsuki, N.; Nibu, K.; Sugimura, K. Value of retrospective image fusion of ^{18}F -FDG PET and MRI for preoperative staging of head and neck cancer: Comparison with PET/CT and contrast-enhanced neck MRI. *Eur. J. Radiol.* **2013**, *82*, 2005–2010. [CrossRef] [PubMed]
327. Baghi, M.; Mack, M.G.; Hambek, M.; Rieger, J.; Vogl, T.; Gstoettner, W.; Knecht, R. The efficacy of MRI with ultrasmall superparamagnetic iron oxide particles (uspio) in head and neck cancers. *Anticancer Res.* **2005**, *25*, 3665–3670. [PubMed]
328. Rahbari, R.; Zhang, L.S.; Kebebew, E. Thyroid cancer gender disparity. *Future Oncol.* **2010**, *6*, 1771–1779. [CrossRef] [PubMed]
329. Dinauer, C.A.; Breuer, C.; Rivkees, S.A. Differentiated thyroid cancer in children: Diagnosis and management. *Curr. Opin. Oncol.* **2008**, *20*, 59–65. [CrossRef]
330. Massimino, M.; Evans, D.B.; Podda, M.; Spinelli, C.; Collini, P.; Pizzi, N.; Bleyer, A. Thyroid cancer in adolescents and young adults. *Pediatr. Blood Cancer* **2018**, *65*, 9. [CrossRef]
331. Regalbuto, C.; Frasca, F.; Pellegriti, G.; Malandrino, P.; Marturano, I.; Di Carlo, I.; Pezzino, V. Update on thyroid cancer treatment. *Future Oncol.* **2012**, *8*, 1331–1348. [CrossRef]
332. CynaGorse, F.; Toubert, M.E.; Zagdanski, A.M.; deKerviler, E.; Feger, C.; Benchaib, N.; Attal, P.; Frija, J.; LavalJeantet, M. Recurrence of differentiated thyroid carcinomas: Value of MRI. *J. Radiol.* **1996**, *77*, 1195–1200.
333. Chen, Q.H.; Raghavan, P.; Mukherjee, S.; Jameson, M.J.; Patrie, J.; Xin, W.J.; Xian, J.F.; Wang, Z.C.; Levine, P.A.; Wintermark, M. Accuracy of MRI for the diagnosis of metastatic cervical lymphadenopathy in patients with thyroid cancer. *Radiol. Med.* **2015**, *120*, 959–966. [CrossRef] [PubMed]
334. Gross, N.D.; Weissman, J.L.; Talbot, J.M.; Andersen, P.E.; Wax, M.K.; Cohen, J.I. MRI detection of cervical metastasis from differentiated thyroid carcinoma. *Laryngoscope* **2001**, *111*, 1905–1909. [CrossRef] [PubMed]
335. Liu, Z.M.; Xun, X.Q.; Wang, Y.Z.; Mei, L.; He, L.; Zeng, W.; Wang, C.Y.; Tao, H. MRI and ultrasonography detection of cervical lymph node metastases in differentiated thyroid carcinoma before reoperation. *Am. J. Transl. Res.* **2014**, *6*, 147–154. [PubMed]
336. Samanci, C.; Onal, Y.; Sager, S.; Asa, S.; Ustabasioglu, F.E.; Alis, D.; Akman, C.; Sonmezoglu, K. Diagnostic capabilities of MRI versus ^{18}F FDG PET-ct in postoperative patients with thyroglobulin positive, ^{131}I -negative local recurrent or metastatic thyroid cancer. *Curr. Med. Imaging* **2019**, *15*, 956–964. [CrossRef]
337. Wang, J.C.; Takashima, S.; Matsushita, T.; Takayama, F.; Kobayashi, T.; Kadoya, M. Esophageal invasion by thyroid carcinomas: Prediction using magnetic resonance imaging. *J. Comput. Assist. Tomogr.* **2003**, *27*, 18–25. [CrossRef]
338. Brown, A.M.; Nagala, S.; McLean, M.A.; Lu, Y.G.; Scoffings, D.; Apte, A.; Gonen, M.; Stambuk, H.E.; Shah, A.R.; Tuttle, R.M.; et al. Multi-institutional validation of a novel textural analysis tool for preoperative stratification of suspected thyroid tumors on diffusion-weighted MRI. *Magn. Reson. Med.* **2016**, *75*, 1708–1716. [CrossRef]
339. Naglah, A.; Khalifa, F.; Khaled, R.; Razek, A.; Ghazal, M.; Giridharan, G.; El-Baz, A. Novel MRI-based cad system for early detection of thyroid cancer using multi-input CNN. *Sensors* **2021**, *21*, 3878. [CrossRef]
340. Abd el Aziz, L.M.; Hamisa, M.; Badwy, M.E. Differentiation of thyroid nodules using diffusion-weighted MRI. *Alex. J. Med.* **2015**, *51*, 305–309. [CrossRef]
341. Taha, M.S.; Hassan, O.; Amir, M.; Taha, T.; Riad, M.A. Diffusion-weighted MRI in diagnosing thyroid cartilage invasion in laryngeal carcinoma. *Eur. Arch. Oto-Rhino-Laryngol.* **2014**, *271*, 2511–2516. [CrossRef]
342. Binse, I.; Poeppel, T.D.; Ruhlmann, M.; Gomez, B.; Umutlu, L.; Bockisch, A.; Rosenbaum-Krumme, S.J. Imaging with i-124 in differentiated thyroid carcinoma: Is PET/MRI superior to PET/CT? *Eur. J. Nucl. Med. Mol. Imaging* **2016**, *43*, 1011–1017. [CrossRef]
343. Seiboth, L.; Van Nostrand, D.; Wartofsky, L.; Ousman, Y.; Jonklaas, J.; Butler, C.; Atkins, F.; Burman, K. Utility of PET/neck MRI digital fusion images in the management of recurrent or persistent thyroid cancer. *Thyroid* **2008**, *18*, 103–111. [CrossRef] [PubMed]
344. Teller, P.; Jefford, V.J.; Gabram, S.G.A.; Newell, M.; Carlson, G.W. The utility of breast MRI in the management of breast cancer. *Breast J.* **2010**, *16*, 394–403. [CrossRef] [PubMed]
345. Mann, R.M.; Cho, N.; Moy, L. Breast MRI: State of the art. *Radiology* **2019**, *292*, 520–536. [CrossRef] [PubMed]
346. Militello, C.; Rundo, L.; Dimarco, M.; Orlando, A.; Conti, V.; Woitek, R.; D’Angelo, I.; Bartolotta, T.V.; Russo, G. Semi-automated and interactive segmentation of contrast-enhancing masses on breast DCE-MRI using spatial fuzzy clustering. *Biomed. Signal Process. Control* **2022**, *71*, 12. [CrossRef]
347. Vogel, W.V.; Nestle, U.; Valli, M.C. Pet/MRI in breast cancer. *Clin. Transl. Imaging* **2017**, *5*, 71–78. [CrossRef]

348. Heacock, L.; Reig, B.; Lewin, A.A.; Toth, H.K.; Moy, L.; Lee, C.S. Abbreviated breast MRI: Road to clinical implementation. *J. Breast Imaging* **2020**, *2*, 201–214. [CrossRef]
349. Mann, R.M.; Kuhl, C.K.; Moy, L. Contrast-enhanced MRI for breast cancer screening. *J. Magn. Reson. Imaging* **2019**, *50*, 377–390. [CrossRef]
350. van Zandwijk, N. Chemoprevention of lung cancer. *Lung Cancer* **2001**, *34*, S91–S94. [CrossRef]
351. Hintze, C.; Dinkel, J.; Biederer, J.; Heussel, C.P.; Puderbach, M. New procedures—Comprehensive staging of lung cancer with MRI. *Radiologe* **2010**, *50*, 699–705. [CrossRef]
352. Khalil, A.; Bouhela, T.; Carette, M.F. Contribution of MRI in lung cancer staging. *JBR-BTR* **2013**, *96*, 132–141. [CrossRef]
353. Biederer, J.; Ohno, Y.; Hatabu, H.; Schiebler, M.L.; van Beek, E.J.R.; Vogel-Claussen, J.; Kauczor, H.U. Screening for lung cancer: Does MRI have a role? *Eur. J. Radiol.* **2017**, *86*, 353–360. [CrossRef]
354. Meier-Schroers, M.; Homsi, R.; Schild, H.H.; Thomas, D. Lung cancer screening with MRI: Characterization of nodules with different non-enhanced MRI sequences. *Acta Radiol.* **2019**, *60*, 168–176. [CrossRef]
355. Meier-Schroers, M.; Homsi, R.; Skowasch, D.; Buermann, J.; Zipfel, M.; Schild, H.H.; Thomas, D. Lung cancer screening with MRI: Results of the first screening round. *J. Cancer Res. Clin. Oncol.* **2018**, *144*, 117–125. [CrossRef]
356. Cobben, D.C.P.; de Boer, H.C.J.; Tijssen, R.H.; Rutten, E.; van Vulpen, M.; Peerlings, J.; Troost, E.G.C.; Hoffmann, A.L.; van Lier, A. Emerging role of MRI for radiation treatment planning in lung cancer. *Technol. Cancer Res. Treat.* **2016**, *15*, NP47–NP60. [CrossRef]
357. Weller, A.; Papoutsaki, M.V.; Waterton, J.C.; Chiti, A.; Stroobants, S.; Kuijer, J.; Blackledge, M.; Morgan, V.; deSouza, N.M. Diffusion-weighted (dw) MRI in lung cancers: Adc test-retest repeatability. *Eur. Radiol.* **2017**, *27*, 4552–4562. [CrossRef]
358. Kim, H.S.; Lee, K.S.; Ohno, Y.; van Beek, E.J.R.; Biederer, J.; Int Workshop Pulm, F. PET/CT versus MRI for diagnosis, staging, and follow-up of lung cancer. *J. Magn. Reson. Imaging* **2015**, *42*, 247–260. [CrossRef]
359. Ohno, Y.; Koyama, H.; Lee, H.Y.; Yoshikawa, T.; Sugimura, K. Magnetic resonance imaging (MRI) and positron emission tomography (PET)/MRI for lung cancer staging. *J. Thorac. Imaging* **2016**, *31*, 215–227. [CrossRef]
360. Pasechnikov, V.; Chukov, S.; Fedorov, E.; Kikuste, I.; Leja, M. Gastric cancer: Prevention, screening and early diagnosis. *World J. Gastroenterol.* **2014**, *20*, 13842–13862. [CrossRef]
361. Renzulli, M.; Clemente, A.; Spinelli, D.; Ierardi, A.M.; Marasco, G.; Farina, D.; Brocchi, S.; Ravaioli, M.; Pettinari, I.; Cescon, M.; et al. Gastric cancer staging: Is it time for magnetic resonance imaging? *Cancers* **2020**, *12*, 1402. [CrossRef]
362. Zhang, Y.J.; Yu, J.C. The role of MRI in the diagnosis and treatment of gastric cancer. *Diagn. Interv. Radiol.* **2020**, *26*, 176–182. [CrossRef]
363. De Vuysere, S.; Vandecaveye, V.; De Bruecker, Y.; Carton, S.; Vermeiren, K.; Tollens, T.; De Keyzer, F.; Dresen, R.C. Accuracy of whole-body diffusion-weighted MRI (wb-dwi/MRI) in diagnosis, staging and follow-up of gastric cancer, in comparison to CT: A pilot study. *BMC Med. Imaging* **2021**, *21*, 9. [CrossRef]
364. Caivano, R.; Rabasco, P.; Lotumolo, A.; D'Antuono, F.; Zandolino, A.; Villonio, A.; Macarini, L.; Guglielmi, G.; Salvatore, M.; Cammarota, A. Gastric cancer: The role of diffusion weighted imaging in the preoperative staging. *Cancer Investig.* **2014**, *32*, 184–190. [CrossRef]
365. Hasbahceci, M.; Akcakaya, A.; Memmi, N.; Turkmen, I.; Cipe, G.; Yildiz, P.; Arici, D.S.; Muslumanoglu, M. Diffusion MRI on lymph node staging of gastric adenocarcinoma. *Quant. Imaging Med. Surg.* **2015**, *5*, 392–400.
