The Danish Pesticide Leaching Assessment Programme

Monitoring results May 1999–June 2010

Jeanne Kjær, Annette E. Rosenbom, Walter Brüsch, René K. Juhler, Lasse Gudmundsson, Finn Plauborg, Ruth Grant and Preben Olsen

Geological Survey of Denmark and Greenland Ministry of Climate and Energy

Department of Agroecology Aarhus University

Department of Bioscience Aarhus University

Editor: Jeanne Kjær *Cover photo:* Lasse Gudmundsson *Cover:* Henrik Klinge Pedersen *Layout and graphic production*: Authors *Printed:* September 2011 *Price:* DKK 200

ISBN 978-87-7871-312-4

Available from:

Geological Survey of Denmark and Greenland Øster Voldgade 10, DK-1350 Copenhagen K, Denmark Phone: +45 38 14 20 00. Fax: +45 38 14 20 50 E-mail: geus@geus.dk Homepage: www.geus.dk

The report is also available as a pdf file at www.pesticidvarsling.dk

© De Nationale Geologiske Undersøgelser for Danmark og Grønland, 2011

Table of contents

PREFACE

SI D	JMMARY ANSK SAI	MMENDRAG	
1	INTR	ODUCTION	13
1	1.1		10
	1.1 C 1.2 S	DBJECTIVE	13
2	PESTI	ICIDE LEACHING AT TVI STRUP	17
-	2.1		17
	2.1 N	VIATERIALS AND METHODS	1/ 17
	2.1.1 2.1.2	A oricultural management	18
	2.1.2	Model setup and calibration	.18
	2.2 F	RESULTS AND DISCUSSION	19
	2.2.1	Soil water dynamics and water balances	. 19
	2.2.2	Bromide leaching	21
	2.2.3	Pesticide leaching	. 23
3	PESTI	ICIDE LEACHING AT JYNDEVAD	29
	3.1 N	ATERIALS AND METHODS	29
	3.1.1	Site description and monitoring design	29
	3.1.2	Agricultural management	29
	3.1.3	Model setup and calibration	. 31
	3.2 F	Results and discussion	33
	3.2.1	Soil water dynamics and water balances	33
	3.2.2	Bromide leaching	. 34
	5.2.5		. 50
4	PESTI	ICIDE LEACHING AT SILSTRUP	43
	4.1 N	MATERIALS AND METHODS	43
	4.1.1	Site description and monitoring design	43
	4.1.2	Agricultural management	43
	4.1.3 4.2 I	Model setup and caubration	45
	4.2 r 421	Soil water dynamics and water balances	40
	4.2.2	Bromide leaching	49
	4.2.3	Pesticide leaching	50
5	PESTI	ICIDE LEACHING AT ESTRUP	61
	51 N		61
	5.1.1	Site description and monitoring design	61
	5.1.2	Agricultural management	61
	5.1.3	Model setup and calibration	63
	5.2 F	RESULTS AND DISCUSSION	63
	5.2.1	Soil water dynamics and water balances	. 63
	5.2.2	Bromide leaching	66
	5.2.3	Pesticide leaching	
6	PESTI	ICIDE LEACHING AT FAARDRUP	77
	6.1 N	MATERIALS AND METHODS	77
	6.1.1	Site description and monitoring design	77
	6.1.2	Agricultural management	.80
	0.1.3 6.2 I	model setup and calibration	. 0U . 0U
	0.∠ F 621	Soil water dynamics and water balances	01
	6.2.2	Bromide leaching	. 83
	6.2.3	Pesticide leaching	85
		-	

7	PESTICIDE ANALYSIS QUALITY ASSURANCE	
	7.1 MATERIALS AND METHODS	
	7.1.1 Internal QA	
	7.1.2 External QA	
	7.2 Results and discussion	
	7.2.1 Internal QA	
	7.2.2 External QA	
	7.3 SUMMARY AND CONCLUDING REMARKS	94
8	SUMMARY OF MONITORING RESULTS	95
9	REFERENCES	

APPENDIX 1. CHEMICAL ABSTRACTS NOMENCLATURE FOR THE PESTICIDES ENCOMPASSED BY THE PLAP.

APPENDIX 2. PESTICIDE MONITORING PROGRAMME - SAMPLING PROCEDURE.

APPENDIX 3. AGRICULTURAL MANAGEMENT.

APPENDIX 4. PRECIPITATION DATA FOR THE PLAP SITES.

APPENDIX 5. PESTICIDE DETECTION IN SAMPLES FROM DRAINAGE SYSTEM, SUCTION CUPS AND MONITORING SCREENS.

APPENDIX 6. LABORATORY INTERNAL CONTROL CARDS.

Preface

In 1998, the Danish Parliament initiated the Danish Pesticide Leaching Assessment Programme (PLAP), an intensive monitoring programme aimed at evaluating the leaching risk of pesticides under field conditions. The Danish Government funded the first phase of the programme from 1998 to 2001. The programme has now been prolonged twice, initially with funding from the Ministry of the Environment and the Ministry of Food, Agriculture and Fisheries for the period 2002 to 2009, and presently with funding from the Danish Environmental Protection Agency for the period 2010 to 2015.

The work was conducted by the Geological Survey of Denmark and Greenland (GEUS), the Department of Agroecology (DJF) at Aarhus University and the Department of Bioscience (NERI), Aarhus University under the direction of a management group comprising Jeanne Kjær (GEUS), Annette E. Rosenbom (GEUS), Walter Brüsch (GEUS), Lis Wollesen de Jonge (DJF), Preben Olsen (DJF), Ruth Grant (NERI) and Steen Marcher (Danish Environmental Protection Agency).

This report presents the results for the period May 1999–June 2010. Results covering part of the period May 1999–June 2009 have been reported previously (Kjær *et al.*, 2002, Kjær *et al.*, 2003, Kjær *et al.*, 2004, Kjær *et al.*, 2005c, Kjær *et al.*, 2007, Kjær *et al.*, 2008, Kjær *et al.*, 2009, and Rosenbom *et al.*, 2010b). The present report should therefore be seen as a continuation of previous reports with the main focus on the leaching risk of pesticides applied during 2008.

The report was prepared jointly by Annette E. Rosenbom, Walter Brüsch, René K. Juhler, Jeanne Kjær, and Lasse Gudmundsson (all GEUS), Preben Olsen, and Finn Plauborg (DJF), and Ruth Grant (NERI). While all authors contributed to the whole report, authors were responsible for separate aspects as follows:

- Pesticide and bromide leaching: Walter Brüsch, Preben Olsen and Jeanne Kjær.
- Soil water dynamics and water balances: Annette E. Rosenborn, Finn Plauborg, and Ruth Grant.
- Pesticide analysis quality assurance: René K. Juhler.

Jeanne Kjær September 2011

Summary

In 1998, the Danish Parliament initiated the Pesticide Leaching Assessment Programme (PLAP), an intensive monitoring programme aimed at evaluating the leaching risk of pesticides under field conditions. The objective of the PLAP is to improve the scientific foundation for decision-making in the Danish regulation of pesticides. The specific aim is to analyse whether pesticides applied in accordance with current regulations leach to groundwater in unacceptable concentrations. The programme currently evaluates the leaching risk of 42 pesticides and 41 degradation products at five agricultural sites ranging in size from 1.1 to 2.4 ha. The evaluation is based upon monitoring results representing detections in 1 meters depth (water collected via drains and suction cups) and detections in groundwater monitoring screens (1.5-4.5 meter below ground surface, hereafter m b.g.s.). This report presents the results for the entire monitoring period May 1999–June 2010. Results covering part of the period May 1999–June 2009 have been reported previously.

Highlights from **monitoring period 2009-2010** where **6** pesticides were applied, show that:

- Bifenox acid (degradation product of bifenox) can on loamy soil leach through the root zone and enter both drainage water system and groundwater monitoring wells in concentrations exceeding 0.1 μ g/l. Average concentration in the drainage water exceeded 0.1 μ g/l and concentrations exceeding 0.1 μ g/l were observed up to six months after application. Similar evidence of pronounced leaching was **not** observed on the coarse sandy soil as there was only a single detection of bifenox acid in soil water, whereas bifenox was detected very sporadically in soil and groundwater, concentrations always less than 0.1 μ g/l.
- Ethofumesate, this year used in a new admissible dose that is five times lower than in past applications, was detected once in groundwater, concentrations at less than $0.1 \mu g/l$. When, in the past (before the imposed regulation), ethofumesate was used at a much higher dose, leaching above $0.1 \mu g/l$ to both drains and groundwater monitoring wells was observed.
- Metamitron, this year used in a dose 33% lower than the permitted, did not cause leaching above 0.1 μ g/l of either metamitron or its metabolite metamitron-desamino. When, in the past, metamitron was used at maximum allowed dose, leaching above 0.1 μ g/l to both drainage system and groundwater monitoring wells was observed. It is not possible to say if the low leaching this year is related to the climatic conditions/timing or the reduced dose.
- The leaching pattern of the remaining three pesticides (bentazone, azoxystrobin, triasulfuron) was in line with the previous observations (outlined below).

The results of the **entire monitoring period 1999-2010** covering 42 pesticides, show that:

- Of the 42 pesticides applied, 11 pesticides and/or their degradation product(s) (clopyralid, chlormequat, desmedipham, fenpropimorph, florasulam, iodosulfuron-methyl-sodium, linuron, metsulfuron-methyl, thiamethoxam, tribenuron-methyl, and triasulfuron) did not leach during the entire monitoring period.
- The monitoring data indicate pronounced leaching of 14 of the applied pesticides and/or their degradation products. The following compounds leached through the soil entering drains and suction cups (placed 1 m b.g.s) in average concentrations exceeding 0.1 µg/l:
 - o *azoxystrobin* and its degradation product *CyPM*,
 - o *bentazone*
 - *CL153815* (degradation product of picolinafen)
 - o *pirimicarb-desmethyl-formamido* (degradation product of pirimicarb)
 - o propyzamide
 - o *tebuconazole*
 - *glyphosate* and its degradation product *AMPA*
 - **PPU** (degradation products of rimsulfuron)
 - **Bifenox acid** (degradation product of bifenox)
 - o ethofumesate
 - **TFMP** (degradation product of fluazifop-P-butyl)
 - *metamitron* and its degradation product **metamitron-desamino**
 - **metribuzin-desamino-diketo** and **metribuzin-diketo** (degradation products of metribuzin)
 - **terbuthylazine** and its degradation products: **desethyl-terbuthylazine**, 2*hydroxy-desethyl-terbuthylazine*, and 2-*hydroxy-terbuthylazine*

For the pesticides and/or their degradation products marked in Italics, pronounced leaching is mainly confined to the depth of 1 meter, where pesticides were frequently found in samples collected from drains and suction cups, while a limited number of detections (fewer than 5 samples per field) exceeding 0.1 µg/l were found in groundwater monitoring wells. For the pesticides and/or their degradation products marked in bold, pronounced leaching below the depth of 1 m was observed. Apart from PPU, these were all frequently detected in concentrations exceeding 0.1 μ g/l in groundwater monitoring wells, exceedance of 0.1 μ g/l being observed more than six months after application. Although PPU was only detected in a few samples in concentrations exceeding 0.1 µg/l, elevated concentrations just below 0.1 µg/l were found in groundwater monitoring wells during a twoyear period, thus confirming the pronounced leaching and high persistency of PPU in soil and groundwater. Repeated applications of PPU may thus pose a contamination risk of the shallow groundwater. Moreover, for the glyphosate being frequently applied on one loamy soil, detections in groundwater monitoring wells have gradually increased over time. On two occasions heavy rain events and snowmelt induced leaching to the groundwater monitoring wells in concentrations exceeding 0.1 µg/l more than two years after the application.

• The monitoring data also indicate leaching of an additional 17 pesticides, but in low concentrations. Although concentrations exceeded 0.1 μ g/l in several samples collected from suction cups and drains (1 m b.g.s.), average leaching concentrations on a yearly basis did not. None of the compounds were found in groundwater monitoring wells in concentrations exceeding 0.1 μ g/l.

The PLAP initially evaluated the leaching risk at six agricultural sites representing a range of Danish soil and climate conditions. Monitoring at the Slaeggerup site was terminated on 1 July 2003, and results from that site are not included in the present report. For the monitoring results from this site see Kjær *et al.* (2004).

In order to describe water transport, a bromide tracer was applied to the fields. Bromide and pesticide concentrations are measured monthly in both the unsaturated and the saturated zones, and weekly in the drainage water. This report covers the period May 1999–June 2010 and presents the monitoring results from the five agricultural sites presently monitored. The main focus is on evaluating the leaching risk of the pesticides applied during 2008.