366. Takahashi, H.; Yano, H.; Matsushita, M.; Monden, T.; Kinuta, M.; Tateishi, H.; Nakano, Y.; Matsui, S.; Iwazawa, T.; Kanoh, T.; et al. Preoperative staging of gastric cancer: Diagnosing the depth of invasion in the gastric wall with MRI. In Proceedings of the 3rd International Gastric Cancer Congress, Seoul, Korea, 27–30 April 1999; pp. 411–414.
367. Tokuhara, T.; Tanigawa, N.; Matsuki, M.; Nomura, E.; Mabuchi, H.; Lee, S.W.; Tatsumi, Y.; Nishimura, H.; Yoshinaka, R.; Kurisu, Y.; et al. Evaluation of lymph node metastases in gastric cancer using magnetic resonance imaging with ultrasmall superparamagnetic iron oxide (uspio): Diagnostic performance in post-contrast images using new diagnostic criteria. *Gastric Cancer* **2008**, *11*, 194–200. [CrossRef]
368. Arocena, M.G.; Barturen, A.; Bujanda, L.; Casado, O.; Ramirez, M.M.; Oleagoitia, J.M.; Iturri, M.G.; Mugica, P.; Cosme, A.; Gutierrez-Stampa, M.A.; et al. MRI and endoscopic ultrasonography in the staging of gastric cancer. *Rev. Esp. Enferm. Dig.* **2006**, *98*, 582–586. [CrossRef]
369. Malaj, A.; Bilaj, F.; Shahini, A.; Miraka, M. Ct/MRI accuracy in detecting and determining preoperative stage of gastric adenocarcinoma in albania. *Wspolczesna Onkol.* **2017**, *21*, 168–173. [CrossRef]
370. Heye, T.; Kuntz, C.; Dux, M.; Encke, J.; Palmowski, M.; Autschbach, F.; Volke, F.; Kauffmann, G.W.; Grenacher, L. Ct and endoscopic ultrasound in comparison to endoluminal MRI-preliminary results in staging gastric carcinoma. *Eur. J. Radiol.* **2009**, *70*, 336–341. [CrossRef]
371. Kim, A.Y.; Han, J.K.; Seong, C.K.; Kim, T.K.; Choi, B.I. MRI in staging advanced gastric cancer: Is it useful compared with spiral CT? *J. Comput. Assist. Tomogr.* **2000**, *24*, 389–394. [CrossRef]
372. Maccioni, F.; Marcelli, G.; Al Ansari, N.; Zippi, M.; De Marco, V.; Kagamanova, A.; Vestri, A.; Marcheg-giano-Clarke, L.; Marini, M. Preoperative t and n staging of gastric cancer: Magnetic resonance imaging (MRI) versus multi detector computed tomography (MDCT). *Clin. Ter.* **2010**, *161*, e57–e62.
373. Ma, L.; Xu, X.W.; Zhang, M.; Zheng, S.Q.; Zhang, B.; Zhang, W.; Wang, P.J. Dynamic contrast-enhanced MRI of gastric cancer: Correlations of the pharmacokinetic parameters with histological type, lauren classification, and angiogenesis. *Magn. Reson. Imaging* **2017**, *37*, 27–32. [CrossRef]

374. Joo, I.; Lee, J.M.; Han, J.K.; Yang, H.K.; Lee, H.J.; Choi, B.I. Dynamic contrast-enhanced MRI of gastric cancer: Correlation of the perfusion parameters with pathological prognostic factors. *J. Magn. Reson. Imaging* **2015**, *41*, 1608–1614. [CrossRef]
375. Bae, S.W.; Berlith, F.; Jeong, K.Y.; Suh, Y.S.; Kong, S.H.; Lee, H.J.; Kim, W.H.; Chung, J.K.; Yang, H.K. Establishment of a ¹⁸F FDG-PET/MRI imaging protocol for gastric cancer pdx as a preclinical research tool. *J. Gastric Cancer* **2020**, *20*, 60–71. [CrossRef]
376. Lee, D.H.; Kim, S.H.; Joo, I.; Hur, B.Y.; Han, J.K. Comparison between ¹⁸F-FDG PET/MRI and mdct for the assessment of preoperative staging and resectability of gastric cancer. *Eur. J. Radiol.* **2016**, *85*, 1085–1091. [CrossRef]
377. Liu, Y.; Zheng, D.; Liu, J.J.; Cui, J.X.; Xi, H.Q.; Zhang, K.C.; Huang, X.H.; Wei, B.; Wang, X.X.; Xu, B.X.; et al. Comparing PET/MRI with PET/CT for pretreatment staging of gastric cancer. *Gastroenterol. Res. Pract.* **2019**, *2019*, 9564627. [CrossRef]
378. Yuan, Y.; Chen, L.G.; Ren, S.N.; Wang, Z.; Chen, Y.K.; Jin, A.G.; Li, S.; Fang, X.; Wang, T.G.; Bian, Y.; et al. Diagnostic performance in t staging for patients with esophagogastric junction cancer using high-resolution MRI: A comparison with conventional MRI at 3 tesla. *Cancer Imaging* **2019**, *19*, 9. [CrossRef]
379. Pang, L.; Wang, J.; Fan, Y.; Xu, R.; Bai, Y.P.; Bai, L.C. Correlations of tnM staging and lymph node metastasis of gastric cancer with MRI features and vegf expression. *Cancer Biomark.* **2018**, *23*, 53–59. [CrossRef]
380. Giganti, F.; Orsenigo, E.; Arcidiacono, P.G.; Nicoletti, R.; Albarello, L.; Ambrosi, A.; Salerno, A.; Esposito, A.; Petrone, M.C.; Chiari, D.; et al. Preoperative locoregional staging of gastric cancer: Is there a place for magnetic resonance imaging? Prospective comparison with eus and multidetector computed tomography. *Gastric Cancer* **2016**, *19*, 216–225. [CrossRef]
381. Garrido-Laguna, I.; Hidalgo, M. Pancreatic cancer: From state-of-the-art treatments to promising novel therapies. *Nat. Rev. Clin. Oncol.* **2015**, *12*, 319–334. [CrossRef]
382. Barnes, C.A.; Krzywda, E.; Lahiff, S.; McDowell, D.; Christians, K.K.; Knechtges, P.; Tolat, P.; Hohenwalter, M.; Dua, K.; Khan, A.H.; et al. Development of a high risk pancreatic screening clinic using 3.0 t MRI. *Fam. Cancer* **2018**, *17*, 101–111. [CrossRef]
383. Gaa, J.; Fingerle, A.A.; Holzapfel, K.; Rummeny, E.J. MRI for malignant pancreatic tumors. *Radiologe* **2009**, *49*, 124–130. [CrossRef]
384. Ueno, M.; Niwa, T.; Ohkawa, S.; Amano, A.; Masaki, T.; Miyakawa, K.; Yoshida, T. The usefulness of perfusion-weighted magnetic resonance imaging in advanced pancreatic cancer. *Pancreas* **2009**, *38*, 644–648. [CrossRef] [PubMed]
385. Gassert, F.G.; Ziegelmayer, S.; Luitjens, J.; Gassert, F.T.; Tollens, F.; Rink, J.; Makowski, M.R.; Rubenthaler, J.; Froelich, M.F. Additional MRI for initial m-staging in pancreatic cancer: A cost-effectiveness analysis. *Eur. Radiol.* **2021**, *1*–9. [CrossRef] [PubMed]
386. Visser, B.C.; Yeh, B.M.; Qayyum, A.; Way, L.W.; McCulloch, C.E.; Coakley, F.V. Characterization of cystic pancreatic masses: Relative accuracy of CT and MRI. *Am. J. Roentgenol.* **2007**, *189*, 648–656. [CrossRef] [PubMed]
387. Chen, F.M.; Ni, J.M.; Zhang, Z.Y.; Zhang, L.; Li, B.; Jiang, C.J. Presurgical evaluation of pancreatic cancer: A comprehensive imaging comparison of CT versus MRI. *Am. J. Roentgenol.* **2016**, *206*, 526–535. [CrossRef] [PubMed]
388. Deng, Y.; Ming, B.; Wu, J.L.; Zhou, T.; Zhang, S.Y.; Chen, Y.; Lan, C.; Zhang, X.M. Magnetic resonance imaging for preoperative staging of pancreatic cancer based on the 8th edition of ajcc guidelines. *J. Gastrointest. Oncol.* **2020**, *11*, 329–336. [CrossRef] [PubMed]
389. Litjens, G.; Riviere, D.M.; van Geenen, E.J.M.; Radema, S.A.; Brosens, L.A.A.; Prokop, M.; van Laarhoven, C.; Hermans, J.J. Diagnostic accuracy of contrast-enhanced diffusion-weighted MRI for liver metastases of pancreatic cancer: Towards adequate staging and follow-up of pancreatic cancer—DIA-PANC study: Study protocol for an international, multicenter, diagnostic trial. *BMC Cancer* **2020**, *20*, 8. [CrossRef]
390. Jhaveri, K.S.; Jandaghi, A.B.; Thippavong, S.; Espin-Garcia, O.; Dodd, A.; Hutchinson, S.; Reichman, T.W.; Moulton, C.A.; McGilvary, I.D.; Gallinger, S. Can preoperative liver MRI with gadoxetic acid help reduce open-close laparotomies for curative intent pancreatic cancer surgery? *Cancer Imaging* **2021**, *21*, 12. [CrossRef]
391. Riviere, D.M.; van Geenen, E.J.M.; van der Kolk, B.M.; Nagtegaal, I.D.; Radema, S.A.; van Laarhoven, C.; Hermans, J.J. Improving preoperative detection of synchronous liver metastases in pancreatic cancer with combined contrast-enhanced and diffusion-weighted MRI. *Abdom. Radiol.* **2019**, *44*, 1756–1765. [CrossRef]
392. Yang, M.; Hopp, A.C.; Bekaii-Saab, T.S.; Collins, J.M. Sister mary joseph nodule in advanced pancreatic adenocarcinoma identified on ¹⁸F-FDG PET/MRI. *J. Nucl. Med. Technol.* **2019**, *47*, 341–342. [CrossRef]
393. Sandrasegaran, K.; Nutakki, K.; Tahir, B.; Dhanabal, A.; Tann, M.; Cote, G.A. Use of diffusion-weighted MRI to differentiate chronic pancreatitis from pancreatic cancer. *Am. J. Roentgenol.* **2013**, *201*, 1002–1008. [CrossRef]
394. Kim, J.K.; Altun, E.; Elias, J.; Pamuklar, E.; Rivero, H.; Semelka, R.C. Focal pancreatic mass: Distinction of pancreatic cancer from chronic pancreatitis using gadolinium-enhanced 3d-gradient-echo MRI. *J. Magn. Reson. Imaging* **2007**, *26*, 313–322. [CrossRef] [PubMed]
395. Liu, S.A.; Fu, W.W.; Liu, Z.J.; Liu, M.; Ren, R.M.; Zhai, H.X.; Li, C.L. MRI-guided celiac plexus neurolysis for pancreatic cancer pain: Efficacy and safety. *J. Magn. Reson. Imaging* **2016**, *44*, 923–928. [CrossRef] [PubMed]
396. Niwa, T.; Ueno, M.; Shinya, N.; Gotoh, T.; Kwee, T.C.; Takahara, T.; Yoshida, T.; Ohkawa, S.; Doiuchi, T.; Inoue, T. Dynamic susceptibility contrast MRI in advanced pancreatic cancer: Semi-automated analysis to predict response to chemotherapy. *NMR Biomed.* **2010**, *23*, 347–352. [CrossRef] [PubMed]
397. Zhang, T.; Zhang, F.; Meng, Y.F.; Wang, H.; Le, T.; Wei, B.J.; Lee, D.; Willis, P.; Shen, B.Z.; Yang, X.M. Diffusion-weighted MRI monitoring of pancreatic cancer response to radiofrequency heat-enhanced intratumor chemotherapy. *NMR Biomed.* **2013**, *26*, 1762–1767. [CrossRef]

398. Grandhi, M.S.; Kim, A.K.; Ronneklev-Kelly, S.M.; Kamel, I.R.; Ghasebeh, M.A.; Pawlik, T.M. Hepatocellular carcinoma: From diagnosis to treatment. *Surg. Oncol.* **2016**, *25*, 74–85. [[CrossRef](#)]
399. Wang, G.B.; Zhu, S.C.; Li, X.K. Comparison of values of CT and MRI imaging in the diagnosis of hepatocellular carcinoma and analysis of prognostic factors. *Oncol. Lett.* **2019**, *17*, 1184–1188. [[CrossRef](#)]
400. Kloeckner, R.; dos Santos, D.P.; Kreitner, K.F.; Leicher-Duber, A.; Weinmann, A.; Mittler, J.; Duber, C. Quantitative assessment of washout in hepatocellular carcinoma using MRI. *BMC Cancer* **2016**, *16*, 8. [[CrossRef](#)]
401. Sanghvi, T.; Boyum, J.; Spilseth, B.; Schat, R.; Estby, H.; Taylor, A. MRI for hepatocellular carcinoma: A primer for magnetic resonance imaging interpretation. *Abdom. Radiol.* **2018**, *43*, 1143–1151. [[CrossRef](#)]
402. Hectors, S.J.; Lewis, S.; Besa, C.; King, M.J.; Said, D.; Putra, J.; Ward, S.; Higashi, T.; Thung, S.; Yao, S.; et al. MRI radiomics features predict immuno-oncological characteristics of hepatocellular carcinoma. *Eur. Radiol.* **2020**, *30*, 3759–3769. [[CrossRef](#)]
403. Low, H.M.; Choi, J.Y.; Tan, C.H. Pathological variants of hepatocellular carcinoma on MRI: Emphasis on histopathologic correlation. *Abdom. Radiol.* **2019**, *44*, 493–508. [[CrossRef](#)]
404. Kim, Y.N.; Song, J.S.; Moon, W.S.; Hwang, H.P.; Kim, Y.K. Intra-individual comparison of hepatocellular carcinoma imaging computed tomography-enhanced computed tomography, gadopentetate dimeglumine-enhanced MRI, and gadoxetic acid-enhanced MRI. *Acta Radiol.* **2018**, *59*, 639–648. [[CrossRef](#)] [[PubMed](#)]
405. Gabr, A.E.; Mikhael, H.S.W.; El-Maadawy, S.M. Comparison between subtraction and dynamic MRI in assessing treatment response following radiofrequency ablation in patients with hepatocellular carcinoma. *Egypt. J. Radiol. Nucl. Med.* **2021**, *52*, 10. [[CrossRef](#)]
406. Jha, R.C.; Zanello, P.A.; Nguyen, X.M.; Pehlivanova, M.; Johnson, L.B.; Fishbein, T.; Shetty, K. Small hepatocellular carcinoma: MRI findings for predicting tumor growth rates. *Acad. Radiol.* **2014**, *21*, 1455–1464. [[PubMed](#)]
407. Abdullah, S.S.; Pialat, J.B.; Wiart, M.; Duboeuf, F.; Mabrut, J.Y.; Bancel, B.; Rode, A.; Ducerf, C.; Baulieux, J.; Berthezene, Y. Characterization of hepatocellular carcinoma and colorectal liver metastasis by means of perfusion MRI. *J. Magn. Reson. Imaging* **2008**, *28*, 390–395. [[CrossRef](#)]
408. Gong, X.Q.; Tao, Y.Y.; Wu, Y.K.; Liu, N.; Yu, X.; Wang, R.; Zheng, J.; Liu, N.; Huang, X.H.; Li, J.D.; et al. Progress of MRI radiomics in hepatocellular carcinoma. *Front. Oncol.* **2021**, *11*, 13. [[CrossRef](#)]
409. Carbonell, G.; Kennedy, P.; Bane, O.; Kirmani, A.; El Homsi, M.; Stocker, D.; Said, D.; Mukherjee, P.; Gevaert, O.; Lewis, S.; et al. Precision of MRI radiomics features in the liver and hepatocellular carcinoma. *Eur. Radiol.* **2021**, *1*–11. [[CrossRef](#)]
410. Kim, H.; Park, M.S.; Choi, J.Y.; Park, Y.N.; Kim, M.J.; Kim, K.S.; Choi, J.S.; Han, K.H.; Kim, E.; Kim, K.W. Can microvessel invasion of hepatocellular carcinoma be predicted by pre-operative MRI? *Eur. Radiol.* **2009**, *19*, 1744–1751. [[CrossRef](#)]
411. Hennedige, T.; Venkatesh, S.K. Imaging of hepatocellular carcinoma: Diagnosis, staging and treatment monitoring. *Cancer Imaging* **2013**, *12*, 530–547. [[CrossRef](#)]
412. Taneja, S.; Taneja, R.; Kashyap, V.; Jha, A.; Jena, A. Ga-68-psma uptake in hepatocellular carcinoma. *Clin. Nucl. Med.* **2017**, *42*, E69–E70. [[CrossRef](#)]
413. Dondi, F.; Albano, D.; Cerudelli, E.; Gazzilli, M.; Giubbini, R.; Treglia, G.; Bertagna, F. Radiolabelled psma PET/CT or PET/MRI in hepatocellular carcinoma (HCC): A systematic review. *Clin. Transl. Imaging* **2020**, *8*, 461–467. [[CrossRef](#)]
414. Qiu, Z.L.; Pan, Y.S.; Wei, J.; Wu, D.J.; Xia, Y.; Shen, D.G. Predicting symptoms from multiphasic MRI via multi-instance attention learning for hepatocellular carcinoma grading. In Proceedings of the International Conference on Medical Image Computing and Computer Assisted Intervention (MICCAI), Strasbourg, France, 27 September–1 October 2021; Springer: Cham, Switzerland, 2021; pp. 439–448.
415. Saito, K.; Ledsam, J.; Sugimoto, K.; Sourbron, S.; Araki, Y.; Tokuyue, K. Dce-MRI for early prediction of response in hepatocellular carcinoma after tace and sorafenib therapy: A pilot study. *J. Belg. Soc. Radiol.* **2018**, *102*, 9. [[CrossRef](#)] [[PubMed](#)]
416. Fan, Y.F.; Yu, Y.X.; Wang, X.M.; Hu, M.J.; Du, M.Z.; Guo, L.C.A.; Sun, S.F.; Hu, C.H. Texture analysis based on gd-eob-dtpa-enhanced MRI for identifying vessels encapsulating tumor clusters (vtc)-positive hepatocellular carcinoma. *J. Hepatocell. Carcinoma* **2021**, *8*, 349–359. [[CrossRef](#)] [[PubMed](#)]
417. Li, X.Q.; Wang, X.; Zhao, D.W.; Sun, J.; Liu, J.J.; Lin, D.D.; Yang, G.; Liu, H.; Xia, Z.Y.; Jia, C.Y.; et al. Application of gd-eob-dtpa-enhanced magnetic resonance imaging (MRI) in hepatocellular carcinoma. *World J. Surg. Oncol.* **2020**, *18*, 219. [[CrossRef](#)] [[PubMed](#)]
418. Hanna, R.F.; Kased, N.; Kwan, S.W.; Gamst, A.C.; Santosa, A.C.; Hassanein, T.; Sirlin, C.B. Double-contrast MRI for accurate staging of hepatocellular carcinoma in patients with cirrhosis. *Am. J. Roentgenol.* **2008**, *190*, 47–57. [[CrossRef](#)]
419. Wu, Y.N.; Huang, L.N.; Li, B.S.; Li, H.G. The diagnostic value of gd-eob-dtpa-enhanced MRI scans in small hepatocellular carcinoma in patients with liver cirrhosis. *Int. J. Clin. Exp. Med.* **2020**, *13*, 6909–6915.
420. Taouli, B.; Johnson, R.S.; Hajdu, C.H.; Oei, M.T.H.; Merad, M.; Yee, H.; Rusinek, H. Hepatocellular carcinoma: Perfusion quantification with dynamic contrast-enhanced MRI. *Am. J. Roentgenol.* **2013**, *201*, 795–800. [[CrossRef](#)]
421. Akhtar, S.; Hussain, M.; Ali, S.; Maqsood, S.; Akram, S.; Abbas, N. Comparison of positive predictive value of multiphasic dynamic contrast enhanced MRI with dynamic contrast enhanced ct for the detection of hepatocellular carcinoma. *Pak. J. Med. Health Sci.* **2020**, *14*, 562–564.
422. Gluskin, J.S.; Chegai, F.; Monti, S.; Squillaci, E.; Mannelli, L. Hepatocellular carcinoma and diffusion-weighted MRI: Detection and evaluation of treatment response. *J. Cancer* **2016**, *7*, 1565–1570. [[CrossRef](#)]

423. Chen, H.Y.; Hou, Y.L.; Ma, X.M.; Xie, H.Y.; Ye, M.; Bai, Y.R. Ct and MRI in target delineation in primary hepatocellular carcinoma. *Cancer Radiother.* **2013**, *17*, 750–754. [CrossRef]
424. Zemour, J.; Marty, M.; Lapuyade, B.; Collet, D.; Chiche, L. Gallbladder tumor and pseudotumor: Diagnosis and management. *J. Visc. Surg.* **2014**, *151*, 289–300. [CrossRef]
425. Tseng, J.H.; Wan, Y.L.; Hung, C.F.; Ng, K.K.; Pan, K.T.; Chou, A.S.B.; Liu, N.J. Diagnosis and staging of gallbladder carcinoma—Evaluation with dynamic MR imaging. *Clin. Imaging* **2002**, *26*, 177–182. [CrossRef]
426. Vendrami, C.L.; Magnetta, M.J.; Mittal, P.K.; Moreno, C.C.; Miller, F.H. Gallbladder carcinoma and its differential diagnosis at MRI: What radiologists should know. *Radiographics* **2021**, *41*, 78–95. [CrossRef] [PubMed]
427. Wu, S.S.; Zou, X.L.; Wang, Q.X.; Hu, D.Y.; Li, Z.; Xu, C.O. Gallbladder carcinoma: An initial clinical experience of reduced field-of-view diffusion-weighted MRI. *Cancer Imaging* **2020**, *20*, 8. [CrossRef] [PubMed]
428. Kuipers, H.; Hoogwater, F.J.H.; Holtman, G.A.; van der Hoorn, A.; de Boer, M.T.; de Haas, R.J. Clinical value of diffusion-weighted MRI for differentiation between benign and malignant gallbladder disease: A systematic review and meta-analysis. *Acta Radiol.* **2021**, *62*, 987–996. [CrossRef]
429. Corgna, E.; Betti, M.; Gatta, G.; Roila, F.; De Mulder, P.H.M. Renal cancer. *Crit. Rev. Oncol./Hematol.* **2007**, *64*, 247–261. [CrossRef]
430. Bensalah, K.; Albiges, L.; Bernhard, J.C.; Bigot, P.; Bodin, T.; Boissier, R.; Correas, J.M.; Gimel, P.; Hetet, J.F.; Long, J.A.; et al. French ccaf guidelines—Update 2018–2020: Management of kidney cancer. *Prog. Urol.* **2018**, *28*, R5–R33. [CrossRef]
431. Couvidat, C.; Eiss, D.; Verkarre, V.; Merran, S.; Correas, J.M.; Mejean, A.; Helenon, O. Renal papillary carcinoma: CT and MRI features. *Diagn. Interv. Imaging* **2014**, *95*, 1055–1063. [CrossRef]
432. Shinagare, A.B.; Davenport, M.S.; Park, H.; Pedrosa, I.; Remer, E.M.; Chandarana, H.; Doshi, A.M.; Schieda, N.; Smith, A.D.; Vikram, R.; et al. Lexicon for renal mass terms at CT and MRI: A consensus of the society of abdominal radiology disease-focused panel on renal cell carcinoma. *Abdom. Radiol.* **2021**, *46*, 703–722. [CrossRef]