Dansk sammendrag

I 1998 vedtog Folketinget at iværksætte projektet "Varslingssystem for udvaskning af Pesticider til grundvandet" (VAP). VAP er et omfattende moniteringsprogram, der undersøger udvaskning af pesticider anvendt i landbrug under reelle markforhold. Programmet har til formål at undersøge, om godkendte pesticider eller deres nedbrydningsprodukter - ved regelret brug - udvaskes til grundvandet i koncentrationer over grænseværdien for herigennem at udvide det videnskabelige grundlag for danske myndigheders (Miljøstyrelsen) procedurer for regulering af sprøjtemidler. Udvaskningsrisikoen for 42 pesticider og 41 nedbrydningsprodukter er således op til i dag undersøgt på fem marker, der har en størrelse på mellem 1,1 og 2,4 ha. Undersøgelsen bygger på moniteringsresultater henholdsvis repræsenterende fund i en meters dybde (indhentet via dræn og sugeceller) og fund i grundvandsmoniteringsfiltre (1.5-4.5 meter under terræn, herefter m u.t.). Denne rapport opsummerer resultaterne for den samlede moniteringsperiode maj 1999 - juni 2010; resultater fra de foregående år maj 1999 – juni 2009 er blevet afrapporteret i de tidligere rapporter.

"Highlights" fra den seneste moniteringsperiode **2009-2010**, hvor **6** pesticider blev udbragt, viser følgende:

- Bifenox-syre (nedbrydningsprodukt af bifenox) blev på lerjorde udvasket fra rodzonen (1 m.u.t.) til både dræn og grundvandsfiltre i koncentrationer over 0,1 µg/l. I drænene oversteg den årlige middelkoncentration 0,1 µg/l, og i grundvandsfiltrene blev der på en ud af to testede lerjorde fundet koncentrationer over 0,1 µg/l op til seks måneder efter udbringning. Tilsvarende udvaskning blev ikke set på den grovsandede jord, hvor der kun var et enkelt fund af bifenox-syre (i jordvandet) samt sporadiske fund af bifenox i lave koncentrationer (under 0,1 µg/l).
- Ethofumesat, der som følge af Miljøstyrelsens restriktioner dette år blev udbragt i en dosis 5 gange lavere end tidligere, blev denne gang kun fundet i en enkelt prøve (<0,1 μg/l). Ved tidligere anvendelser med den høje dosering, blev ethofumesat udvasket til både dræn og grundvandsfiltre i koncentrationer over 0,1 μg/l.
- Metamitron, som dette år blev udbragt i den anbefalede dosis, hvilket var 33% mindre end den maximalt tilladte mængde, blev ikke udvasket i koncentrationer, der oversteg 0,1 μ g/l. Ved tidligere udbringninger (ved den maximalt tilladte dosis) blev metamitron og dets nedbrydningsprodukt metamitron-desamino udvasket i koncentrationer der oversteg 0,1 μ g/l i både dræn og grundvandsfiltre. Det er imidlertid ikke til at sige om den mindskede udvaskning dette år skyldes den lavere dosering eller de klimatiske forhold.
- For de øvrige 3 stoffer (bentazon, azoxystrobin, triasulfuron) var de observerede udvaskningsforløb meget lig tidligere observationer (beskrevet nedenfor).

Resultater for hele **moniteringsperioden 1999-2010**, som omfatter **42** pesticider viser følgende:

- Af de 42 pesticider, der er blevet udbragt, blev 11 pesticider eller nedbrydningsprodukter heraf (clopyralid, chlormequat, desmedipham, fenpropimorph, florasulam, iodosulfuron-methyl-natrium, linuron, metsulfuron-methyl, thiamethoxam, tribenuronmethyl og triasulfuron) **ikke** fundet udvasket i løbet af den samlede moniteringsperiode.
- 14 af de udbragte stoffer eller nedbrydningsprodukter heraf gav anledning til en markant udvaskning. Følgende stoffer blev udvasket til dræn og sugeceller, beliggende i ca. 1 meters dybde i gennemsnitskoncentrationer over 0,1 µg/l:
 - o *azoxystrobin* og dets nedbrydningsprodukt *CyPM*
 - o *bentazon*
 - o *CL153815* (nedbrydningsprodukt af picolinafen)
 - o *pirimicarb-desmethyl-formamido* (nedbrydningsprodukt af pirimicarb)
 - o propyzamid
 - o tebuconazol
 - o glyphosat og dets nedbrydningsprodukt AMPA
 - **PPU** (nedbrydningsprodukt af rimsulfuron)
 - **bifenox-syre** (nedbrydningsprodukt af bifenox)
 - ethofumesat
 - **TFMP** (nedbrydningsprodukt af fluazifop-P-butyl),
 - o metamitron og dets nedbrydningsprodukt metamitron-desamino
 - **metribuzin-desamino-diketo** og **metribuzin-diketo** (nedbrydningsprodukter af metribuzin)
 - **terbuthylazin** og dets nedbrydningsprodukter **desethyl-terbuthylazin**, *2-hydroxy-desethyl-terbuthylazin*, *and 2-hydroxy-terbuthylazin*

For de pesticider eller nedbrydningsprodukter heraf fremhævet med kursiv var udvaskningen primært begrænset til 1 m.u.t., hvor de blev fundet hyppigt i dræn og sugeceller. Selvom hovedparten af stofferne blev fundet i koncentrationer over 0,1 µg/l i grundvandsfiltrene, var antallet af overskridelser få (mindre end 5 pr. mark). Pesticider markeret med fed blev derimod udvasket til grundvandsfiltrene i en større grad. På nær PPU blev samtlige stoffer relativt hyppigt fundet i koncentrationer over 0,1 µg/l i grundvandsfiltrene, hvor koncentrationer over 0,1 µg/l blev fundet mere end seks måneder efter udbringning. Om end det kun var enkelte prøver som indeholdt mere end 0,1 µg/l PPU, blev der igennem en toårig periode fundet PPU i grundvandet i koncentrationer tæt på de 0,1 µg/l, hvilket bekræfter den høje persistens af PPU i jord og grundvand. Gentagne udbringninger af PPU kan potentielt forurene det allerøverste grundvand. Glyphosat er blevet udbragt flere gange på en af de lerede forsøgslokaliteter. På denne mark er der igennem de seneste år konstateret et stigende antal fund af glyphosat i grundvandsfiltrene. To gange har markante nedbørshændelse samt snesmeltning forårsaget udvaskning af glyphosat til grundvandfiltrene i koncentrationer over 0,1 µg/l mere end to år efter udbringning.

 Andre 17 stoffer gav anledning til udvaskning. Selv om flere af disse stoffer i én meters dybde ofte blev fundet i koncentrationer over 0,1 µg/l, var der ikke tale om, at udvaskningen som årsmiddel oversteg 0,1 µg/l. Stofferne blev heller ikke fundet i grundvandsfiltrene i koncentration over 0.1 µg/L. VAP-programmet omfattede oprindeligt seks marker placeret, så de repræsenterer forskellige typer geologi og tillige tager hensyn til de klimatiske variationer i Danmark, specielt hvad angår nedbørforhold. Monitering på den ene forsøgsmark (Slæggerup) stoppede 1. juli, 2003. Resultater fra denne mark er ikke inkluderet i denne rapport, men kan findes i Kjær *et al.* (2004). De anvendte pesticider bliver udbragt i maksimalt tilladte doser. Bromid anvendes som sporstof for at beskrive vandtransporten. Bromidog pesticidkoncentrationer bliver analyseret månedligt i prøver udtaget i den umættede og mættede zone og ugentligt i prøver af drænvand. I denne rapport præsenteres moniteringsresultaterne for de fem områder for perioden maj 1999 - juni 2010 primært med fokus på pesticider udbragt i 2008. En del af stofferne har kun været inkluderet i konkludere noget endeligt.

1 Introduction

There is growing public concern in Denmark about pesticide contamination of our surface waters and groundwater. Pesticides and their degradation products have increasingly been detected in groundwater during the past decade and are now present in much of the Danish groundwater. Under the Danish National Groundwater Monitoring Programme (GRUMO) pesticides have so far been detected in 53% of all screens monitored and in 61% of the screens placed in the upper groundwater (Thorling, L. (red), 2010).

The increasing detection of pesticides in groundwater over the past 10 years has given rise to the desire to enhance the scientific foundation for the existing approval procedure for pesticides and to improve the present risk assessment tools. A main issue in this respect is that the EU assessment and hence also the Danish assessment of the risk of pesticide leaching to groundwater is largely based on data from modelling, laboratory or lysimeter studies. However, these types of data may not adequately describe the leaching that may occur under actual field conditions. Although models are widely used within the registration process, their validation requires further work, not least because of the limited availability of field data (Boesten, 2000). Moreover, laboratory and lysimeter studies do not include the spatial variability of the soil parameters (hydraulic, chemical, physical and microbiological soil properties) affecting pesticide leaching. This is of particular importance for silty and loamy soils, where preferential transport may have a major impact on pesticide leaching. In fact, various field studies suggest that considerable preferential transport of several pesticides occurs to a depth of 1 m under conditions comparable to those pertaining in Denmark (Kördel, 1997).

The inclusion of field studies, i.e. test plots exceeding 1 ha, in risk assessment of pesticide leaching to groundwater is considered an important improvement to the risk assessment procedures. For example, the US Environmental Protection Agency (US-EPA) has included field-scale studies in its risk assessments since 1987. Pesticides that may potentially leach to the groundwater are required to be included in field studies as part of the registration procedure. The US-EPA has therefore conducted field studies on more than 50 pesticides (US Environmental Protection Agency, 1998). A similar concept has also been adopted within the European Union (EU), where Directive 91/414/EEC, Annexe VI (Council Directive 97/57/EC of 22 September 1997) enables field leaching study results to be included in the risk assessments.

1.1 Objective

In 1998, the Danish Government initiated the Pesticide Leaching Assessment Programme (PLAP), an intensive monitoring programme with the purpose of evaluating the leaching risk of pesticides under field conditions. The PLAP is intended to serve as an early warning system providing decision-makers with advance warning if approved pesticides leach in unacceptable concentrations. The programme focuses on pesticides used in arable farming and monitors leaching at five agricultural test sites representative of Danish conditions.

The objective of the PLAP is to improve the scientific foundation for decision-making in the Danish registration and approval procedures for pesticides, enabling field studies to be included in risk assessment of selected pesticides. The specific aim is to analyse whether pesticides applied in accordance with current regulations leach at levels exceeding the maximum allowable concentration of $0.1 \,\mu g/l$.

1.2 Structure of the PLAP

The pesticides included in the PLAP were selected by the Danish Environmental Protection Agency on the basis of expert judgement. At present, 42 pesticides and 41 degradation products are included in the PLAP. All the compounds analysed are listed in Appendix 1.



Figure 1. Location of the PLAP sites Tylstrup, Jyndevad, Silstrup, Estrup, and Faardrup. Monitoring at Slaeggerup was terminated on 1 July 2003.

Soil type and climatic conditions are considered to be some of the most important parameters controlling pesticide leaching. The PLAP initially encompassed six test sites representative of the dominant soil types and the climatic conditions in Denmark (Figure 1). Monitoring at the Slaeggerup site was terminated on 1 July 2003, and results from that site are not included in the present report. For the monitoring results from this site see Kjær et al. (2003). The groundwater table at all the sites is shallow, thereby enabling pesticide leaching to groundwater to be rapidly detected (Table 1). Cultivation of the PLAP sites is in line with conventional agricultural practice in the vicinity. The pesticides are applied at maximum permitted doses and in the manner specified in the regulations. Hence any pesticides or degradation products appearing in the groundwater downstream of the sites can be related to the current approval conditions pertaining to the individual pesticides. The PLAP was initiated in autumn 1998. The five test sites encompassed by the present report were selected and established during 1999. Monitoring was initiated at Tylstrup, Jyndevad and Faardrup in 1999 and at Silstrup and Estrup in 2000 (Table 1).