433. Vannier, M.W. *Imaging Renal Carcinoma*; Demos Medical Publications: New York, NY, USA, 2011; Volume 2, pp. 37–56.
434. Chiarello, M.A.; Mali, R.D.; Kang, S.K. Diagnostic accuracy of MRI for detection of papillary renal cell carcinoma: A systematic review and meta-analysis. *Am. J. Roentgenol.* **2018**, *211*, 812–821. [CrossRef]
435. Wehrli, N.E.; Kim, M.J.; Matza, B.W.; Melamed, J.; Taneja, S.S.; Rosenkrantz, A.B. Utility of MRI features in differentiation of central renal cell carcinoma and renal pelvic urothelial carcinoma. *Am. J. Roentgenol.* **2013**, *201*, 1260–1267. [CrossRef]
436. Palmowski, M.; Schifferdecker, I.; Zwick, S.; Macher-Goeppinger, S.; Laue, H.; Haferkamp, A.; Kauczor, H.U.; Kiessling, F.; Hallscheidt, P. Tumor perfusion assessed by dynamic contrast-enhanced MRI correlates to the grading of renal cell carcinoma: Initial results. *Eur. J. Radiol.* **2010**, *74*, e177–e181. [CrossRef] [PubMed]
437. Lei, Y.; Wang, H.; Li, H.F.; Rao, Y.W.; Liu, J.H.; Tian, S.F.; Ju, Y.; Li, Y.; Chen, A.L.; Chen, L.H.; et al. Diagnostic significance of diffusion-weighted MRI in renal cancer. *BioMed Res. Int.* **2015**, *2015*, 172165. [CrossRef] [PubMed]
438. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2016. *CA Cancer J. Clin.* **2016**, *66*, 7–30. [CrossRef]
439. Saito, W.; Amanuma, M.; Tanaka, J.; Heshiki, A. Histopathological analysis of a bladder cancer stalk observed on MRI. *Magn. Reson. Imaging* **2000**, *18*, 411–415. [CrossRef]
440. Tillou, X.; Grardel, E.; Fourmarier, M.; Bernasconi, T.; Demainly, M.; Hakami, F.; Saint, F.; Petit, J. Can MRI be used to distinguish between superficial and invasive transitional cell bladder cancer? *Prog. Urol.* **2008**, *18*, 440–444. [CrossRef]
441. Dolz, J.; Xu, X.P.; Rony, J.; Yuan, J.; Liu, Y.; Granger, E.; Desrosiers, C.; Zhang, X.; Ben Ayed, I.; Lu, H.B. Multiregion segmentation of bladder cancer structures in MRI with progressive dilated convolutional networks. *Med. Phys.* **2018**, *45*, 5482–5493. [CrossRef] [PubMed]
442. Barentsz, J.O.; Jager, G.J.; Witjes, J.A.; Ruijs, J.H.J. Primary staging of urinary bladder carcinoma: The role of MRI and a comparison with ct. *Eur. Radiol.* **1996**, *6*, 129–133. [CrossRef]
443. Mehraj, A.; Hameed, A.; Nanda, S.; Wazir, B.S.; Khan, M.A. Role of MRI in staging urinary bladder tumors. *Ann. Med. Health Sci. Res.* **2017**, *7*, 116–121.
444. Abouelkheir, R.T.; Abdelhamid, A.; Abou El-Ghar, M.; El-Diasty, T. Imaging of bladder cancer: Standard applications and future trends. *Med. Lith.* **2021**, *57*, 220. [CrossRef]
445. Wong, V.C.K.; Ganeshan, D.; Jensen, C.T.; Devine, C.E. Imaging and management of bladder cancer. *Cancers* **2021**, *13*, 1396. [CrossRef]
446. Woo, S.; Suh, C.H.; Kim, S.Y.; Cho, J.Y.; Kim, S.H. Diagnostic performance of MRI for prediction of muscle-invasiveness of bladder cancer: A systematic review and meta-analysis. *Eur. J. Radiol.* **2017**, *95*, 46–55. [CrossRef] [PubMed]
447. Ghafoori, M.; Shakiba, M.; Ghiasi, A.; Asvadi, N.; Hosseini, K.; Alavi, M. Value of MRI in local staging of bladder cancer. *Urol. J.* **2013**, *10*, 866–872.
448. Rosenkrantz, A.B.; Mussi, T.C.; Melamed, J.; Taneja, S.S.; Huang, W.C. Bladder cancer: Utility of MRI in detection of occult muscle-invasive disease. *Acta Radiol.* **2012**, *53*, 695–699. [CrossRef] [PubMed]
449. Badawy, M.; Farg, H.; Gadelhak, B.; ElGhar, M.A.; Sadeq, A.G.; Borg, M. Diagnostic performance of 3-tesla multiparametric MRI for assessment of the bladder cancer t stage and histologic grade. *Egypt. J. Radiol. Nucl. Med.* **2020**, *51*, 11. [CrossRef]
450. Juri, H.; Narumi, Y.; Panebianco, V.; Osuga, K. Staging of bladder cancer with multiparametric MRI. *Br. J. Radiol.* **2020**, *93*, 7. [CrossRef] [PubMed]

451. Naish, J.H.; McGrath, D.M.; Bains, L.J.; Passera, K.; Roberts, C.; Watson, Y.; Cheung, S.; Taylor, M.B.; Logue, J.P.; Buckley, D.L.; et al. Comparison of dynamic contrast-enhanced MRI and dynamic contrast-enhanced CT biomarkers in bladder cancer. *Magn. Reson. Med.* **2011**, *66*, 219–226. [[CrossRef](#)]
452. Hijab, A.; Tocco, B.; Hanson, I.; Meijer, H.; Nyborg, C.J.; Bertelsen, A.S.; Smeenk, R.J.; Smith, G.; Michalski, J.; Baumann, B.C.; et al. Mr-guided adaptive radiotherapy for bladder cancer. *Front. Oncol.* **2021**, *11*, 12. [[CrossRef](#)]
453. Wollin, D.A.; Deng, F.M.; Huang, W.C.; Babb, J.S.; Rosenkrantz, A.B. Conventional and diffusion-weighted MRI features in diagnosis of metastatic lymphadenopathy in bladder cancer. *Can. J. Urol.* **2014**, *21*, 7454–7459.
454. Watanabe, H.; Kanematsu, M.; Kondo, H.; Goshima, S.; Tsuge, Y.; Onozuka, M.; Moriyama, N. Preoperative t staging of urinary bladder cancer: Does diffusion-weighted MRI have supplementary value? *Am. J. Roentgenol.* **2009**, *192*, 1361–1366. [[CrossRef](#)]
455. Howlader, N.; Noone, A.M.; Krapcho, M.; Miller, D.; Bishop, K.; Kosary, C.L.; Yu, M.; Ruhl, J.; Tatalovich, Z.; Mariotto, A.; et al. *Seer Cancer Statistics Review, 1975–2014*, April 2017 ed.; National Cancer Institute: Bethesda, MD, USA, 2017.
456. Sharma, S.K.; Nemieboka, B.; Sala, E.; Lewis, J.S.; Zeglis, B.M. Molecular imaging of ovarian cancer. *J. Nucl. Med.* **2016**, *57*, 827–833. [[CrossRef](#)]
457. Engbersen, M.P.; van't Sant, I.; Lok, C.; Lambregts, D.M.J.; Sonke, G.S.; Beets-Tan, R.G.H.; van Driel, W.J.; Lahaye, M.J. MRI with diffusion-weighted imaging to predict feasibility of complete cytoreduction with the peritoneal cancer index (PCI) in advanced stage ovarian cancer patients. *Eur. J. Radiol.* **2019**, *114*, 146–151. [[CrossRef](#)] [[PubMed](#)]
458. Michielsen, K.; Dresen, R.; Vanslembrouck, R.; De Keyzer, F.; Amant, F.; Mussen, E.; Leunen, K.; Berteloot, P.; Moerman, P.; Vergote, I.; et al. Diagnostic value of whole body diffusion-weighted MRI compared to computed tomography for pre-operative assessment of patients suspected for ovarian cancer. *Eur. J. Cancer* **2017**, *83*, 88–98. [[CrossRef](#)] [[PubMed](#)]
459. Rockall, A.G. Diffusion weighted MRI in ovarian cancer. *Curr. Opin. Oncol.* **2014**, *26*, 529–535. [[CrossRef](#)] [[PubMed](#)]
460. Carter, J.S.; Koopmeiners, J.S.; Kuehn-Hajder, J.E.; Metzger, G.J.; Lakkadi, N.; Downs, L.S.; Bolan, P.J. Quantitative multiparametric MRI of ovarian cancer. *J. Magn. Reson. Imaging* **2013**, *38*, 1501–1509. [[CrossRef](#)] [[PubMed](#)]
461. Mikkelsen, M.S.; Petersen, L.K.; Blaakaer, J.; Marinovskij, E.; Rosenkilde, M.; Andersen, G.; Bouchelouche, K.; Iversen, L.H. Assessment of peritoneal metastases with DW-MRI, CT, and FDG PET/CT before cytoreductive surgery for advanced stage epithelial ovarian cancer. *Eur. J. Surg. Oncol.* **2021**, *47*, 2134–2141. [[CrossRef](#)]
462. Khiewvan, B.; Torigian, D.A.; Emamzadehfard, S.; Paydar, K.; Salavati, A.; Houshmand, S.; Werner, T.J.; Alavi, A. An update on the role of PET/CT and PET/MRI in ovarian cancer. *Eur. J. Nucl. Med. Mol. Imaging* **2017**, *44*, 1079–1091. [[CrossRef](#)] [[PubMed](#)]
463. Tsuyoshi, H.; Tsujikawa, T.; Yamada, S.; Okazawa, H.; Yoshida, Y. Diagnostic value of $[^{18}\text{F}]$ FDG PET/MRI for staging in patients with ovarian cancer. *EJNMMI Res.* **2020**, *10*, 117. [[CrossRef](#)]
464. Kim, C.K.; Park, B.K.; Choi, J.Y.; Kim, B.G.; Han, H. Detection of recurrent ovarian cancer at MRI: Comparison with integrated PET/CT. *J. Comput. Assist. Tomogr.* **2007**, *31*, 868–875. [[CrossRef](#)]
465. Schmidt, S.; Meuli, R.A.; Achtari, C.; Prior, J.O. Peritoneal carcinomatosis in primary ovarian cancer staging comparison between mdct, MRI, and $[^{18}\text{F}]$ -FDG PET/CT. *Clin. Nucl. Med.* **2015**, *40*, 371–377. [[CrossRef](#)]
466. Sanli, Y.; Turkmen, C.; Bakir, B.; Iyibozkurt, C.; Ozel, S.; Has, D.; Yilmaz, E.; Topuz, S.; Yavuz, E.; Unal, S.N.; et al. Diagnostic value of PET/CT is similar to that of conventional MRI and even better for detecting small peritoneal implants in patients with recurrent ovarian cancer. *Nucl. Med. Commun.* **2012**, *33*, 509–515. [[CrossRef](#)]
467. Nam, E.J.; Yun, M.J.; Oh, Y.T.; Kim, J.W.; Kim, J.H.; Kim, S.; Jung, Y.W.; Kim, S.W.; Kim, Y.T. Diagnosis and staging of primary ovarian cancer: Correlation between PET/CT, doppler us, and CT or MRI. *Gynecol. Oncol.* **2010**, *116*, 389–394. [[PubMed](#)]
468. Reinhold, C.; Rockall, A.; Sadowski, E.A.; Siegelman, E.S.; Maturen, K.E.; Vargas, H.A.; Forstner, R.; Glanc, P.; Andreotti, R.F.; Thomassin-Naggara, I. Ovarian-adnexal reporting lexicon for MRI: A white paper of the acr ovarian-adnexal reporting and data systems MRI committee. *J. Am. Coll. Radiol.* **2021**, *18*, 713–729. [[CrossRef](#)] [[PubMed](#)]
469. Murakami, N.; Ando, K.; Murata, M.; Murata, K.; Ohno, T.; Aoshika, T.; Kato, S.; Okonogi, N.; Saito, A.I.; Kim, J.Y.; et al. Why not de-intensification for uterine cervical cancer? *Gynecol. Oncol.* **2021**, *163*, 105–109. [[CrossRef](#)] [[PubMed](#)]
470. Novellas, S.; Fournol, M.; Marcotte-Bloch, C.; Mondot, L.; Caramella, T.; Bongain, A.; Chevallier, P. Magnetic resonance staging of uterine cervix carcinoma. *Feuill. Rad.* **2008**, *48*, 147–154. [[CrossRef](#)]
471. Bourgioti, C.; Chatoupis, K.; Moulopoulos, L.A. Current imaging strategies for the evaluation of uterine cervical cancer. *World J. Radiol.* **2016**, *8*, 342–354. [[CrossRef](#)]
472. Theodore, C.; Levaillant, J.M.; Capmas, P.; Chabi, N.; Skalli, D.; Vienet-Legue, L.; Haddad, B.; Fernandez, H.; Touboul, C. MRI and ultrasound fusion imaging for cervical cancer. *Anticancer Res.* **2017**, *37*, 5079–5085.
473. Marzouk, F.; Jalaguier-Coudray, A.; Villard-Mahjoub, R. Uterine cervical cancer: The new figo classification. *Imaging Femme* **2021**, *31*, 30–39.
474. Kasuya, G.; Toita, T.; Furutani, K.; Kodaira, T.; Ohno, T.; Kaneyasu, Y.; Yoshimura, R.; Uno, T.; Yogi, A.; Ishikura, S.; et al. Distribution patterns of metastatic pelvic lymph nodes assessed by ct/MRI in patients with uterine cervical cancer. *Radiat. Oncol.* **2013**, *8*, 6. [[CrossRef](#)]
475. Zhu, Y.X.; Shen, B.M.; Pei, X.; Liu, H.X.; Li, G.Y. Ct, MRI, and PET imaging features in cervical cancer staging and lymph node metastasis. *Am. J. Transl. Res.* **2021**, *13*, 10536–10544.
476. Grueneisen, J.; Schaarschmidt, B.M.; Heubner, M.; Aktas, B.; Kinner, S.; Forsting, M.; Lauenstein, T.; Ruhlmann, V.; Umutlu, L. Integrated PET/MRI for whole-body staging of patients with primary cervical cancer: Preliminary results. *Eur. J. Nucl. Med. Mol. Imaging* **2015**, *42*, 1814–1824. [[CrossRef](#)]

477. Monteil, J.; Maubon, A.; Leobon, S.; Roux, S.; Marin, B.; Renaudie, J.; Genet, D.; Fermeaux, V.; Aubard, Y.; Tubiana-Mathieu, N. Lymph node assessment with ^{18}F -FDG-PET and MRI in uterine cervical cancer. *Anticancer Res.* **2011**, *31*, 3865–3871. [PubMed]
478. de Boer, P.; Adam, J.A.; Buist, M.R.; van de Vijver, M.J.; Rasch, C.R.; Stoker, J.; Bipat, S.; Stalpers, L.J.A. Role of MRI in detecting involvement of the uterine internal os in uterine cervical cancer: Systematic review of diagnostic test accuracy. *Eur. J. Radiol.* **2013**, *82*, E422–E428. [CrossRef] [PubMed]
479. Kitajima, K.; Suenaga, Y.; Ueno, Y.; Kanda, T.; Maeda, T.; Deguchi, M.; Ebina, Y.; Yamada, H.; Takahashi, S.; Sugimura, K. Fusion of PET and MRI for staging of uterine cervical cancer: Comparison with contrast-enhanced ^{18}F -FDG PET/CT and pelvic MRI. *Clin. Imaging* **2014**, *38*, 464–469. [CrossRef] [PubMed]
480. Balleyguier, C.; Fournet, C.; Ben Hassen, W.; Zareski, E.; Morice, P.; Haie-Meder, C.; Uzan, C.; Gouy, S.; Duvillard, P.; Lhomme, C. Management of cervical cancer detected during pregnancy: Role of magnetic resonance imaging. *Clin. Imaging* **2013**, *37*, 70–76. [CrossRef]
481. Surov, A.; Meyer, H.J.; Schob, S.; Hohn, A.K.; Bremicker, K.; Exner, M.; Stumpp, P.; Purz, S. Parameters of simultaneous ^{18}F -FDG-PET/MRI predict tumor stage and several histopathological features in uterine cervical cancer. *Oncotarget* **2017**, *8*, 28285–28296. [CrossRef]
482. Lee, J.; Kim, C.K.; Gu, K.W.; Park, W. Value of blood oxygenation level-dependent MRI for predicting clinical outcomes in uterine cervical cancer treated with concurrent chemoradiotherapy. *Eur. Radiol.* **2019**, *29*, 6256–6265. [CrossRef]
483. Saida, T.; Sakata, A.; Tanaka, Y.O.; Ochi, H.; Ishiguro, T.; Sakai, M.; Takahashi, H.; Satoh, T.; Minami, M. Clinical and MRI characteristics of uterine cervical adenocarcinoma: Its variants and mimics. *Korean J. Radiol.* **2019**, *20*, 14. [CrossRef]
484. Steiner, A.; Narva, S.; Rinta-Kiikka, I.; Hietanen, S.; Hyyninen, J.; Virtanen, J. Diagnostic efficiency of whole-body ^{18}F -FDG PET/MRI, MRI alone, and suv and adc values in staging of primary uterine cervical cancer. *Cancer Imaging* **2021**, *21*, 11. [CrossRef]
485. Ran, C.; Sun, J.; Qu, Y.H.; Long, N. Clinical value of MRI, serum scca, and ca125 levels in the diagnosis of lymph node metastasis and para-uterine infiltration in cervical cancer. *World J. Surg. Oncol.* **2021**, *19*, 11. [CrossRef]
486. Park, B.K.; Kim, T.J. Useful MRI findings for minimally invasive surgery for early cervical cancer. *Cancers* **2021**, *13*, 4078. [CrossRef]
487. Hauge, A.; Wegner, C.S.; Gaustad, J.V.; Simonsen, T.G.; Andersen, L.M.K.; Rofstad, E.K. Diffusion-weighted MRI-derived adc values reflect collagen i content in pdx models of uterine cervical cancer. *Oncotarget* **2017**, *8*, 105682–105691. [CrossRef] [PubMed]
488. Liu, Y.; Ye, Z.X.; Sun, H.R.; Bai, R.J. Clinical application of diffusion-weighted magnetic resonance imaging in uterine cervical cancer. *Int. J. Gynecol. Cancer* **2015**, *25*, 1073–1078. [CrossRef] [PubMed]
489. Liu, Y.; Liu, H.D.; Bai, X.; Ye, Z.X.; Sun, H.R.; Bai, R.J.; Wang, D.H. Differentiation of metastatic from non-metastatic lymph nodes in patients with uterine cervical cancer using diffusion-weighted imaging. *Gynecol. Oncol.* **2011**, *122*, 19–24. [CrossRef] [PubMed]
490. Liu, Y.; Bai, R.J.; Sun, H.R.; Liu, H.D.; Wang, D.H. Diffusion-weighted magnetic resonance imaging of uterine cervical cancer. *J. Comput. Assist. Tomogr.* **2009**, *33*, 858–862.
491. Hallac, R.R.; Ding, Y.; Yuan, Q.; McColl, R.W.; Lea, J.; Sims, R.D.; Weatherall, P.T.; Mason, R.P. Oxygenation in cervical cancer and normal uterine cervix assessed using blood oxygenation level-dependent (bold) MRI at 3t. *Nmr Biomed.* **2012**, *25*, 1321–1330. [CrossRef]
492. Ryan, A.J.; Susil, B.; Jobling, T.W.; Oehler, M.K. Endometrial cancer. *Cell Tissue Res.* **2005**, *322*, 53–61. [CrossRef]
493. Soneji, N.D.; Bharwani, N.; Ferri, A.; Stewart, V.; Rockall, A. Pre-operative MRI staging of endometrial cancer in a multicentre cancer network: Can we match single centre study results? *Eur. Radiol.* **2018**, *28*, 4725–4734. [CrossRef]
494. Taieb, S.; Rocourt, N.; Narducci, F.; Ceugnart, L. Endometrial cancer imaging. *Bull. Cancer* **2012**, *99*, 13–20.
495. Nougaret, S.; Horta, M.; Sala, E.; Lakhman, Y.; Thomassin-Naggara, I.; Kido, A.; Masselli, G.; Bharwani, N.; Sadowski, E.; Ertmer, A.; et al. Endometrial cancer MRI staging: Updated guidelines of the european society of urogenital radiology. *Eur. Radiol.* **2019**, *29*, 792–805. [CrossRef]
496. Meissnitzer, M.; Forstner, R. MRI of endometrium cancer—How we do it. *Cancer Imaging* **2016**, *16*, 9. [CrossRef]
497. Ortoft, G.; Dueholm, M.; Mathiesen, O.; Hansen, E.S.; Lundorf, E.; Moller, C.; Marinovskij, E.; Petersen, L.K. Preoperative staging of endometrial cancer using tvs, MRI, and hysteroscopy. *Acta Obstet. Gynecol. Scand.* **2013**, *92*, 536–545. [CrossRef] [PubMed]
498. Sadowski, E.A.; Robbins, J.B.; Guite, K.; Patel-Lippmann, K.; del Rio, A.M.; Kushner, D.M.; Al-Niaimi, A. Preoperative pelvic MRI and serum cancer antigen-125: Selecting women with grade 1 endometrial cancer for lymphadenectomy. *Am. J. Roentgenol.* **2015**, *205*, W556–W564. [CrossRef] [PubMed]
499. Tanaka, T.; Terai, Y.; Ono, Y.J.; Fujiwara, S.; Tanaka, Y.; Sasaki, H.; Tsunetoh, S.; Kanemura, M.; Yamamoto, K.; Yamada, T.; et al. Preoperative MRI and intraoperative frozen section diagnosis of myometrial invasion in patients with endometrial cancer. *Int. J. Gynecol. Cancer* **2015**, *25*, 879–883. [CrossRef] [PubMed]
500. Abu Freij, M.; Saleh, H.; Rawlins, H.; Duncan, T.; Nieto, J. The use of MRI for selecting patients with endometrial cancer and significant co-morbidities for vaginal hysterectomy. *Arch. Gynecol. Obstet.* **2011**, *283*, 1097–1101. [CrossRef] [PubMed]
501. Ben-Shachar, I.; Vitellas, K.M.; Cohn, D.E. The role of MRI in the conservative management of endometrial cancer. *Gynecol. Oncol.* **2004**, *93*, 233–237. [CrossRef]
502. Bakir, B.; Sanli, S.; Bakir, V.L.; Ayas, S.; Yildiz, S.O.; Iyibozkurt, A.C.; Kartal, M.G.; Yavuz, E. Role of diffusion weighted MRI in the differential diagnosis of endometrial cancer, polyp, hyperplasia, and physiological thickening. *Clin. Imaging* **2017**, *41*, 86–94. [CrossRef]

503. Zhou, Y.; Jiang, L.; Nuerlan, T. Application of diffusion-weighted MRI for endometrial cancer diagnosis. *Eur. J. Gynaecol. Oncol.* **2018**, *39*, 655–658.