	Tylstrup	Jyndevad	Silstrup	Estrup	Faardrup
Location	Brønderslev	Tinglev	Thisted	Askov	Slagelse
Precipitation ¹⁾ (mm/y)	668	858	866	862	558
Pot. evapotransp. ¹⁾ (mm/y)	552	555	564	543	585
W x L (m)	70 x 166	135 x 184	91 x 185	105 x 120	150 x 160
Area (ha)	1.1	2.4	1.7	1.3	2.3
Tile drain Depths to tile drain (m b.g.s.) Monitoring initiated	No May 1999	No Sep 1999	Yes 1.1 Apr 2000	Yes 1.1 Apr 2000	Yes 1.2 Sep 1999
Geological characteristics					
– Deposited by	Saltwater	Meltwater	Glacier	Glacier/meltwater	Glacier
– Sediment type – DGU symbol	Fine sand YS	Coarse sand TS	Clayey till ML	Clayey till ML	Clayey till ML
Depth to the calcareous matrix (m b.g.s.)Depth to the reduced matrix (m b.g.s.)	6 >12	5–9 10–12	1.3 5	$1-4^{(2)}$ >5 $^{(2)}$	1.5 4.2
$-$ Max. fracture depth $^{3)}$ (m)	_	_	4	>6.5	8
- Fracture intensity 3-4 m depth (fractures/m)	-	- 1 2 10 ⁻⁴	<1	11 8 0 10 ⁻⁸	4
- KS III C HOHZON (III/S)	2.0.10	1.5.10	5.4.10	8.0.10	7.2.10
	IDO	ID 1	107	ID 5 / 6	
- Classification	JB2 Loamy sand	Sand	JB / Sandy clay loam/	Sandy loam	JB5/6 Sandy loam
– Clay content (%)	6	5	18–26	10-20	14–15
– Silt content (%)	13	4	27	20-27	25
– Sand content (%)	78	88	8	50-65	57
– pH	4-4.5	5.6-6.2	6.7–7	6.5-7.8	6.4–6.6
– TOC (%)	2.0	1.8	2.2	1.7–7.3	1.4

Table 1.	Characteristics	of the five Pl	LAP sites	(modified from	Lindhardt et al	2001).
	Circle corrigered	01 010 11 0 1 1	DI II DICCO	,	Difference of criti	

^{T)} Yearly normal based on a time series for the period 1961–90. The data refer to precipitation measured 1.5 m above ground. ²⁾ Large variation within the field.

³⁾ Maximum fracture depth refers to the maximum fracture depth found in excavations and wells.

Site characterization and monitoring design are described in detail in Lindhardt *et al.* (2001). The present report presents the results of the monitoring period May 1999–June 2009. The main focus of this report is on the leaching risk of pesticides applied during 2007. For a detailed description of the earlier part of the monitoring period (May 1999–June 2009), see previous publications on http://pesticidvarsling.dk/publ_result/index.html.

Under the PLAP the leaching risk of pesticides is evaluated on the basis of at least two years of monitoring data. For some pesticides the present report must be considered preliminary because they have been monitored for an insufficient length of time.

Hydrological modelling of the unsaturated zone at each PLAP site supports the monitoring data. The MACRO model (version 5.1), see Larsbo *et al.* (2005), was used to describe the soil water dynamics at each site during the entire monitoring period from May 1999–June 2010. The five site models have been calibrated for the monitoring period May 1999–June 2004 and validated for the monitoring period July 2004–June 2010.

Scientifically valid methods of analysis are essential to ensure the integrity of the PLAP. The field monitoring work has therefore been supported by intensive quality assurance entailing continuous evaluation of the analyses employed. The quality assurance methodology and results are presented in Section 7.

2 Pesticide leaching at Tylstrup

2.1 Materials and methods

2.1.1 Site description and monitoring design

Tylstrup is located in northern Jutland (Figure 1). The test field covers a cultivated area of 1.1 ha (70 x 166 m) and is practically flat, with windbreaks bordering the eastern and western sides. Based on two soil profiles dug in the buffer zone around the test field the soil was classified as a Humic Psammentic Dystrudept (Soil Survey Staff, 1999). The topsoil is characterised as loamy sand with 6% clay and 2.0% total organic carbon (Table 1). The aquifer material consists of an approx. 20 m deep layer of marine sand sediment deposited in the Yoldia Sea. The southern part is rather homogeneous, consisting entirely of fine-grained sand, whereas the northern part is more heterogeneous due to the intrusion of several silt and clay lenses (Lindhardt *et al.*, 2001). The overall direction of groundwater flow is towards the west (Figure 2). During the monitoring period the groundwater table was 2.6–4.5 m b.g.s. (Figure 3). A brief description of the sampling procedure is provided in Appendix 2. The monitoring design and test site are described in detail in Lindhardt *et al.* (2001), and the analysis methods in Kjær *et al.* (2002).



Figure 2.Overview of the **Tylstrup** site. The innermost white area indicates the cultivated land, while the grey area indicates the surrounding buffer zone. The positions of the various installations are indicated, as is the direction of groundwater flow (by an arrow). Pesticide monitoring is conducted monthly and half-yearly from suction cups and selected monitoring wells as described in Table A2.1 in Appendix 2.

2.1.2 Agricultural management

Management practice during the 2009 and 2010 growing seasons is briefly summarized below and detailed in Appendix 3 (Table A3.1). For information about management practice during the previous monitoring periods, see previous monitoring reports available on http://pesticidvarsling.dk/publ_result/index.html.

The field was ploughed on 10 April 2009 and on 14 April sown with spring barley (cv. Keops), which emerged on 21 April. On 15 May, when the barley had three detectable tillers, the herbicides MCPA and bentazone were applied, of which only the latter was monitored. Fungi were treated on 23 June at 80% inflorescence using azoxystrobin, and pests were treated on 8 July at late milk stage using tau-fluvalinate. Tau-fluvalinate and azoxystrobin were not included in the monitoring programme. The barley received 26 mm irrigation on 29 June at the end of flowering and 27 mm at late milk stage on 8 July. An amount of 53.4 hkg/ha of grain (85% dry matter (DM)) was harvested on 20 August, slightly above the average for the soil type this year (Plantedirektoratet, 2009). On 28 august, 17.4 hkg/ha of straw (100% DM) was removed from the field.

On 4 April 2010 the field was ploughed and on 6 May planted with potatoes (cv. Kuras). On 26 May, before emergence, the field was sprayed with the herbicides aclonifen and rimsulfuron. Rimsulfuron was applied again on 8 June, when the fifth leaf of the main stem had unfolded. Cyazofamid was used against fungi six times between 15 June and 2 August. On 9 July a fungicide containing mancozeb and metalaxyl was used, of which only metalaxyl was monitored. Between 9 August and 23 August the fungicide mancozeb was again applied, but not monitored. The crop was irrigated twice, with 29 mm on 6 July and 28 mm on 27 July. The yield of potatoes, harvested on 20 October, was 470.3 hkg/ha with 27.2% DM (128.0 hkg/ha at 100% DM), yields being below the average for the year and soil type (Plantedirektoratet 2010).

2.1.3 Model setup and calibration

The numerical model MACRO (version 5.1) was applied to the Tylstrup site covering the soil profile to a depth of 5 m b.g.s., always including the groundwater table. The model was used to simulate water and bromide transport in the unsaturated zone during the full monitoring period May 1999–June 2010 and to establish an annual water balance.

Compared to Rosenbom *et al.* (2010b), a year of validation was added to the MACROsetup for the Tylstrup site. The setup was hereby calibrated for the monitoring period May 1999-June 2004 and validated for the monitoring period July 2004-June 2010. Daily time series of groundwater table measured in the piezometers located in the buffer zone, soil water content measured at three different depths (25, 60, and 110 cm b.g.s.) from the two profiles S1 and S2 (Figure 2) and the bromide concentration measured in the suction cups located 1 and 2 m b.g.s. were used in the calibration and validation process. Data acquisition, model setup and results related to simulated bromide transport are described in Barlebo *et al.* (2007).

r					
	Normal	Dessinitation	Imigation	Actual	Groundwater
	precipitation	Precipitation	Infigation	evapotranspiration	recharge
1.5.99–30.6.99 ¹⁾	120	269	0	112	156
1.7.99-30.6.00	773	1073	33	498	608
1.7.00-30.6.01	773	914	75	487	502
1.7.01-30.6.02	773	906	80	570	416
1.7.02-30.6.03	773	918	23	502	439
1.7.03-30.6.04	773	758	0	472	287
1.7.04-30.6.05	773	854	57	477	434
1.7.05-30.6.06	773	725	67	488	304
1.7.06-30.6.07	773	1147	59	591	615
1.7.07-30.6.08	773	913	126	572	467
1.7.08-30.6.09	773	1269	26	600	695
1.7.09-30.6.10	773	863	27	419	471

Table 2. Annual water balance for **Tylstrup** (mm/y). Precipitation is corrected to soil surface according to the method of Allerup and Madsen (1979).

¹⁾Accumulated for a two-month period.

²⁾ Normal values based on time series for 1961–1990.

³⁾ Groundwater recharge is calculated as precipitation + irrigation - actual evapotranspiration.

2.2 Results and discussion

2.2.1 Soil water dynamics and water balances

The model simulations were generally consistent with the observed data, thus indicating a good model description of the overall soil water dynamics in the unsaturated zone (Figure 3). The overall trends in soil water saturation were modelled successfully, with the model capturing soil water dynamics at all depths (Figure 3C-E). During the last three hydraulic years the level in water saturation at 25 cm b.g.s. was, however, overestimated. Moreover the initial decrease in water saturation observed during the summer periods at 25, 60, and 110 cm b.g.s. was less well captured. The dynamics of groundwater table were captured with some exceptions, but as with previous simulations the amplitude of the fluctuations was less well described (Figure 3B).

The resulting annual water balance is shown for each hydraulic year of the monitoring period (July–June) in Table 2. Values for precipitation and actual evapotranspiration for the most recent hydraulic year, July 2009–June 2010, were among the lowest observed since monitoring began at the site, and the monthly precipitation pattern for this year was low to medium compared with earlier years, except for the wettest November monitored. Especially January was very dry (Appendix 4). Artificial irrigation was minimal, which could be the result of a wet July. The groundwater recharge/percolation was medium compared to the other hydraulic years, and continuous (Figure 3A).



Figure 3. Soil water dynamics at **Tylstrup**: Measured precipitation, irrigation and simulated percolation 1 m b.g.s. (A), simulated and measured groundwater level GWT (B), and simulated and measured soil water saturation (SW sat.) at three different soil depths (C, D and E). The measured data in B derive from piezometers located in the buffer zone. The measured data in C, D and E derive from TDR probes installed at S1 and S2 (Figure 2). The broken vertical line indicates the beginning of the validation period (July 2004-June 2010).



Figure 4. Measured bromide concentration in the unsaturated zone at Tylstrup. The measured data derive from suction cups installed 1 m b.g.s. and 2 m b.g.s. at locations S1 (A) and S2 (B) indicated in Figure 2. The green vertical lines indicate the dates of bromide applications.

2.2.2 Bromide leaching

Bromide has now been applied twice at Tylstrup. The bromide concentrations measured until April 2003 (Figure 4 and Figure 5) relate to the bromide applied in May 1999, as described further in Kjær *et al.* (2003). Unsaturated transport of the bromide applied in March 2003 is evaluated in Barlebo *et al.* (2007).



Figure 5. Bromide concentration in the groundwater at **Tylstrup**. The data derive from monitoring wells M1–M6. Monitoring at well M6 was suspended September 2008 (Appendix 2). Screen depth is indicated in m b.g.s. The green vertical lines indicate the dates of bromide applications.

2.2.3 Pesticide leaching

Monitoring at Tylstrup began in May 1999 and presently encompasses several pesticides and their degradation products, as shown in Table 3. Pesticide applications during the latest two growing seasons are shown together with precipitation and simulated precipitation in Figure 6.

It should be noted that precipitation in Table 3 is corrected to soil surface according to Allerup and Madsen (1979), whereas percolation (1 m b.g.s.) refers to accumulated percolation as simulated with the MACRO model. It should also be noted that some of the applied pesticides degrade rapidly, e.g. mancozeb (applied here as Dithane DG), tribenuron-methyl (applied here as Express ST) and rimsulfuron (applied here as Titus). The risk of leaching is therefore associated with their respective degradation products: ETU, triazinamin-methyl, PPU and PPU-desamino. This is why the degradation products and not the parent compounds are monitored in PLAP (Table 3). Pesticides applied later than April 2010 are not evaluated in this report and hence are not included in Table 3 and Figure 6.

The current report focuses on the pesticide applied from 2008 and onwards, while leaching risk of pesticides applied before 2008 has been evaluated in previous monitoring reports (see http://pesticidvarsling.dk/publ_result/index.html). The leaching of metribuzin is further detailed in Kjær *et al.* (2005b) and Rosenborn *et al.* (2009).



Figure 6. Application of pesticides included in the monitoring programme, precipitation and irrigation (primary axis) together with simulated percolation 1 m b.g.s. (secondary axis) at **Tylstrup** in 2008/2009 upper) and 2009/2010 (lower).