504. Ma, Q.L.; Wang, H.Y.; Tao, C.; Duan, F.; Li, X.L. Evaluation of 3.0t multimodal MRI in patients with endometrial cancer. *Acta Med. Mediterr.* **2021**, *37*, 1887–1892.
505. Chen, J.Y.; Gu, H.L.; Fan, W.M.; Wang, Y.H.; Chen, S.; Chen, X.; Wang, Z.Q. MRI-based radiomic model for preoperative risk stratification in stage I endometrial cancer. *J. Cancer* **2021**, *12*, 726–734. [[CrossRef](#)]
506. Fasmer, K.E.; Hodneland, E.; Dybvik, J.A.; Wagner-Larsen, K.; Trovik, J.; Salvesen, O.; Krakstad, C.; Haldorsen, I.H.S. Whole-volume tumor MRI radiomics for prognostic modeling in endometrial cancer. *J. Magn. Reson. Imaging* **2021**, *53*, 928–937. [[CrossRef](#)]
507. Keles, D.K.; Evrimler, S.; Merd, N.; Erdemoglu, E. Endometrial cancer: The role of MRI quantitative assessment in preoperative staging and risk stratification. *Acta Radiol* **2021**. [[CrossRef](#)] [[PubMed](#)]
508. Chen, J.Y.; Fan, W.M.; Gu, H.L.; Zhang, W.; Liu, Y.T.; Wang, Y.J.; Pan, Z.C.; Wang, Z.Q. Preoperative MRI and immunohistochemical examination for the prediction of high-risk endometrial cancer. *Gland Surg.* **2021**, *10*, 2180–2191. [[CrossRef](#)] [[PubMed](#)]
509. Kim, H.J.; Cho, A.; Yun, M.; Kim, Y.T.; Kang, W. Comparison of FDG PET/CT and MRI in lymph node staging of endometrial cancer. *Ann. Nucl. Med.* **2016**, *30*, 104–113. [[CrossRef](#)] [[PubMed](#)]
510. Tsuyoshi, H.; Tsujikawa, T.; Yamada, S.; Okazawa, H.; Yoshida, Y. Diagnostic value of ¹⁸F-FDG PET/MRI for staging in patients with endometrial cancer. *Cancer Imaging* **2020**, *20*, 9. [[CrossRef](#)] [[PubMed](#)]
511. Bezzini, C.; Zambella, E.; Ghezzo, S.; Fallanca, F.; Samanes Gajate, A.M.; Franchini, A.; Ironi, G.; Bergamini, A.; Monaco, L.; Evangelista, L.; et al. ¹⁸F-FDG PET/MRI in endometrial cancer: Systematic review and meta-analysis. *Clin. Transl. Imaging* **2021**, *1*–14. [[CrossRef](#)]
512. Kitajima, K.; Suenaga, Y.; Ueno, Y.; Kanda, T.; Maeda, T.; Takahashi, S.; Ebina, Y.; Miyahara, Y.; Yamada, H.; Sugimura, K. Value of fusion of PET and MRI for staging of endometrial cancer: Comparison with ¹⁸F-FDG contrast-enhanced PET/CT and dynamic contrast-enhanced pelvic MRI. *Eur. J. Radiol.* **2013**, *82*, 1672–1676. [[CrossRef](#)]
513. Bian, L.H.; Wang, M.; Gong, J.; Liu, H.H.; Wang, N.; Wen, N.; Fan, W.S.; Xu, B.X.; Wang, M.Y.; Ye, M.X.; et al. Comparison of integrated PET/MRI with PET/ct in evaluation of endometrial cancer: A retrospective analysis of 81 cases. *PeerJ* **2019**, *7*, 10. [[CrossRef](#)]
514. Hama, Y.; Tate, E. Palliative MRI-guided intensity modulated radiation therapy (imrt) for locally recurrent endometrial cancer. *Aktualni Gynekol. Porod.* **2020**, *12*, 70–74.
515. Whitaker, H.; Tam, J.O.; Connor, M.J.; Grey, A. Prostate cancer biology & genomics. *Transl. Androl. Urol.* **2020**, *9*, 1481–1491.
516. Shukla-Dave, A.; Hricak, H. Role of MRI in prostate cancer detection. *NMR Biomed.* **2014**, *27*, 16–24. [[CrossRef](#)]
517. Hotker, A.M.; Dappa, E.; Mazaheri, Y.; Ehdaie, B.; Zheng, J.T.; Capanu, M.; Hricak, H.; Akin, O. The influence of background signal intensity changes on cancer detection in prostate MRI. *Am. J. Roentgenol.* **2019**, *212*, 823–829. [[CrossRef](#)] [[PubMed](#)]
518. Ventrella, E.; Eusebi, L.; Carpagnano, F.A.; Bartelli, F.; Cormio, L.; Guglielmi, G. Multiparametric MRI of prostate cancer: Recent advances. *Curr. Radiol. Rep.* **2020**, *8*, 14. [[CrossRef](#)]
519. Sun, Y.; Reynolds, H.M.; Parameswaran, B.; Wraith, D.; Finnegan, M.E.; Williams, S.; Haworth, A. Multiparametric MRI and radiomics in prostate cancer: A review. *Australas. Phys. Eng. Sci. Med.* **2019**, *42*, 3–25. [[CrossRef](#)] [[PubMed](#)]
520. Kumar, V.; Bora, G.S.; Kumar, R.; Jagannathan, N.R. Multiparametric (MP) MRI of prostate cancer. *Prog. Nucl. Magn. Reson. Spectrosc.* **2018**, *105*, 23–40. [[CrossRef](#)] [[PubMed](#)]
521. Ghai, S.; Haider, M.A. Multiparametric-MRI in diagnosis of prostate cancer. *Indian J. Urol.* **2015**, *31*, 194–201. [[CrossRef](#)]
522. Wu, R.C.; Lebastchi, A.H.; Hadachik, B.A.; Emberton, M.; Moore, C.; Laguna, P.; Futterer, J.J.; George, A.K. Role of MRI for the detection of prostate cancer. *World J. Urol.* **2021**, *39*, 637–649. [[CrossRef](#)]
523. Piert, M.; El Naqa, I.; Davenport, M.S.; Incerti, E.; Mapelli, P.; Picchio, M. Pet/MRI and prostate cancer. *Clin. Transl. Imaging* **2016**, *4*, 473–485. [[CrossRef](#)]
524. Park, H.; Wood, D.; Hussain, H.; Meyer, C.R.; Shah, R.B.; Johnson, T.D.; Chenevert, T.; Piert, M. Introducing parametric fusion PET/MRI of primary prostate cancer. *J. Nucl. Med.* **2012**, *53*, 546–551. [[CrossRef](#)]
525. Lindenberg, L.; Ahlman, M.; Turkbey, B.; Mena, E.; Choyke, P. Evaluation of prostate cancer with PET/MRI. *J. Nucl. Med.* **2016**, *57*, 111S–116S. [[CrossRef](#)]
526. Kreydin, E.I.; Barrisford, G.W.; Feldman, A.S.; Preston, M.A. Testicular cancer: What the radiologist needs to know. *Am. J. Roentgenol.* **2013**, *200*, 1215–1225. [[CrossRef](#)]
527. Nestler, T.; Baunacke, M.; Drager, D.; von Landenberg, N.; Groeben, C.; Huber, J. Testicular cancer guideline adherence and patterns of care in germany: A nationwide survey. *Eur. J. Cancer Care* **2019**, *28*, 9. [[CrossRef](#)] [[PubMed](#)]
528. Hale, G.R.; Teplitsky, S.; Truong, H.; Gold, S.A.; Bloom, J.B.; Agarwal, P.K. Lymph node imaging in testicular cancer. *Transl. Androl. Urol.* **2018**, *7*, 864–874. [[CrossRef](#)] [[PubMed](#)]
529. Dugal, M.; Lei, J.; Dou, S.W.; Zhu, S.C. Role of imaging in testicular cancer. *Curr. Med. Imaging Rev.* **2017**, *13*, 38–49. [[CrossRef](#)]
530. Rud, E.; Langberg, C.W.; Baco, E.; Lauritzen, P.; Sandbaek, G. MRI in the follow-up of testicular cancer: Less is more. *Anticancer Res.* **2019**, *39*, 2963–2968. [[CrossRef](#)] [[PubMed](#)]
531. Tsili, A.C.; Sofikitis, N.; Stiliaris, E.; Argyropoulou, M.I. MRI of testicular malignancies. *Abdom. Radiol.* **2019**, *44*, 1070–1082. [[CrossRef](#)] [[PubMed](#)]

532. Larsen, S.K.A.; Agerbaek, M.; Jurik, A.G.; Pedersen, E.M. Ten years of experience with MRI follow-up of testicular cancer stage i: A retrospective study and an MRI protocol with dwi. *Acta Oncol.* **2020**, *59*, 1374–1381. [CrossRef]
533. Manganaro, L.; Saldari, M.; Pozza, C.; Vinci, V.; Gianfrilli, D.; Greco, E.; Franco, G.; Sergi, M.E.; Scialpi, M.; Catalano, C.; et al. Dynamic contrast-enhanced and diffusion-weighted mr imaging in the characterisation of small, non-palpable solid testicular tumours. *Eur. Radiol.* **2018**, *28*, 554–564. [CrossRef] [PubMed]
534. Laukka, M.; Mannisto, S.; Beule, A.; Kouri, M.; Blomqvist, C. Comparison between ct and MRI in detection of metastasis of the retroperitoneum in testicular germ cell tumors: A prospective trial. *Acta Oncol.* **2020**, *59*, 660–665. [CrossRef] [PubMed]
535. Reinges, M.H.T.; Kaiser, W.A.; Miersch, W.D.; Vogel, J.; Reiser, M. Dynamic MRI of benign and malignant testicular lesions—Preliminary-observations. *Eur. Radiol.* **1995**, *5*, 615–622. [CrossRef]
536. Rocher, L.; Ksouri, A.; Maxwell, F.; Bresson, B.; Hindawi, G.; Balasa, C.; Bellin, M.F.; Albiges, L. Testicular tumors: A diagnostic challenge of imaging. *Bull. Cancer* **2019**, *106*, 875–886. [CrossRef]
537. Khanna, M.; Abualruz, A.R.; Yadav, S.K.; Mafraji, M.; Al-Rumaihi, K.; Al-Bozom, I.; Kumar, D.; Tsili, A.C.; Schieda, N. Diagnostic performance of multi-parametric MRI to differentiate benign sex cord stromal tumors from malignant (non-stromal and stromal) testicular neoplasms. *Abdom. Radiol.* **2021**, *46*, 319–330. [CrossRef] [PubMed]
538. Sohaib, S.A.; Koh, D.M.; Barbachano, Y.; Parikh, J.; Husband, J.E.S.; Dearnaley, D.P.; Horwich, A.; Huddart, R. Prospective assessment of MRI for imaging retroperitoneal metastases from testicular germ cell tumours. *Clin. Radiol.* **2009**, *64*, 362–367. [CrossRef] [PubMed]
539. Ortiz, A.F.H.; Beaujon, L.J.F.; Villamizar, S.Y.G.; Lopez, F.F.F. Magnetic resonance versus computed tomography for the detection of retroperitoneal lymph node metastasis due to testicular cancer: A systematic literature review. *Eur. J. Radiol. Open* **2021**, *8*, 7.