Table 3. Pesticides analysed at **Tylstrup** with the products used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from the date of first application until the end of monitoring. 1^{st} month perc. refers to accumulated percolation within the first month after the application. C_{mean} refers to average leachate concentration at 1 m b.g.s. the first year after application (See Appendix 2 for calculation method).

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	Date	monitoring	(mm)	(mm)	perc. (mm)	(µg/L)
Potatoes 1999						
Linuron (Afalon)	May 99	Jul 01	2550	1253	87	< 0.01
- ETU^{I} (Dithane DG)	Jun 99	Oct 01	2381	1169	73	< 0.01
Metribuzine (Sencor WG)	Jun 99	Jul 03	4223	2097	85	< 0.01
- metribuzine-diketo		Jul 10^{\dagger}	11142	5387	85	0.05-0.36
- metribuzine-desamino		Jul 03	4223	2097	85	< 0.02
- metribuzine-desamino-diketo		Apr 08	8689	4192	85	0.14-0.97
Spring barley 2000						
Triasulfuron (Logran 20 WG)	May 00	Apr 03	2740	1283	13	< 0.02
- triazinamin						< 0.02
Propiconazole (Tilt Top)	Jun 00	Jul 03	2948	1341	11	< 0.01
Fenpropimorph (Tilt Top)	Jun 00	Jul 03	2948	1341	11	< 0.01
- fenpropimorphic acid						< 0.02
Pirimicarb (Pirimor G)	Jun 00	Apr 03	2622	1263	17	< 0.01
- pirimicarb-desmethyl						< 0.02
- pirimicarb-desmethyl-formamido						< 0.02
Winter rye 2001						
Pendimethalin (Stomp SC)	Nov 00	Apr 03	2271	1219	109	< 0.01
<i>Triazinamin-methyl</i> ²⁾ (Express)	Nov 00	Apr 03	2271	1219	109	< 0.02
Propiconazole (Tilt Top)	May 01	Jul 03	2948	1341	11	< 0.01
Fenpropimorph (Tilt Top)	May 01	Jul 03	2948	1341	11	< 0.01
- fenpropimorphic acid						< 0.01
Winter rape 2002						
Clomazone (Command CS)	Sep 01	Jul 04	2534	1194	9	< 0.01
- FMC65317 (propanamide-clomazone)	-					< 0.02

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1. $\frac{1}{2}$

¹⁾ Degradation product of mancozeb. The parent compound degrades too rapidly to be detected by monitoring.

²⁾ Degradation product of tribenuron-methyl. The parent compound degrades too rapidly to be detected by monitoring.

† Monitoring will continue during the following year. The values for prec. and perc. are accumulated up to July 2009.

Degradation products of rimsulfuron, which was applied in June 2004, continued to leach also in 2009/2010, but now only in small concentrations. The results of this 2004 application are summarised below and in Rosenborn *et al.* (2010a).

Rimsulfuron degrades rapidly in the soil, and the leaching risk is therefore associated with the degradation products PPU and PPU-desamino. PPU has been found several times in suction cups situated 1 m and 2 m b.g.s. at both S1 and S2 (Figure 7). The first detection of PPU occurred 10 months after the application of rimsulfuron (Figure 7B), after which PPU was found in 122 out of 192 analysed samples with concentrations ranging between 0.017 and 0.150 µg/l. PPU-desamino has been found in 35 out of 192 analysed samples with concentrations ranging between 0.01 and 0.042 µg/l. A little more than two years after application, PPU was found at 1 m depth at S2, whereafter PPU was detected in 46 out of 99 analysed samples with concentrations ranging between 0.01 and 0.067 µg/l (Figure 7D). At S2, the number of detections and concentration levels of PPU-desamino were low (Figure 7D and 7E; Appendix 5). Small concentrations of PPU were seen in both S1 and S2 at the end of the monitoring period, indicating that although leaching had reduced, it had not yet ceased. After application of rimsulfuron, average concentrations did not exceed 0.1 µg/l in any of the five years for either of the degradation products (Table 4).

Table 3 continued. Pesticides analysed at Tylstrup with the products used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from date of first application until the end of monitoring. 1st month perc. refers to accumulated percolation within the first month after the application. C_{mean} refers to average leachate concentration at 1 m b.g.s. the first year after application (See Appendix 2 for calculation method).

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	date	monitoring	(mm)	(mm)	perc. (mm)	(µg/l)
Winter wheat 2003						
Bromoxynil (Oxitril CM)	Oct 02	Apr 05	2082	995	53	< 0.01
Ioxynil (Oxitril CM)	Oct 02	Apr 05	2082	995	53	< 0.01
Fluroxypyr (Starane 180)	May 03	Jul 05	1867	787	50	< 0.02
Flamprop-M-isopropyl (Barnon Plus 3)	May 03	Jul 05	2635	1031	42	< 0.01
- Flamprop-M (free acid)	•					
Dimethoate (Perfekthion 500 S)	Jul 03	Jul 05	1629	722	14	< 0.01
Potatoes 2004						
-Fluazifop-P (free acid) ^{1} (Fusilade X-tra)	May 04	Jul 06	1754	704	16	< 0.01
$-PPU^{2}$ (Titus)	Jun 04	Jul 10 [†]	6211	3008	13	< 0.01 3)
- PPU-desamino ²⁾ (Titus)	Jun 04	Jul 10 [†]	6211	3008	13	< 0.013)
Maize 2005						
Terbuthylazine (Inter-Terbutylazine)	May 05	Jul 07	2145	933	16	< 0.01
-desethyl-terbuthylazine	May 05	Juior	2110	100	10	< 0.01
-2-hydroxy-terbuthylazine						< 0.01
-desisopropyl-atrazine						< 0.01 4)
-2-hvdroxy-desethvl-terbuthvlazine						< 0.01
Bentazone (Laddok TE)	Jun 05	Jul 07	2061	927	33	< 0.01
-AIBA						< 0.01
Spring barley 2006						
-triazinamin-methyl ⁵ (Express ST)	Jun 06	Jul 08	2349	1184	43	< 0.02
Epoxiconazole (Opus)	Jul 06	Jul 08	2233	1148	24	< 0.01
Winter rane 2007						
Thiamethoxam (Cruiser RAPS) ⁶⁾	Aug 06	Apr 08	2030	1123	57	< 0.01
-CGA 322704	riug oo	ripi öö	2000	1125	57	<0.02
Propyzamide (Kerb 500 SC)	Feb 07	Apr 09	2400	1172	40	< 0.01
-RH-24644						< 0.01
-RH-24580						< 0.01
-RH-24655						< 0.01
Clopyralid (Matrigon)	Mar 07	Apr 09	2317	1112	24	< 0.01
Winter wheat 2008		-				
Pendimethalin (Stomp)	Oct 07	Dec 09	2595	1324	27	< 0.01
Tebuconazole (Folicur EC250)	Nov 07	Mar 10	2696	1427	46	< 0.01
Azoxystrobin (Amistar)	Jun 08	Jul 10 [†]	2265	1151	0	< 0.01
-CyPM						< 0.01
Spring harley 2009						
Bentazone (Basagran M75)	May 09	Inf 10^{\dagger}	1084	512	22	< 0.01
Azoxystrobin (Amistar)	Iun 09	$\operatorname{Iul} 10^{\dagger}$	920	485	11	<0.01
-CvPM	- un ()	241 10	/20	100		< 0.01

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1. ¹⁾ Degradation product of fluazifop-P-butyl. The parent compound degrades too rapidly to be detected by monitoring.

²⁾ Degradation product of rimsulfuron. The parent compound degrades too rapidly to be detected by monitoring.

³⁾ Leaching increased the second and third year after application (see Figure 7 and Table 4).

⁴⁾ Leaching increased during the second year after application but measured concentrations did not exceed 0.042 μ g/l (see *Kjær et al.*, 2008).

⁵⁾Degradation product of tribenuron-methyl. The parent compound degrades too rapidly to be detected by monitoring.

⁶⁾ Unlike the other pesticide applied via surface spray application, thiamethoxam was directly applied in the soil as the rape seeds (cv. Lioness) were dressed with thiamethoxam.

† Monitoring will continue during the following year. The values for prec. and perc. are accumulated up to July 2009.

PPU was found in two groundwater samples (0.045 µg/l on 7 December 2005 in the monitoring screen of M4 located 4.4-5.4 m b.g.s. and 0.011 µg/l on 11 February 2009 in M5.5). PPU-desamino has not been detected in the groundwater (Table A5.1 in Appendix 5).



Figure 7. Precipitation, irrigation and simulated percolation 1 m b.g.s. (A) together with measured concentration of *PPU* and *PPU*-desamino (μ g/l) in suction cups installed at location S1 at 1 m b.g.s. (B) and 2 m b.g.s. (C) and location S2 at 1 m b.g.s. (D) and 2 m b.g.s. (E) at **Tylstrup**. The green vertical line indicates the date of pesticide application. Open symbols indicate concentrations below the limit of detection (0.02 μ g/l prior to July 2006 and 0.01 μ g/l thereafter). While PPU-desamino has not been detected in the groundwater, PPU was detected in two groundwater samples (see text).

	Percolation	PPU		PPU-de	samino
	(mm/y)	Suction cup – S1	Suction cup - S2	Suction cup – S1	Suction cup - S2
1.7.04-30.6.05	528	< 0.02	< 0.02	< 0.02	< 0.02
1.7.05-30.6.06	257	0.01-0.03	< 0.02	< 0.02	< 0.02
1.7.06-30.6.07	529	0.07	0.01-0.02	0.02	< 0.01
1.7.07-30.6.08	529	0.04	0.03	0.01	< 0.01
1.7.08-30.6.09	672	0.02	0.02	< 0.01	< 0.01
1.7.09-30.6.10	476	0.01	0.01	< 0.01	< 0.01

Table 4. Percolation together with estimated average concentration $(\mu g/l)$ of PPU and PPU-desamino 1 m b.g.s. at Tylstrup.

When evaluating these results it should be noted that precipitation following the application of rimsulfuron (applied on 3 June 2004) amounted to 68 mm in May 2004 (20% higher than normal) and 51 mm in June 2004 (21% lower than normal). Precipitation and percolation following the application at Tylstrup were thus much lower than at Jyndevad in 2003 where rimsulfuron was also applied. Finally, it should be noted that the concentration of PPU is likely to be underestimated by 28-47%. Results from the field-spiked samples revealed that PPU is degraded slightly during analysis (see Rosenbom *et al.*, 2010b; section 7.2.2.). Thus, the observed PPU-desamino probably derives from degradation in the sample during analysis rather than from degradation occurring in the soil. As a consequence, the concentration of PPU is likely to be underestimated.

The pesticides applied on winter wheat in 2008 and spring barley in 2009 and their degradation products (Table 3) have not been found in any of the analysed water samples, the exception being tebuconazole detected once in a groundwater sample (0.011 μ g/l on 4 April 2008 in M4, 2.5–3.5 m b.g.s.). Moreover, bentazone has been found in one sample (10 April 2006, at S1 1 m b.g.s.) originating from the application in 2005, indicating that no leaching occurred from the 2009 application.

3 Pesticide leaching at Jyndevad

3.1 Materials and methods

3.1.1 Site description and monitoring design

Jyndevad is located in southern Jutland (Figure 1). The test site covers a cultivated area of 2.4 ha (135 x 184 m) and is practically flat. A windbreak borders the eastern side of the test site. The area has a shallow groundwater table ranging from 1 to 3 m b.g.s. (Figure 9B) The overall direction of groundwater flow is towards the northwest (Figure 8). The soil can be classified as Arenic Eutrudept and Humic Psammentic Dystrudept (Soil Survey Staff, 1999) with coarse sand as the dominant texture class and topsoil containing 5% clay and 1.8% total organic carbon (Table 1). The geological description points to a rather homogeneous aquifer of meltwater sand, with local occurrences of thin clay and silt beds. A brief description of the sampling procedure is provided in Appendix 2. The monitoring design and test site are described in detail in Lindhardt *et al.* (2001) and the analysis methods in Kjær *et al.* (2002).

3.1.2 Agricultural management

Management practice during the 2008-2009 growing seasons is briefly summarized below and detailed in Appendix 3 (Table A3.2). For information about management practice during the previous monitoring periods, see previous monitoring reports available on http://pesticidvarsling.dk/publ_result/index.html.



Figure 8. Overview of the **Jyndevad** site. The innermost white area indicates the cultivated land, while the grey area indicates the surrounding buffer zone. The positions of the various installations are indicated, as is the direction of groundwater flow (by an arrow). Pesticide monitoring is conducted monthly and half-yearly from selected monitoring wells and suctions cups as described in Table A2.1 in Appendix 2.

The field was ploughed on 17 March 2009 and the following day sown with spring barley (cv. Simba). The herbicide bifenox was applied on 27 April before the start of tillering. The herbicides bentazone and MCPA were applied two weeks later, at the beginning of stem elongation, although MCPA was not included in the monitoring. Fungicides were applied around late boot stage on 26 May, using boscalide and epoxiconazole, of which only the latter was monitored. The field was irrigated on three occasions: 30 mm on 26 May, at the late boot stage; 27 mm on 5 June at the beginning of heading and finally 27 mm on 29 June, at the beginning of flowering. The crop was harvested on 7 August, yielding 64.0 hkg/ha of grain and 19.5 hkg of straw (85 and 100% DM, respectively), grain yield being nearly 30% above the average for the soil type and year (Plantedirektoratet, 2009).

Having been ploughed on 14 April 2010, the field was planted with potatoes (cv. Kuras) on 4 May. Before the potatoes emerged the field was sprayed with the herbicides aclonifen and rimsulfuron on 27 May. Rimsulfuron was applied again on 8 June. The fungicide cyazofamid was used five times between 28 June and 9 August , whereas azoxystrobin was applied on 6 July and a combination of mancozeb and metalaxyl on 25 July. Aphids were sprayed once using lambda-cyhalothrin on 16 July. The field was irrigated three times with 25, 25, and 30 mm on 24 June, 30 June and 8 July, respectively. Neither mancozeb, azoxystrobin nor lambda-cyhalothrin was included in the monitoring. The potatoes were harvested on 19 October yielding 450 hkg/ha with 26.8% DM (120.6 hkg/ha at 100% DM), yields being below the average for the soil type and year (Plantedirektoratet, 2010).

3.1.3 Model setup and calibration

The numerical model MACRO (version 5.1, Larsbo *et al.*, 2005) was applied to the Jyndevad site covering the soil profile to a depth of 5 m b.g.s., always including the groundwater table. The model was used to simulate water flow and bromide transport in the unsaturated zone during the full monitoring period July 1999–June 2010 and to establish an annual water balance.

Compared with the setup in Rosenbom *et al.* (2010b), a year of validation was added to the MACRO-setup for the Jyndevad site. The setup was hereby calibrated for the monitoring period May 1999-June 2004 and validated for the monitoring period July 2004-June 2010. For this purpose, the following time series have been used: the groundwater table measured in the piezometers located in the buffer zone, soil water content measured at three different depths (25, 60, and 110 cm b.g.s.) from the two profiles S1 and S2 (location indicated at Figure 8), and the bromide concentration measured in the suction cups located 1 and 2 m b.g.s (Figure 11). Data acquisition, model setup as well as results related to simulated bromide transport are described in Barlebo *et al.* (2007).



Figure 9. Soil water dynamics at **Jyndevad**: Measured precipitation, irrigation and simulated percolation 1 m b.g.s. (A), simulated and measured groundwater level (B), and simulated and measured soil water saturation (SW sat.) at three different soil depths (C, D and E). The measured data in B derive from piezometers located in the buffer zone. The measured data in C, D and E derive from TDR probes installed at S1 and S2 (Figure 8). The broken vertical line indicates the beginning of the validation period (July 2004-June 2010).

3.2 Results and discussion

3.2.1 Soil water dynamics and water balances

The model simulations were generally consistent with the observed data, thus indicating a good model description of the overall soil water dynamics in the unsaturated zone (Figure 9). The dynamics of the simulated groundwater table were well described with MACRO 5.1 (Figure 9B). For the recent hydraulic year, no measurements of the water saturation were obtained during the following two periods: 1 June – 25 August 2009 (given a breakdown in the TDR measuring system) and 7 February – 6 March 2010 (given a sensor error). However, as noted earlier in Rosenbom *et al.* (2010b), the model still had some difficulty in capturing the degree of soil water saturation 1.1 m b.g.s. (Figure 9E) and also the decrease in water saturation observed during summer periods at 25 and 60 cm b.g.s.

The resulting water balance for Jyndevad for the 11 monitoring periods is shown in Table 5. Compared with the previous ten years, the latest hydraulic year July 2009-June 2010 was characterised by having medium precipitation, simulated actual evapotranspiration and irrigation values. Precipitation in the latest hydraulic year was characterised by November being very wet, and January to June being very dry (Appendix 4). Periods without continuous percolation 1 m b.g.s. were therefore simulated in the spring of 2010.

	Normal			Actual	Groundwater		
	Precipitation ¹⁾	Precipitation	Irrigation	Evapotranspiration	Recharge ²⁾		
1.7.99-30.6.00	995	1073	29	500	602		
1.7.00-30.6.01	995	810	0	461	349		
1.7.01-30.6.02	995	1204	81	545	740		
1.7.02-30.6.03	995	991	51	415	627		
1.7.03-30.6.04	995	937	27	432	531		
1.7.04-30.6.05	995	1218	87	578	727		
1.7.05-30.6.06	995	857	117	490	484		
1.7.06-30.6.07	995	1304	114	571	847		
1.7.07-30.6.08	995	1023	196	613	605		
1.7.08-30.6.09	995	1048	114	547	615		
1.7.09-30.6.10	995	1034	80	516	599		

Table 5. Annual water balance for **Jyndevad** (mm/yr). Precipitation is corrected to the soil surface according to the method of Allerup and Madsen (1979).

¹⁾ Normal values based on time series for 1961–1990.

²⁾ Groundwater recharge is calculated as precipitation + irrigation - actual evapotranspiration.



Figure 10. Bromide concentration in the unsaturated zone at **Jyndevad**. The measured data derive from suction cups installed 1 m b.g.s. (upper)and 2 m b.g.s. (lower)at locations S1 and S2 (Figure 8). The green vertical lines indicate the dates of bromide applications.

3.2.2 Bromide leaching

Bromide has now been applied twice at Jyndevad. The bromide concentrations measured until April 2003 (Figure 10 and Figure 11) relate to the bromide applied in autumn 1999, as described further in Kjær *et al.* (2003). Leaching of the bromide applied in March 2003 is evaluated in Barlebo *et al.* (2007).


Figure 11. Bromide concentration in the groundwater at **Jyndevad**. The data derive from monitoring wells M1–M7. Monitoring at well M6 was suspended September 2008 (Appendix 2). Screen depth is indicated in m b.g.s. The green vertical lines indicate the dates of bromide applications.



Figure 12. Application of pesticides included in the monitoring programme, precipitation and irrigation (primary axis) together with simulated percolation 1 m b.g.s. (secondary axis) at Jyndevad in 2008/2009 (upper) and 2009/2010 (lower).

3.2.3 Pesticide leaching

Monitoring at Jyndevad began in September 1999 and presently encompasses several pesticides and their degradation products, as indicated in Table 6. Pesticide application during the two most recent growing seasons is shown together with precipitation and simulated precipitation in Figure 12. It should be noted that precipitation is corrected to the soil surface according to Allerup and Madsen (1979), whereas percolation (1 m b.g.s.) refers to accumulated percolation as simulated with the MACRO model. It should also be noted that as tribenuron-methyl (applied here as Express), pyridate (applied here as Lido) and rimsulfuron (applied here as Titus) degrade rapidly. The leaching risk is therefore associated with their respective degradation products: triazinamin-methyl, PHCP, PPU, and PPU-desamino rather than the parent compounds. For the same reasons the degradation products and not the parent compounds are monitored in PLAP (Table 6). The product Basagran M75 contains two active substances, bentazone and MCPA, but only bentazone is monitored. Pesticides applied later than April 2010 are not evaluated in this report and hence not included in Table 6 and Figure 12.

Table 6. Pesticides analysed at Jyndevad with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from date of first application until end of monitoring. 1st month perc. refers to accumulated percolation within the first month after application. Cmean refers to average leachate concentration 1 m b.g.s the first year after application (See Appendix 2 for calculation method).

Crop and analysed pesticides	Application	End of Monitoring	Prec.	Perc.	1 st month	C_{mean}
W/indow	Date	Womoning	(11111)	(11111)	perc. (iiiii)	(μg/1)
Glyphosate (Roundup 2000)	Son 00	$\Delta pr 02$	2750	1607	130	<0.01
- AMPA	Sep 99	Apr 02	2139	1007	139	< 0.01
Triazinamin-methyl ¹⁾ (Express)	Nov 99	Apr 02	2534	1451	86	<0.01
Propiconazole (Tilt Top)	Apr 00	Jul 02	2301	1061	3	< 0.01
Fenpropimorph (Tilt Top)	Apr 00	Apr 02	2015	1029	3	< 0.01
- fenpropimorphic acid						< 0.01
Maize 2001						
Terbuthylazine (Lido 410 SC)	May 01	Apr 04	3118	1809	4	< 0.01
- desethyl-terbuthylazine	May 01	Apr 07	6742	3826	4	< 0.01 - 0.02
PHCP ²⁾ (Lido 410 SC)	May 01	Jul 03	2413	1366	4	< 0.02
Potatoes 2002						
- PPU (Titus) ³⁾	May 02	Jul 10 [†]	9389	5126	11	$0.06^{4}-0.13$
- PPU-desamino (Titus) ³⁾		Jul 10 [†]	9389	5126	11	0.01-0.03
Spring barley 2003					_	
MCPA (Metaxon)	Jun 03	Jul 05	2340	1233	0	< 0.01
- 4-chlor,2-methylphenol	I 02	L-1.05	2279	1020	1	< 0.01
Dimethoate (Perfektition 500 S)	Jun 03	Jul 05	2278	1232	1	<0.01
Pea 2004 Deptazone (Decement 180)	Max 04	Lul 07	2000	2044	4	0.02 0.12
AIDA	May 04	Jul 07	2000	2044	4	0.02 - 0.15
- AIDA Pendimethalin (Stomp SC)	May 04	Apr 07	3557	1996	Δ	< 0.01
Pirimicarb (Pirimor G)	Jun 04	Apr 07	3493	1993	27	< 0.01
- Pirimicarb-desmethyl	bull of	ripi ov	0.70	1770	_,	< 0.01
-Pirimicarb-desmethyl-						< 0.02
formamido						
- fluazifop-P(free acid) ⁵⁾	Jun 04	Jul 06	2395	1233	27	< 0.01
(Fusilade X-tra)						
Winter wheat 2005						
Ioxynil (Oxitril CM)	Oct 04	Apr 07	2955	1791	81	< 0.01
Bromoxynil (Oxitril CM)	Oct 04	Apr 07	2955	1791	81	< 0.01
Amidosulfuron (Gratil 75 WG)	Apr 05	Jul 07	1070	515	33	< 0.01
Fluroxypyr (Starane 180 S)	May 05	Jul 07	2683	1360	37	< 0.02
Azoxystrobin (Amistar)	May 05	Apr 07	2274	1283	49	< 0.01
- CyPM						< 0.02
Spring barley 2006						
Florasulam (Primus)	May 06	Jul 08	2779	1487	34	< 0.01
- florasulam-desmethyl	T OC	D 00	1.600	2502		< 0.03
Epoxiconazole (Opus)	Jun 06	Dec 09	4698	2592	31	<0.01
Triticale 2007		D 00		2410	50	0.01
Mesosulfuron-methyl (Atlantis WG)	Oct 06	Dec 09	4177	2418	73	< 0.01
- mesosulfuron Chlormaguat (Cyapael 750)	Apr 07	Jul 08	15/9	680	1	< 0.01
Enovicence (Cycocer 750)	Apr 07	Jui 08	2465	1015	I C	< 0.01
Epoxicoliazole (Opus)	Way 07	Dec 09	5405	1015	0	<0.01
Winter wheat 2008	0 + 07	M 10	2024	1605	<i></i>	-0.01
Picolinaten (Pico 750 WG)	Oct 07	Mar 10	2934	1685	22	< 0.01
- CLISSOIS Tebuconazole (Folicur FC 250)	Dec 07	Mar 10	2807	1626	97	< 0.01
Service header 2000	Dec 07	Wai 10	2807	1020	21	<0.01
Bifency (Ecy 490 SC)	A mm 00	$\mathbf{L} 1 1 0^{\dagger}$	1276	ENT	2	<0.02
- hifenox acid	Apr 09	Jul 10	13/0	047	3	<0.02
- oyenos ucu - nitrofen						<0.05
Bentazone (Basagran M75)	May 09	Jul 10^{\dagger}	1328	646	14	0.06
Epoxiconazole (Bell)	May 09	Dec 09	972	518	43	< 0.01

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1. ¹⁾ Degradation product of tribenuron-methyl. The parent compound degrades too rapidly to be detected by monitoring. ²⁾ Degradation product of pyridate. The parent compound degrades too rapidly to be detected by monitoring.