540. Kubik-Huch, R.A.; Hailemariam, S.; Hamm, B. Ct and MRI of the male genital tract: Radiologic-pathologic correlation. *Eur. Radiol.* **1999**, *9*, 16–28. [CrossRef] [PubMed]
541. Bleiberg, H. Colorectal cancer: The challenge. *Eur. J. Cancer* **1996**, *32A*, S2–S6. [CrossRef]
542. de Souza, G.D.; Souza, L.R.Q.; Cuenca, R.M.; Vilela, V.M.; Santos, B.; de Aguiar, F.S. Pre- and postoperative imaging methods in colorectal cancer. *ABCD-Arq. Bras. Cir. Dig.-Braz. Arch. Dig. Surg.* **2018**, *31*, 5. [CrossRef]
543. Georgiou, P.A.; Tekkis, P.P.; Constantinides, V.A.; Patel, U.; Goldin, R.D.; Darzi, A.W.; Nicholls, R.J.; Brown, G. Diagnostic accuracy and value of magnetic resonance imaging (MRI) in planning exenterative pelvic surgery for advanced colorectal cancer. *Eur. J. Cancer* **2013**, *49*, 72–81. [CrossRef]
544. Annemans, L.; Lencioni, R.; Warie, H.; Bartolozzi, C.; Ciceri, M.; Muller, U. Health economic evaluation of ferucarbotran-enhanced MRI in the diagnosis of liver metastases in colorectal cancer patients. *Int. J. Colorectal Dis.* **2008**, *23*, 77–83. [CrossRef]
545. Engbersen, M.P.; Rijsement, C.J.V.; Nederend, J.; Aalbers, A.G.J.; de Hingh, I.; Retel, V.; Lambregts, D.M.J.; Van der Hoeven, E.; Boerma, D.; Wiezer, M.J.; et al. Dedicated MRI staging versus surgical staging of peritoneal metastases in colorectal cancer patients considered for crs-hipec; the disco randomized multicenter trial. *BMC Cancer* **2021**, *21*, 8. [CrossRef]
546. Westberg, K.; Othman, B.; Suzuki, C.; Blomqvist, L.; Martling, A.; Iversen, H. Magnetic resonance imaging as a predictor of surgical outcome in patients with local pelvic recurrence of colorectal cancer. *Eur. J. Surg. Oncol.* **2021**, *47*, 2119–2124. [CrossRef]
547. Nasseri, Y.; Langenfeld, S.J. Imaging for colorectal cancer. *Surg. Clin.* **2017**, *97*, 503–513. [CrossRef] [PubMed]
548. Dresen, R.C.; De Vuysere, S.; De Keyzer, F.; Van Cutsem, E.; Prenen, H.; Vanslembrouck, R.; De Hertogh, G.; Wolthuis, A.; D’Hoore, A.; Vandecaveye, V. Whole-body diffusion-weighted MRI for operability assessment in patients with colorectal cancer and peritoneal metastases. *Cancer Imaging* **2019**, *19*, 10. [CrossRef] [PubMed]
549. Sabry, M.S.; Rady, A.E.E.; Niazi, G.E.M.; Ali, S.A. Role of diffusion-weighted MRI in diagnosis and post therapeutic follow-up of colorectal cancer. *Egypt. J. Radiol. Nucl. Med.* **2021**, *52*, 11. [CrossRef]
550. Ono, K.; Ochiai, R.; Yoshida, T.; Kitagawa, M.; Omagari, J.; Kobayashi, H.; Yamashita, Y. Comparison of diffusion-weighted MRI and 2- fluorine-18 -fluoro-2-deoxy-d-glucose positron emission tomography (FDG-PET) for detecting primary colorectal cancer and regional lymph node metastases. *J. Magn. Reson. Imaging* **2009**, *29*, 336–340. [CrossRef] [PubMed]
551. Kim, T.H.; Woo, S.; Han, S.; Suh, C.H.; Vargas, H.A. The diagnostic performance of MRI for detection of extramural venous invasion in colorectal cancer: A systematic review and meta-analysis of the literature. *Am. J. Roentgenol.* **2019**, *213*, 575–585. [CrossRef] [PubMed]
552. Smith, N.J.; Shihab, O.; Arnaout, A.; Swift, R.I.; Brown, G. MRI for detection of extramural vascular invasion in rectal cancer. *Am. J. Roentgenol.* **2008**, *191*, 1517–1522. [CrossRef]
553. Soomro, M.H.; De Cola, G.; Conforto, S.; Schmid, M.; Giunta, G.; Guidi, E.; Neri, E.; Caruso, D.; Ciolina, M.; Laghi, A.; et al. Automatic segmentation of colorectal cancer in 3D MRI by combining deep learning and 3D level-set algorithm—a preliminary study. In Proceedings of the 2018 IEEE 4th Middle East Conference on Biomedical Engineering (MECBME), Tunis, Tunisia, 28–30 March 2018; pp. 198–203.
554. Ogawa, M.; Ichiba, N.; Watanabe, M.; Yanaga, K. The usefulness of diffusion MRI in detection of lymph node metastases of colorectal cancer. *Anticancer Res.* **2016**, *36*, 815–819.
555. Ichikawa, T.; Erturk, S.M.; Motosugi, U.; Sou, H.; Iino, H.; Araki, T.; Fujii, H. High-b-value diffusion-weighted MRI in colorectal cancer. *Am. J. Roentgenol.* **2006**, *187*, 181–184. [CrossRef]
556. García-Figueiras, R.; Baleato-González, S.; Canedo-Antelo, M.; Alcalá, L.; Marhuenda, A. Imaging advances on ct and MRI in colorectal cancer. *Curr. Colorectal Cancer Rep.* **2021**, *17*, 113–130. [CrossRef]

557. Kijima, S.; Sasaki, T.; Nagata, K.; Utano, K.; Lefor, A.T.; Sugimoto, H. Preoperative evaluation of colorectal cancer using ct colonography, MRI, and PET/CT. *World J. Gastroenterol.* **2014**, *20*, 16964–16975. [[CrossRef](#)]
558. Kulemann, V.; Schima, W.; Tamandl, D.; Kaczirek, K.; Gruenberger, T.; Wrba, F.; Weber, M.; Ba-Salamah, A. Preoperative detection of colorectal liver metastases in fatty liver: MDCT or MRI? *Eur. J. Radiol.* **2011**, *79*, E1–E6. [[CrossRef](#)] [[PubMed](#)]
559. Kang, B.; Lee, J.M.; Song, Y.S.; Woo, S.; Hur, B.Y.; Jeon, J.H.; Paeng, J.C. Added value of integrated whole-body PET/MRI for evaluation of colorectal cancer: Comparison with contrast-enhanced mdct. *Am. J. Roentgenol.* **2016**, *206*, W10–W20. [[CrossRef](#)] [[PubMed](#)]
560. Schmidt, G. Importance of whole body MRI for staging of colorectal cancer. *Radiolge* **2012**, *52*, 537–544. [[CrossRef](#)] [[PubMed](#)]
561. Datta, N.R.; Krishnan, S.; Speiser, D.E.; Neufeld, E.; Kuster, N.; Bodis, S.; Hofmann, H. Magnetic nanoparticle-induced hyperthermia with appropriate payloads: Paul ehrlich's "magic (nano)bullet" for cancer theranostics? *Cancer Treat. Rev.* **2016**, *50*, 217–227. [[CrossRef](#)] [[PubMed](#)]
562. Douziech-Eyrolles, L.; Marchais, H.; Herve, K.; Munnier, E.; Souce, M.; Linassier, C.; Dubois, P.; Chourpa, I. Nanovectors for anticancer agents based on superparamagnetic iron oxide nanoparticles. *Int. J. Nanomed.* **2007**, *2*, 541–550.
563. Balaita, L.; Popa, M. Polymer magnetic particles in biomedical applications. *Rev. Roum. Chim.* **2009**, *54*, 185–199.
564. Rotariu, O.; Udrea, L.E.; Strachan, N.J.C.; Badescu, V. The guidance of magnetic colloids in simulated tissues for targeted drug delivery. *J. Optoelectron. Adv. Mater.* **2007**, *9*, 942–945.
565. Alexiou, C.; Arnold, W.; Klein, R.J.; Parak, F.G.; Hulin, P.; Bergemann, C.; Erhardt, W.; Wagenpfeil, S.; Lubbe, A.S. Locoregional cancer treatment with magnetic drug targeting. *Cancer Res.* **2000**, *60*, 6641–6648.
566. Wahajuddin; Arora, S. Superparamagnetic iron oxide nanoparticles: Magnetic nanoplatforms as drug carriers. *Int. J. Nanomed.* **2012**, *7*, 3445–3471. [[CrossRef](#)]
567. Mansouri, M.; Nazarpak, M.H.; Solouk, A.; Akbari, S.; Hasani-Sadrabadi, M.M. Magnetic responsive of paclitaxel delivery system based on spion and palmitoyl chitosan. *J. Magn. Magn. Mater.* **2017**, *421*, 316–325. [[CrossRef](#)]
568. Yang, Y.; Guo, Q.F.; Peng, J.R.; Su, J.; Lu, X.L.; Zhao, Y.X.; Qian, Z.Y. Doxorubicin-conjugated heparin-coated superparamagnetic iron oxide nanoparticles for combined anticancer drug delivery and magnetic resonance imaging. *J. Biomed. Nanotechnol.* **2016**, *12*, 1963–1974. [[CrossRef](#)] [[PubMed](#)]
569. Mattingly, S.J.; O'Toole, M.G.; James, K.T.; Clark, G.J.; Nantz, M.H. Magnetic nanoparticle-supported lipid bilayers for drug delivery. *Langmuir* **2015**, *31*, 3326–3332. [[CrossRef](#)] [[PubMed](#)]
570. Ghorbani, M.; Hamishehkar, H.; Arsalani, N.; Entezami, A.A. Preparation of thermo and ph-responsive polymer@au/fe₃o₄ core/shell nanoparticles as a carrier for delivery of anticancer agent. *J. Nanopart. Res.* **2015**, *17*, 13. [[CrossRef](#)]
571. Song, W.X.; Muthana, M.; Mukherjee, J.; Falconer, R.J.; Biggs, C.A.; Zhao, X.B. Magnetic-silk core-shell nanoparticles as potential carriers for targeted delivery of curcumin into human breast cancer cells. *ACS Biomater. Sci. Eng.* **2017**, *3*, 1027–1038. [[CrossRef](#)]
572. Rotariu, O.; Udrea, L.E.; Strachan, N.J.C.; Badescu, V. Targeting magnetic carrier particles in tumour microvasculature—A numerical study. *J. Optoelectron. Adv. Mater.* **2005**, *7*, 3209–3218.
573. Norris, M.D.; Seidel, K.; Kirschning, A. Externally induced drug release systems with magnetic nanoparticle carriers: An emerging field in nanomedicine. *Adv. Ther.* **2019**, *2*, 1800092. [[CrossRef](#)]
574. Ahmad, R.S.; Ali, Z.S.; Mou, X.B.; Wang, J.H.; Yi, H.; He, N.Y. Recent advances in magnetic nanoparticle design for cancer therapy. *J. Nanosci. Nanotechnol.* **2016**, *16*, 9393–9403. [[CrossRef](#)]
575. Ramazanov, M.; Karimova, A.; Shirinova, H. Magnetism for drug delivery, MRI and hyperthermia applications: A review. *Biointerface Res. Appl. Chem.* **2021**, *11*, 8654–8668.