³⁾ Degradation product of pinsulfuron. The parent compound degrades too rapidly to be detected by monitoring.

⁴⁾ Leaching increased the second year after application (see Figure 13).
⁵⁾ Degradation product of fluazifop-P-butyl. The parent compound degrades too rapidly to be detected by monitoring.

† Monitoring will continue for an additional year. The values for prec. and perc. are accumulated up to July 2009.



Figure 13. Precipitation, irrigation and simulated percolation 1 m b.g.s. (A) together with measured concentrations of *PPU* and *PPU-desamino* (μ g/l) in suction cups installed 1 m b.g.s. at location S1 (B) and S2 (C) at **Jyndevad**. The green vertical line indicates the date of pesticide application. Open symbols indicate concentrations below the limit of detection (0.02 μ g/l prior to July 2006 and 0.01 μ g/l thereafter).

The current report focuses on the pesticides applied from 2008 and onwards, while leaching risk of pesticides applied before 2008 has been evaluated in previous monitoring reports (see http://pesticidvarsling.dk/publ_result/index.html). Since PPU and PPU-desamino (degradation products of rimsulfuron applied in 2003) were still included in the current monitoring period, the results of these applications are, however, summarised below and in Rosenbom *et al.* (2010a). For a detailed description of the leaching pattern, including primary data and climatic conditions characterising the monitoring periods, see Kjær *et al.* (2005c).



Figure 14. Precipitation, irrigation and simulated percolation 1 m b.g.s. (A) together with measured concentrations $(\mu g/l)$ in downstream (M1, M2, M4) and upstream monitoring wells (M7) of *PPU* (B) and *PPU-desamino* (C) at **Jyndevad**. The numbers in parentheses indicate the depths of the analysed screens. The green vertical line indicates the date of pesticide application. Open symbols indicate concentrations below the limit of detection (0.02 $\mu g/l$ prior to July 2006 and 0.01 $\mu g/l$ thereafter).

Two degradation products of rimsulfuron, PPU and PPU-desamino, were detected at 1 m depth in the suction cups at S1 and S2 (Figure 13). Both compounds were characterised by continuous leaching over a long period of time. Although the concentration decreased during the last monitoring year, PPU could still be found in low concentrations towards the end of 2009/2010, i.e. eight years after application. Average yearly concentrations of PPU reaching 0.1 μ g/l were seen as long as three years after application (Figure 13 and Table 7). With an overall travel time of about four years, PPU also reached the downstream monitoring screens (Figure 14). The most recent detection of PPU in monitoring screens of M1 (sampled monthly) was 0.011 μ g/l on 3 February 2010. In M2 (sampled half-yearly) the most recent detections were from 30 September 2009 (0.022- 0.046 μ g/l). PPU showed up in M4 (sampled monthly) for the first time in September 2006. From then and until April 2010 PPU has been detected in 119 of a total of 129 samples, concentrations ranging between 0.011 and 0.049 μ g/l. In M7, PPU was still present on 17 March 2010 at 0.026 μ g/l (Figure 9 and Figure 15).

	Percolation	PPU		PPU-des	samino
	(mm/y)	Suction cup – S1	Suction cup – S2	Suction cup – S1	Suction cup - S2
1.7.02-30.6.03	706	0.13	0.06	0.03-0.04	0.01-0.03
1.7.03-30.6.04	468	0.12	0.10	0.04	0.04
1.7.04-30.6.05	759	0.10	0.14	0.03-0.04	0.05
1.7.05-30.6.06	465	0.07	0.09	< 0.02	0.01-0.02
1.7.06-30.6.07	815	0.05	0.08	0.01	0.02
1.7.07-30.6.08	643	0.02	0.04	< 0.01	0.01
1.7.08-30.6.09	623	< 0.01	0.02	< 0.01	< 0.01
1.7.09-30.6.10	619	< 0.01	0.01	< 0.01	< 0.01

Table 7. Percolation together with estimated average concentrations (μ g/l) of PPU and PPU-desamino 1 m b.g.s. at **Jyndevad**. Leached mass refers to the total mass (% of applied rimsulfuron) leached during the monitoring period 1.7.02–30.6.10.

¹⁾ Expressed as rimsulfuron equivalent

The tracer test suggested that water sampled in M7 had not infiltrated at the PLAP site, but originated from the upstream neighbouring fields, where rimsulfuron had also been applied (Kjær *et al.*, 2007).

Furthermore, PPU-desamino was detected in monitoring wells, although the number of detections and concentration levels were lower than those of PPU-desamino (Figure 14C and Table A5.2 in Appendix 5). Finally, it should be noted that the concentration of PPU is likely to be underestimated by up to 22-44% due to stability problems, as described in Rosenbom *et al.* 2010a and section 7.2.2.

Four pesticides (tebuconazole, chlormequat, epoxiconazole and picolinafen, which can degrade to CL153815) were applied during the 2007/08 growing season. Picolinafen was detected in a monitoring well just once, at a concentration of $0.015 \,\mu$ g/l, whereas its metabolite CL153815 was not found. Tebuconazole was detected once at $0.014 \,\mu$ g/l in suction cups. Juhler *et al.* (2010) conducted a detailed analysis on the fate and transport of chlormequat at the site. This analysis was financially supported by Copenhagen Energy (Københavns Energi A/S). Chlormequat was not detected at all and epoxiconazole just once. None of the detections exceeded 0.1 μ g/l (Table A5.2, Appendix 5).

The herbicides bifenox and bentazone were used in the spring barley sown in 2009. Bifenox was found twice in suctions cups, in concentrations of 0.034 and 0.036 μ g/l, four to six months after the application as well as in two monitoring wells five months after application, in concentrations of 0.05 and 0.033 μ g/l (Table A5.2). The metabolite bifenox acid was found once, 0.1 μ g/l, in a monitoring well four months after application. Bentazone was absent in all the samples from the monitoring wells (Table 20). It was, however, found frequently in samples from suction cups (Figure 15B and Table 18), reaching a maximum of 0.71 μ g/l. None of the substances were leached in yearly average concentrations exceeding 0.1 μ g/l (Table 6).



Figure 15. Precipitation, irrigation and simulated percolation 1 m b.g.s. (A) together with measured concentrations of bentazone in suction cups installed 1 m b.g.s. at location S and S2 (B) at **Jyndevad**. The green vertical line indicates the date of pesticide application. Open symbols indicate concentrations below the limit of detection. Bentazone has not been detected in any water samples from the groundwater monitoring wells.

4 Pesticide leaching at Silstrup

4.1 Materials and methods

4.1.1 Site description and monitoring design

The test field at Silstrup is located south of Thisted in north-western Jutland (Figure 1). The cultivated area is 1.69 ha (91 x 185 m) and slopes gently $1-2^{\circ}$ to the north (Figure 15). Based on two profiles excavated in the buffer zone bordering the field the soil was classified as Alfic Argiudoll and Typic Hapludoll (Soil Survey Staff, 1999). The topsoil content of clay in the two profiles was 18 and 26%, and the organic carbon content was 3.4 and 2.8%, respectively (Table 1). The geological description showed rather homogeneous clay till rich in chalk and chert, containing 20–35% clay, 20–40% silt, and 20–40% sand. In some intervals the till was sandier, containing only 12–14% clay. Moreover, thin lenses of silt and sand were found in some of the wells. The gravel content was approx. 5%, but could be as high as 20%. A brief description of the sampling procedure is provided in Appendix 2. The monitoring design and test site are described in detail in Lindhardt *et al.* (2001) and the analysis methods in Kjær *et al.* (2002).

4.1.2 Agricultural management

Management practice during the 2008-2009 growing seasons is briefly summarized below and detailed in Appendix 3 (Table A3.3).. For information about management practice during the previous monitoring periods, see previous reports available on http://pesticidvarsling.dk/publ_result/index.html.

Having been harrowed, levelled and rolled between 21 April and 5 May 2008 the field was sown with fodder beat (cv. Kyros) on 7 May, which emerged on 15 May. A first spraying of weeds was done on 22 May when the first leaf was visible (pinhead-size) and the cotyledons horizontally unfurled, using triflusulfuron, metamitron, and phenmedipham. A second spraying of weeds took place on 30 May, the crop having three leaves unfurled, using triflusulfuron, metamitron, ethofumesate, and phenmedipham.



Figure 16. Overview of the **Silstrup** site. The innermost white area indicates the cultivated land, while the grey area indicates the surrounding buffer zone. The positions of the various installations are indicated, as is the direction of groundwater flow (by an arrow). Pesticide monitoring is conducted weekly from the drainage system (during period of continuous drainage runoff) and monthly and half-yearly from selected groundwater monitoring wells as described in Table A2.1 in Appendix 2.

A third spraying of weeds took place on 17 June, at the stage of five unfurled leaves, using triflusulfuron, metamitron, ethofumesate, and phenmedipham. All of these herbicides except phenmedipham were included in the monitoring programme. On 26 June pests were treated with pirimicarb (not monitored) at the stage of six leaves. On 1 July weeds were sprayed a fourth time using fluazifop-P-butyl, at seven leaves. When the crop had eight unfurled leaves on 4 July a fifth and final treatment of weeds was done using triflusulfuron, metamitron, and phenmedipham. A last spraying with pirimicarb (not monitored) against pests took place on 9 July where the crop covered 20% of the area. Beets were harvested on 27 October, yielding 17.3 t/ha of 100% DM. All beet tops (5.2 t/ha) were shredded and ploughed into the soil on 15 December.

Having been harrowed twice on 30 March 2009, pig slurry was injected and potassium bromide applied as a tracer on 2 April. Sowing of spring barley (cv. Keops), undersown with red fescue (cv. Jasperina), took place on 11 April 2009. The herbicide bentazone was sprayed on 19 May, when the barley had four detectable tillers. The fungicide azoxystrobin was applied on 24 June, but not included in the monitoring. On 16 July the spring barley was harvested as wholecrop, yielding 94.6 hkg/ha (100% DM). The red fescue was later sprayed with the herbicides iodosulfuron on 24 August, four tillers detectable, and with bifenox on 9 September, five tillers detectable. On 2 May 2010 weeds were sprayed with fluazifop-p-butyl and on 5 May with iodosulfuron and MCPA, the latter two were, however, not included in the monitoring. Harvest of grass seeds took place on 20 July, yielding 16.5 hkg/ha of seeds (87% DM). An amount of 69.3 hkg/ha of straw (100% DM) was burned in the field on 21 July 2010.

4.1.3 Model setup and calibration

Compared with the setup in Rosenborn et al. (2010b), a year of validation was added to the MACRO setup for the Silstrup site. The setup was hereby calibrated for the monitoring period May 1999-June 2004 and validated for the monitoring period July 2004-June 2010. For this purpose, the following time series have been used: the observed groundwater table measured in the piezometers located in the buffer zone, soil water content measured at three depths (25, 60, and 110 cm b.g.s.) from the two profiles S1 and S2 (Figure 15), and the measured drainage flow. Data acquisition, model setup and results related to simulated bromide transport are described in Barlebo et al. (2007). Given impounding of water in the drainage water monitoring well, estimates for the measured drainage on 11 December 2006, 13-14 December 2006, and 28 February 2007 were based on expert judgment. Additionally, TDR-measurements at 25 cm b.g.s. in the period 15 December 2009 - 20 March 2010 were discarded given freezing soils (soil temperatures at or below zero degrees Celsius). The soil water content is measured with TDR based on Topp calibration (Topp et al., 1980), which will underestimate the total soil water content at the soil water freezing point as the permittivity of frozen water is much less than that of liquid water (Flerchinger et al., 2006).

4.2 Results and discussion

4.2.1 Soil water dynamics and water balances

The model simulations were largely consistent with the observed data, thus indicating a reasonable model description of the overall soil water dynamics in the unsaturated zone (Figure 17). As in Rosenbom et al. (2010b), the simulated groundwater table of this hydraulic year was validated against the much more fluctuating groundwater table measured in piezometer P3, which yielded the best description of measured drainage (Figure 17B and 17C). The first drainage flow period of the past year was well captured by the model, whereas the magnitude of the second drainage flow period was not captured (Figure 17C). The last period can be characterised by frozen soil and precipitation in the form of snow - a situation, which MACRO has difficulties in describing. Additionally, drainage flow (q) is calculated by means of continuous measurements of the water height (h) at a V-notch in the drainage well and a q/h relationship. At events with extreme water flow it may happen that the discharge pipe has inadequate capacity to discharge the water, so that water will rise above the V-notch plate in the drainage well. This was the case during the snowmelt occurring on 12 March 2010, why adequate water sampling was not possible. At these events the q/h relationship is not valid, and the drainage flow is estimated at a water height (h) corresponding to the maximum height (h) of the V-notch plate, which could have resulted in an overestimation of the measured drainage flow. As in the previous monitoring periods, the overall trends in soil water content were described reasonably well (Figure 17D, 17E, and 17F), although the model still tended to describe the subsoil as being much drier during the summer period than measured by the deeper TDR probes (Figure 17E and 17F).



Figure 17. Soil water dynamics at **Silstrup**: Measured precipitation and simulated percolation 1 m b.g.s. (A), simulated and measured groundwater level GWT (B), simulated and measured drainage flow (C), and simulated and measured soil water saturation (SW sat.) at three different soil depths (D, E, and F). The measured data in B derive from piezometers located in the buffer zone. The measured data in D, E, and F derive from TDR probes installed at S1 and S2 (Figure 16). The broken vertical line indicates the beginning of the validation period (July 2004-June 2010).

	Normal		Actual	Measured	Simulated	Groundwater
	precipitation ²⁾	Precipitation	evapotranspiration	drainage	drainage	recharge ³⁾
$1.7.99 - 30.6.00^{1}$	976	1175	457	-	443	275 ⁴⁾
1.7.00-30.6.01	976	909	413	217	232	279
1.7.01-30.6.02	976	1034	470	227	279	338
1.7.02-30.6.03	976	879	537	81	74	261
1.7.03-30.6.04	976	760	517	148	97	94
1.7.04-30.6.05	976	913	491	155	158	267
1.7.05-30.6.06	976	808	506	101	95	201
1.7.06-30.6.07	976	1150	539	361	307	249
1.7.07-30.6.08	976	877	434	200	184	242
1.7.08-30.6.09	976	985	527	161	260	296
1.7.09-30.6.10	976	835	398	203	222	234

Table 8. Annual water balance for **Silstrup** (mm/year). Precipitation is corrected to the soil surface according to the method of Allerup and Madsen (1979).

¹⁾ The monitoring started in April 2000.

²⁾ Normal values based on time series for 1961–1990 corrected to soil surface.

³⁾ Groundwater recharge calculated as precipitation - actual evapotranspiration - measured drainage.

⁴⁾ Where drainage flow measurements were lacking, simulated drainage flow was used to calculate groundwater recharge.

The resulting water balance for Silstrup for the entire monitoring period is shown in Table 8. Compared with the previous 11 years, the recent hydraulic year July 2009-June 2010 was characterised by having the third lowest precipitation, the lowest simulated actual evapotranspiration, and the fourth-highest measured drainage. Precipitation of this year was characterised by having the wettest month ever monitored at Silstrup in November with more than 300 mm, and August, September, March, April and June being very dry (Appendix 4). Due to this precipitation pattern, the simulated percolation pattern of the year July 2009-June 2010 was not represented by continuous percolation (Figure 17A). The climatic setting of this year gave rise to short periods with the groundwater table above the drainage level, causing the fourth-largest measured drainage since monitoring started in July 2000 (Figure 17B and 17C).



Figure 18. Bromide concentration at **Silstrup**. A and B refer to suction cups located at S1 and S2 (see Figure 16). The bromide concentration is also shown for drainage runoff (C) and the horizontal monitoring wells H1 and H2 (D). In March 2009, bromide measurements in the suction cups were suspended (Appendix 2). The green vertical lines indicate the dates of bromide applications.

4.2.2 Bromide leaching

The bromide concentrations shown in Figure 18 and Figure 19 relate to the bromide applied in May 2000, as described in previous reports (Kjær *et al.* 2003 and Kjær *et al.* 2004) and further evaluated in Barlebo *et al.* (2007). In Marts 2009, bromide measurements in the suction cups and monitoring wells M6 and M11 were suspended. In April 2009, 31.5 kg/ha potassium bromide was applied for the second time.



Figure 19. Bromide concentration at **Silstrup**. The data derive from the vertical monitoring wells (M5–M12). In September 2008, monitoring wells M6 and M11 were suspended (Appendix 2). Screen depth is indicated in m b.g.s. The green vertical lines indicate the date of bromide applications.

4.2.3 Pesticide leaching

Monitoring at Silstrup began in May 2000 and presently encompasses several pesticides and their degradation products, Table 9. Pesticide application during the two most recent growing seasons is shown together with precipitation and simulated percolation in Figure 19. It should be noted that precipitation in Table 9 is corrected to soil surface according to Allerup and Madsen (1979), whereas percolation (1 m b.g.s.) refers to accumulated percolation as simulated with the MACRO model. Moreover, pesticides applied later than April 2010 are not evaluated in this report and hence not included in Table 9.



Figure 20. Application of pesticides included in the monitoring programme, precipitation, and irrigation (primary axis) and simulated percolation 1 m b.g.s. (secondary axis) at Silstrup in 2008/2009 (upper) and 2009/2010 (lower).

It should also be noted that as tribenuronmethyl (applied here as Express), pyridate (applied here as Lido), and fluazifop-P-butyl (Fusilade Max) degrade rapidly, the leaching risk is associated with their respective degradation products: triazinaminmethyl, PHCP, fluazifop-P, and TFMP rather than the parent products. For the same reasons the degradation products and not the parent compounds are monitored in the PLAP (Table 9).

Table 9. Pesticides analysed at **Silstrup** with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from date of first application until end of monitoring. 1st month perc. refers to accumulated percolation within the first month after application. C_{mean} refers to average leachate concentration in the drainage water within the first drainage season after application (See Appendix 2 for calculation methods).

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	date	monitoring	(mm)	(mm)	perc. (mm)	$(\mu g/l)$
Fodder beet 2000					-	
Metamitron (Goltix WG)	May 00	Apr 03	2634	1328	53	0.05
- metamitron-desamino	illuy oo	ripi 05	2051	1520	55	0.05
Ethofumesate (Betanal Optima)	May 00	Apr 03	2634	1328	53	0.00
Desmedinham (Betanal Ontima)	May 00	Apr 03	2634	1328	53	< 0.02
- EHPC	illuy oo	ripi 05	2051	1520	55	< 0.02
Phenmedipham (Betanal Optima)	May 00	Apr 03	2634	1328	53	< 0.01
- <i>MHPC</i>						< 0.02
- 3-aminophenol						< 0.02
Fluazifop-P-butyl (Fusilade X-tra)	Jun 00	Jul 02	1953	1019	5	< 0.01
- fluazifop (free acid)						< 0.02
Pirimicarb (Pirimor G)	Jul 00	Jul 07	6452	2825	1	< 0.01
- pirimicarb-desmethyl						< 0.01
- pirimicarb-desmethyl-formamido						< 0.02
Spring harlow 2001						
Triazinamin methyl ¹⁾ (Express)	May 01	Jul 03	10/1	051	10	<0.02
Flampron-M-isopropyl (Barnon Plus 3)	Jun 01	Jul 03	1978	944	3	<0.02
- flamprop (free acid)	Juli 01	Ju 1 05	1720	744	5	<0.01
Propiconazole (Tilt Top)	Jun 01	Jul 03	1028	944	3	<0.01
Fennronimornh (Tilt Ton)	Jun 01	Jul 03	1928	944	3	<0.01
- fenpropinorphic acid	Juli 01	Ju 1 05	1720	744	5	<0.01
Dimethoate (Perfekthion 500 S)	Jul 01	Jul 03	1882	937	3	0.02
	541 01	Jui 05	1002	201	5	0.02
Maize 2002	0 + 01	• • • • •	2002	1.004	4.4	0.12
Glyphosate (Roundup Bio)	Oct 01	Apr 06	3802	1694	44	0.13
-AMPA	M 02	1.1.04	1764	720	(0.06
PHCP' (Lido 410 SC) Taskuthalasina (Lida 410 SC)	May 02	Jul 04	1/04	1207	0	0.06
dependent for her her her her her her her her her he	May 02	Apr 06	3320	1327	0	0.07
- desethyl-terbuthyldzine		Apr 05				0.15
- 2- nyaroxy-terbuinyiazine		Apr 05				3)
- 2-nyaroxy-aeseinyi-terbuinyiazine		Apr 05				3)
- desisopropyi-dirazine		Apr 05				
Rentazone (Basagran 180)	May 03	Jul 06	2634	1055	44	0.26
AIRA	Way 03	Jui 00	2034	1055	44	<0.20
Pendimethalin (Storm SC)	May 03	Apr 06	2634	1055	44	<0.01
Glyphosate (Roundup Bio)	Sep 03	Apr 06	2034	071		<0.01
- AMBA	Sep 05	Apr 00	2207	7/1	0	0.02
Winter wheat 2004						0.02
Prosulfocarb (Boxer FC)	Oct 03	Apr 06	2125	974	37	0.01
MCPA (Metaxon)	May 04	Jul 06	1797	710	4	< 0.01
- 4-chlor 2-methylphenol	May 04	Ju 1 00	1777	/10	-	< 0.01
Azoxystrobin (Amistar)	Jun 04	Jul 06	1781	706	0	0.01
- CvPM	buil of	Jul 07	2931	1202	õ	0.09
Pirimicarb (Pirimor G)	Jul 04	Jul 07	2818	1205	Ő	< 0.01
- Pirimicarb-desmethvl					~	< 0.01
- Pirimicarb-desmethyl-formamido						< 0.02

¹⁾ Degradation product of tribenuron-methyl. The parent compound degrades too rapidly to be detected by monitoring.

²⁾ Degradation product of pyridate. The parent compound degrades too rapidly to be detected by monitoring.

³⁾ Average leachate concentration within the first drainage season after application could not be calculated, as monitoring started January 2003 (7 mount after application). See Kjær et al.(2007) for further information.

The current report focuses on the pesticides applied from 2008 and onwards, while the leaching risk of pesticides applied before 2008 has been evaluated in previous monitoring reports (see http://pesticidvarsling.dk/publ_result/index.html).

Table 9 continued. Pesticides analysed at Silstrup with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from date of first application until end of monitoring. 1^{st} month perc. refers to accumulated percolation within the first month after application. C_{mean} refers to average leachate concentration in the drainage water within the first drainage season after application (See Appendix 2 for calculation methods).

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	date	monitoring	(mm)	(mm)	Perc. (mm)	(µg/l)
Spring barley 2005						
Fluroxypyr (Starane 180 S)	May 05	Jul 07	2012	830	11	< 0.02
Azoxystrobin (Amistar)	Jun 05	Jul 06	862	332	10	0.01
- CyPM	Jun 05	Jul 07	2012	828	10	0.02
Pirimicarb (Pirimor G)	Jul 05	Jul 07	1933	818	0	< 0.01
- Pirimicarb-desmethyl						< 0.01
- Pirimicarb-desmethyl-formamido						< 0.01
Winter rape 2006						
Propyzamide (Kerb 500 SC)	Nov 05	Apr 08	2345	1115	75	$0.22^{(I)}$
- RH-24644						0.01
- RH-24580						< 0.01
- RH-24655						< 0.01
Clopyralid (Matrigon)	Apr 06	Apr 08	2009	859	8	< 0.01
Winter wheat 2007						
Pendimethalin (Stomp Pentagon)	Sep 06	Apr 08	1686	865	0	< 0.04
Iodosulfuron-methyl-sodium (Hussar OD)	Apr 07	Apr 09	1940	875	3	< 0.01
- Metsulfuron-methyl						< 0.01
- Triazinamin						< 0.01
Chlormequat (Cycocel 750)	Apr 07	Jul 08	1099	392	3	< 0.01
Epoxiconazole (Opus)	Jun 07	Apr 09	1867	873	0	< 0.01
Fodder beet 2008		1				
Triflusulfuron (Safari)	May 08	Jul 10	1894	895	4	< 0.01
- IN-D8526	11249 000	bul 10	1071	070		< 0.01
- IN-E7710						< 0.01
- IN-M7222						< 0.01
Metamitron (Goliath)	May 08	Jul 10^{\dagger}	1894	895	4	0.01
- Metamitron-desamino						0.01
Ethofumesate	May 08	Inf 10^{\dagger}	1893	893	3	< 0.01
- <i>Flugzifon</i> - P^{2} (Fusilade Max)	Jul 08	$Jul 10^{\dagger}$	1820	890	21	<0.01
$-TEMP^{2}$ (Fusilade Max)	Ju 1 00	Jul 10	1020	070	21	0.22
Contract have 2000						0.22
Spring barley 2009	M 00	L-1 10 [†]	020	207	1	-0.01
A = a substant line (A substant)	May 09	Jul 10 [†]	929	397	1	< 0.01
AZOXYSTFODIN (AMISTAF)	Jun 09	Jul 10	833	390	0	0.01
- CyPM Iodosulfuron (Husser OD)	Aug 00	1.110^{\dagger}	726	401	0	0.06
Matsulfunon mathyl	Aug 09	Jul 10	/30	401	0	< 0.01
- Melsuljuron-melnyi Triazinamin						< 0.01
$= 1 \operatorname{maximum}_{\mathrm{Hor}} (\mathrm{For} 480 \ \mathrm{SC})$	Sep 00	$Iu1 10^{\dagger}$	710	402	0	<0.01
- Rifenor acid	Sch 02	Jui 10	/10	402	0	2 22
- Dijenoz uciu - Nitrofen						<0.01
						\0.01

¹⁾ Drainage runoff commenced two weeks prior to the application of propyzamide, and the weighted concentrations refer to the period from the date of application until 1 July 2007. ²⁾ Degradation product of fluazifop-P-butyl. The parent compound degrades too rapidly to be detected by monitoring.