576. Yang, H.Y.; Li, Y.; Lee, D.S. Multifunctional and stimuli-responsive magnetic nanoparticle-based delivery systems for biomedical applications. *Adv. Ther.* **2018**, *1*, 1800011. [[CrossRef](#)]
577. Schneider-Futschik, E.K.; Reyes-Ortega, F. Advantages and disadvantages of using magnetic nanoparticles for the treatment of complicated ocular disorders. *Pharmaceutics* **2021**, *13*, 1157. [[CrossRef](#)] [[PubMed](#)]
578. Zamay, T.N.; Prokopenko, V.S.; Zamay, S.S.; Lukyanenko, K.A.; Kolovskaya, O.S.; Orlov, V.A.; Zamay, G.S.; Galeev, R.G.; Narodov, A.A.; Kichkailo, A.S. Magnetic nanodiscs—a new promising tool for microsurgery of malignant neoplasms. *Nanomaterials* **2021**, *11*, 1459. [[CrossRef](#)]
579. Golovin, Y.I.; Golovin, D.Y.; Vlasova, K.Y.; Veselov, M.M.; Usvaliev, A.D.; Kabanov, A.V.; Klyachko, N.L. Non-heating alternating magnetic field nanomechanical stimulation of biomolecule structures via magnetic nanoparticles as the basis for future low-toxic biomedical applications. *Nanomaterials* **2021**, *11*, 2255. [[CrossRef](#)]
580. Chen, M.W.; Wu, J.J.; Ning, P.; Wang, J.J.; Ma, Z.; Huang, L.Q.; Plaza, G.R.; Shen, Y.J.; Xu, C.; Han, Y.; et al. Remote control of mechanical forces via mitochondrial-targeted magnetic nanospinners for efficient cancer treatment. *Small* **2020**, *16*, 14. [[CrossRef](#)] [[PubMed](#)]
581. Alsharif, N.A.; Aleisa, F.A.; Liu, G.Y.; Ooi, B.S.; Patel, N.; Ravasi, T.; Merzaban, J.S.; Kosel, J. Functionalization of magnetic nanowires for active targeting and enhanced cell-killing efficacy. *ACS Appl. Bio Mater.* **2020**, *3*, 4789–4797. [[CrossRef](#)]
582. Chen, Y.; Han, P.; Wu, Y.; Zhang, Z.F.; Yue, Y.; Li, W.H.; Chu, M.Q. Hedgehog-like gold-coated magnetic microspheres that strongly inhibit tumor growth through magnetomechanical force and photothermal effects. *Small* **2018**, *14*, 13. [[CrossRef](#)]
583. Goirienna-Goikoetxea, M.; Munoz, D.; Orue, I.; Fernandez-Gubieda, M.L.; Bokor, J.; Muela, A.; Garcia-Arribas, A. Disk-shaped magnetic particles for cancer therapy. *Appl. Phys. Rev.* **2020**, *7*, 15. [[CrossRef](#)]

584. Rahban, D.; Doostan, M.; Salimi, A. Cancer therapy; prospects for application of nanoparticles for magnetic-based hyperthermia. *Cancer Investig.* **2020**, *38*, 507–521. [[CrossRef](#)]
585. Kok, H.P.; Cressman, E.N.K.; Ceelen, W.; Brace, C.L.; Ivkov, R.; Grull, H.; ter Haar, G.; Wust, P.; Crezee, J. Heating technology for malignant tumors: A review. *Int. J. Hyperth.* **2020**, *37*, 711–741. [[CrossRef](#)]
586. Liu, X.; Zhang, Y.; Wang, Y.; Zhu, W.; Li, G.; Ma, X.; Zhang, Y.; Chen, S.; Tiwari, S.; Shi, K.; et al. Comprehensive understanding of magnetic hyperthermia for improving antitumor therapeutic efficacy. *Theranostics* **2020**, *10*, 3793–3815. [[CrossRef](#)]
587. Chang, D.; Lim, M.; Goos, J.; Qiao, R.; Ng, Y.Y.; Mansfeld, F.M.; Jackson, M.; Davis, T.P.; Kavallaris, M. Biologically targeted magnetic hyperthermia: Potential and limitations. *Front. Pharm.* **2018**, *9*, 831. [[CrossRef](#)]
588. Frazier, N.; Ghandehari, H. Hyperthermia approaches for enhanced delivery of nanomedicines to solid tumors. *Biotechnol. Bioeng.* **2015**, *112*, 1967–1983. [[CrossRef](#)] [[PubMed](#)]
589. Lee, Y.K.; Lee, S.B.; Kim, Y.U.; Kim, K.N.; Choi, S.Y.; Lee, K.H.; Shim, I.B.; Kim, C.S. Effect of ferrite thermoseeds on destruction of carcinoma cells under alternating magnetic field. *J. Mater. Sci.* **2003**, *38*, 4221–4233. [[CrossRef](#)]
590. Shido, Y.; Nishida, Y.; Suzuki, Y.; Kobayashi, T.; Ishiguro, N. Targeted hyperthermia using magnetite cationic liposomes and an alternating magnetic field in a mouse osteosarcoma model. *J. Bone Jt. Surg. Br. Vol.* **2010**, *92B*, 580–585. [[CrossRef](#)]
591. Manohar, A.; Vijayakanth, V.; Pallavolu, M.R.; Kim, K.H. Effects of Ni—Substitution on structural, magnetic hyperthermia, photocatalytic and cytotoxicity study of MgFe₂O₄ nanoparticles. *J. Alloys Compd.* **2021**, *879*, 160515. [[CrossRef](#)]
592. Chen, S.Z.; Han, F.S.; Huang, D.D.; Meng, J.Q.; Chu, J.P.; Wang, M.; Wang, P.J. Fe₃O₄ magnetic nanoparticle-enhanced radiotherapy for lung adenocarcinoma via delivery of siBIRC5 and AS-ODN. *J. Transl. Med.* **2021**, *19*, 1–14. [[CrossRef](#)]
593. Jiang, K.Y.; Zhang, Q.B.; Hinojosa, D.T.; Zhang, L.L.; Xiao, Z.; Yin, Y.; Tong, S.; Colvin, V.L.; Bao, G. Controlled oxidation and surface modification increase heating capacity of magnetic iron oxide nanoparticles. *Appl. Phys. Rev.* **2021**, *8*, 031407. [[CrossRef](#)]
594. Simeonidis, K.; Kaprara, E.; Rivera-Gil, P.; Xu, R.X.; Teran, F.J.; Kokkinos, E.; Mitropoulos, A.; Maniotis, N.; Balcells, L. Hydrotalcite-embedded magnetite nanoparticles for hyperthermia-triggered chemotherapy. *Nanomaterials* **2021**, *11*, 1796. [[CrossRef](#)]
595. Kawashita, M.; Domi, S.; Saito, Y.; Aoki, M.; Ebisawa, Y.; Kokubo, T.; Saito, T.; Takano, M.; Araki, N.; Hiraoka, M. In vitro heat generation by ferrimagnetic maghemite microspheres for hyperthermic treatment of cancer under an alternating magnetic field. *J. Mater. Sci. Mater. Med.* **2008**, *19*, 1897–1903. [[CrossRef](#)]
596. Tietze, R.; Zaloga, J.; Unterweger, H.; Lyer, S.; Friedrich, R.P.; Janko, C.; Pottler, M.; Durr, S.; Alexiou, C. Magnetic nanoparticle-based drug delivery for cancer therapy. *Biochem. Biophys. Res. Commun.* **2015**, *468*, 463–470. [[CrossRef](#)]
597. Zhi, D.F.; Yang, T.; Yang, J.; Fu, S.; Zhang, S.B. Targeting strategies for superparamagnetic iron oxide nanoparticles in cancer therapy. *Acta Biomater.* **2020**, *102*, 13–34. [[CrossRef](#)]
598. Uskokovic, V.; Drofenik, M. Gold-embellished mixed-valence manganite as a smart, self-regulating magnetoplasmonic nanomaterial. *Mater. Chem. Phys.* **2021**, *271*, 124870. [[CrossRef](#)]
599. Rodrigues, H.F.; Capistrano, G.; Bakuzis, A.F. In vivo magnetic nanoparticle hyperthermia: A review on preclinical studies, low-field nano-heaters, noninvasive thermometry and computer simulations for treatment planning. *Int. J. Hyperth.* **2020**, *37*, 76–99. [[CrossRef](#)] [[PubMed](#)]
600. Rotariu, O.; Iacob, G.; Strachan, N.J.C.; Chiriac, H. Simulating the embolization of blood vessels using magnetic microparticles and acupuncture needle in a magnetic field. *Biotechnol. Prog.* **2004**, *20*, 299–305. [[CrossRef](#)] [[PubMed](#)]
601. Anani, T.; Rahmati, S.; Sultana, N.; David, A.E. MRI-traceable theranostic nanoparticles for targeted cancer treatment. *Theranostics* **2021**, *11*, 579–601. [[CrossRef](#)]
602. Kozissnik, B.; Bohorquez, A.C.; Dobson, J.; Rinaldi, C. Magnetic fluid hyperthermia: Advances, challenges, and opportunity. *Int. J. Hyperth.* **2013**, *29*, 706–714. [[CrossRef](#)]
603. Xie, J.; Yan, C.; Yan, Y.; Chen, L.; Song, L.; Zang, F.; An, Y.; Teng, G.; Gu, N.; Zhang, Y. Multi-modal mn–zn ferrite nanocrystals for magnetically-induced cancer targeted hyperthermia: A comparison of passive and active targeting effects. *Nanoscale* **2016**, *8*, 16902–16915. [[CrossRef](#)]
604. Angelakeris, M. Magnetic nanoparticles: A multifunctional vehicle for modern theranostics. *Biochim. Biophys. Acta Gen. Subj.* **2017**, *1861*, 1642–1651. [[CrossRef](#)] [[PubMed](#)]
605. Cheng, H.W.; Tsao, H.Y.; Chiang, C.S.; Chen, S.Y. Advances in magnetic nanoparticle-mediated cancer immune-theranostics. *Adv. Healthc. Mater.* **2021**, *10*, 2001451. [[CrossRef](#)]
606. Rajan, A.; Sahu, N.K. Review on magnetic nanoparticle-mediated hyperthermia for cancer therapy. *J. Nanopart. Res.* **2020**, *22*, 1–25. [[CrossRef](#)]
607. Soetaert, F.; Korangath, P.; Serantes, D.; Fiering, S.; Ivkov, R. Cancer therapy with iron oxide nanoparticles: Agents of thermal and immune therapies. *Adv. Drug Deliv. Rev.* **2020**, *163*, 65–83. [[CrossRef](#)]
608. Li, X.X.; Li, W.Y.; Wang, M.N.; Liao, Z.H. Magnetic nanoparticles for cancer theranostics: Advances and prospects. *J. Control. Release* **2021**, *335*, 437–448. [[CrossRef](#)] [[PubMed](#)]
609. Mohammed, L.; Gomaa, H.G.; Ragab, D.; Zhu, J. Magnetic nanoparticles for environmental and biomedical applications: A review. *Particuology* **2017**, *30*, 1–14. [[CrossRef](#)]
610. Li, Y.H.; Wang, N.; Huang, X.M.; Li, F.Y.; Davis, T.P.; Qiao, R.R.; Ling, D.S. Polymer-assisted magnetic nanoparticle assemblies for biomedical applications. *ACS Appl. Bio Mater.* **2020**, *3*, 121–142. [[CrossRef](#)] [[PubMed](#)]

611. Pooam, M.; Jourdan, N.; El Esawi, M.; Sherrard, R.M.; Ahmad, M. Hek293 cell response to static magnetic fields via the radical pair mechanism may explain therapeutic effects of pulsed electromagnetic fields. *PLoS ONE* **2020**, *15*, e0243038. [[CrossRef](#)]
612. Chatterjee, R.; Chatterjee, J. Ros and oncogenesis with special reference to emt and stemness. *Eur. J. Cell Biol.* **2020**, *99*, 8. [[CrossRef](#)]