† Monitoring will continue for an additional year. The values for prec. and perc. are accumulated up to July 2009.



Figure 21. Precipitation and simulated percolation 1 m b.g.s. (A) together with the concentration of metamitron (B), metamitron-desamino (C) Ethofumesate (D) and IN-E7710 (E) in the drainage runoff at **Silstrup**. The green vertical lines indicate the dates of applications. Open symbols indicate values below the detection limit of 0.01 μ g/l. While ethofumesate and IN-E7710 were not detected in any samples from groundwater monitoring, metamitron and metamitron-desamino have been found occasionally (see text).

Hitherto, there has been no leaching of triflusulfuron, whereas one of its degradation products IN-E7710 (Table 9 and Figure 21E) was found on four occasions in drainage water. The two other degradation products included in the monitoring programme (IN-D8526 and IN-M7222) could not be detected.

At Silstrup, the herbicides ethofumesate and metamitron have now been applied both in 2000 and 2008 (Table 9). Whereas 345 g/ha of ethofumesate was applied in 2000, only 70 g/ha of ethofumesate was applied in 2008 (71 g/ha every 3^{rd} year being the admissible dose). The reduced application may be part of the reason why there was only a single detection following the 2008 application, as compared to the 24 following the application in 2000, where the average yearly concentrations reduced from 0.03 to less than 0.01 µg/l over the eight-year period. An additional explanation for the reduced leaching could also be that percolation following application was much lower in 2008 than in 2000 (see Table 9).

The maximum allowed doses of metamitron were not regulated between the 2000 and the 2008 application remaining at 2,100 g/ha of metamitron. However, in the PLAP only 1,400 g/ha was applied in 2008. Although both metamitron and its degradation product metamitron-desamino were found in drainage (Table 9, Figure 21B and 21C) and groundwater samples (Table A5.3), the number of findings as well as the concentrations were lower than seen after the application in 2000, and the limit of 0.1 μ g/l was in no case exceeded.

The herbicide triflusulfuron was applied concomitantly with metamitron, and the substance as well as three of its metabolites (IN-D8526, IN-M7222 and IN-E7710) was monitored. Triflusulfuron and the metabolite IN-D8526 were not found at all, and IN-M7222 only once (Table A5.3). IN-E7710 was detected on four occasions (Figure 21E and Table 18).

Fluazifop-P-butyl, a herbicide used against monocotyledons, in this case couch grass (Agropyrum repens, L.), has been included in the PLAP several times over the past 10 years. As fluazifop-P-butyl rapidly degrades, focus has so far been on its degradation product fluazifop-P (free acid). Similar to the 2000/2001 growing season at Silstrup (Kjær et al., 2003), this compound, was found in neither drainage water (Figure 22B) nor groundwater (Figure 22D). When applying fluazifop-P-butyl in July 2008 and including its degradation product TFMP in the monitoring programme, a different picture emerged (Figure 22C and Figure 22E). At the onset of the drainage flow on 11 September 2008, a concentration of 0.52 µg/l TFMP was found. Concentrations remained above 0.1 µg/l throughout the period of drainage runoff. Further, TFMP was found in the screens of the vertical monitoring wells M5.1 (1.5 to 2.5 m b.g.s) and M5.2 (2.5 to 3.5 m b.g.s.) more than one month prior to the detection in the drainage water, in concentrations of 0.11 and 0.064 μ g/l, respectively (Figure 22C). With the groundwater table minimum 1.6 m b.g.s., the root zone being relatively dry, and with percolation 1 m b.g.s. in July-August 2008 (Figure 17 and Figure 22A), this indicates pronounced macropore transport bypassing the tile drainage system.



Figure 22. Precipitation and simulated percolation 1 m b.g.s. (A) together with the concentration of fluazifop-P (free acid) (B) and TFMP (C) in the drainage runoff, and the concentration of fluazifop-P (free acid) (D) and TFMP (E) in the groundwater monitoring screens at **Silstrup**. The green vertical lines indicate the dates of fluazifop-P-butyl applications. Values below the detection limit of 0.01 μ g/l are shown as 0.01 μ g/l (all graphs) and further represented by open symbols in A, B and C.

During the summer of 2009 there was no drainage flow. However, at the onset of flow in the autumn of 2009, TFMP was still present (Figure 22C), the last detection being 0.023 μ g/l on 2 December 2009. Since 18 March 2009 there has been no exceedance of the 0.1 μ g/l in the drainage water, and since 1 April 2009 in the groundwater. The most recent detection of TFMP was 0.03 μ g/l on 1 December 2009. The average leaching concentration in drainage water within the first year of application of fluazifop-p-butyl, amounted to 0.22 μ g/l of TFMP (Table 9). Maximum concentration of TFMP in the groundwater at Silstrup was 0.29 μ g/l (Table 20).

Bifenox, applied in the spring barley on 9 September 2009, was found only in the sampling on 30 September in five screens of three different monitoring wells (Figure 23B); in all but one, the concentrations were less than 0.1 μ g/l. In the groundwater, bifenox acid was found in screens from both horizontal and vertical wells (Figure 23E). Four of the five detections from the horizontal wells exceeded 0.1 μ g/l, maximum, concentrations being 0.86 μ g/l. Concentrations found in the vertical wells were even higher, amounting to 3.1 μ g/l. The bifenox acid appeared in the groundwater before it was found in drainage water, in this case seven days earlier.

Bentazone applied on 19 May 2009 had also been applied in May 2003. Although the amount of active ingredient was the same in both years, the 2003 application was followed by concentrations that were above $0.1 \,\mu g/l$ in both drainage and groundwater (Kjær et all, 2005), as opposed to the 2009 application where $0.1 \,\mu g/l$ was never exceeded (Figure 24B, 24E and Table 9). These results may reflect the importance of different climatic conditions following a spraying.

Azoxystrobin, applied on 24 June 2009, as well as it metabolite CyPM were found in drainage water (Figure 24C and 24D). The concentrations of the metabolite were generally higher than those of the parent compound. All concentrations but one of CyPM were below 0.1 μ g/l. Average yearly concentrations in drainage water amounted to 0.01 and 0.06 for azoxystrobin and CyPM, respectively (Table 9), which is about the same as for the two previous applications in 2004 and 2005 (Table 9). Whereas there were no detections of azoxystrobin in the groundwater, CyPM could be found in both horizontal and vertical wells, concentrations ranging between 0.013 and 0.086 μ g/l and 0.011 and 0.1 μ g/l, respectively.

When evaluating the leaching occurring during the 2009/2010 drainage season (Figure 21–24) it should be noted that the large drainage event that took place during the snowmelt on 12 March 2010 (28 mm/d) could not be sampled due to technical problems caused by the extreme intensity of the drainage runoff. Likewise the drainage runoff could not be measured, and the estimated value for this day is likely to be overestimated (see section 4.2.1 for details).



Figure 23. Precipitation and simulated percolation 1 m b.g.s. (A) together with the concentration of bifenox (B) and bifenox acid (C) in the drainage runoff, and the concentration of bifenox (D) and bifenox acid (E) in the groundwater monitoring screens at **Silstrup**. The green vertical line indicates the date of bifenox application. Values below the detection limit of 0.01 μ g/l are shown as 0.01 μ g/l (all graphs) and further represented as open symbols in B and C.



Figure 24. Precipitation and simulated percolation 1 m b.g.s. (A) together with the concentration of bentazone (B), azoxystrobin (C) and CyPM (D) in the drainage runoff, and the concentration of bentazone (E) and CyPM (F) in the groundwater monitoring screens at **Silstrup**. The green vertical lines indicate the dates of bentazone and azoxystrobin applications. Values below the detection limit of 0.01 μ g/l are shown as 0.01 μ g/l (all graphs) and further represented as open symbols in B, C and D.

5 Pesticide leaching at Estrup

5.1 Materials and methods

5.1.1 Site description and monitoring design

Estrup is located in central Jutland (Figure 1) west of the Main Stationary Line on a hillisland, i.e. a glacial till preserved from the Weichselian Glaciation. Estrup has thus been exposed to weathering, erosion, leaching and other geomorphological processes for a much longer period than the other sites. The test field covers a cultivated area of 1.26 ha (105 x 120 m) and is virtually flat (Figure 25). The site is highly heterogeneous with considerable variation in both topsoil and aquifer characteristics (Lindhardt et al., 2001). Such heterogeneity is quite common for this geological formation, however. Based on three profiles excavated in the buffer zone bordering the field the soil was classified as Abrupt Argiudoll, Aqua Argiudoll and Fragiaquic Glossudalf (Soil Survey Staff, 1999). The topsoil is characterised as sandy loam with a clay content of 10–20%, and an organic carbon content of 1.7-7.3%. A C-horizon of low permeability also characterises the site. The saturated hydraulic conductivity in the C-horizon is 10⁻⁸ m/s, which is about two orders of magnitude lower than at the other loamy sites (Table 1). The geological structure is complex comprising a clay till core with deposits of different age and composition (Lindhardt et al., 2001). A brief description of the sampling procedure is provided in Appendix 2. The monitoring design and test site are described in detail in Lindhardt et al. (2001) and the analysis methods in Kjær et al. (2002). Please note that the geological conditions only allowed one of the planned horizontal wells to be installed.

5.1.2 Agricultural management

Management practice during the 2005-2009 growing seasons is briefly summarized below and detailed in Appendix 3 (Table A3.4). For information about management practice during the previous monitoring periods, see previous monitoring reports available on http://pesticidvarsling.dk/publ_result/index.html.

On 6 April 2009, 30 kg/ha of KBr was applied as a tracer. Two days later the field was sown with spring barley (cv. Keops), which emerged 10 days later. On 1 May, at the beginning of tillering, the herbicide bifenox was used. When on 14 May six tillers were detectable, bentazone, and MCPA were used against weeds. Only bentazone was included in the monitoring. On 4 June, when the first awns were visible, azoxystrobin was used against fungi. At harvest on 7 August, barley yielded 71.4 hkg/ha of grain (85% DM) and 39.9 hkg/ha of straw (100 DM), the latter being shredded at harvest and ploughed in on 24 August 2009. The yield of barley was about 25% above the average for the year and soil type (Plantedirektoratet, 2009).

On the 24 August 2009 the field was ploughed and rotor-harrowed and sown with winter rape (cv. Cabernet). The following day the herbicide clomazone was applied but not included in the monitoring. The herbicide bifenox was used on 30 September when four leaves had unfolded. Pesticide treatment was applied on 9 October using

cypermethrin, but the substance was not monitored. Due to poor overwintering of the winter rape, the field was partially resown on 20 April 2010 using the spring rape variety Pluto.

An area of 2.265 m² was resown after a rotor-harrowing, whereas an area of 2.412 m² was resown without soil cultivation – direct seeding. The resown area amounted to 37% of the total field area. Thiacloprid was used against pests on 10 May and included in the monitoring programme. Harvest of the rape was a two-step procedure. On 23 August the area grown with winter rape was harvested, yielding 38.3 hkg/ha (91% DM) and 41.8 hkg being shredded at harvest (100% DM). On 23 August the section of the field grown with spring rape was shredded, and the resulting biomass of 11.64 hkg/ha spread on the surface. The field was ploughed on 14 September 2010.



Figure 25. Overview of the **Estrup** site. The innermost white area indicates the cultivated land, while the grey area indicates the surrounding buffer zone. The positions of the various installations are indicated, as is the direction of groundwater flow (by an arrow). Pesticide monitoring is conducted weekly from the drainage system (during period of continuous drainage runoff) and monthly and half-yearly from selected groundwater monitoring wells as described in Table A2.1 in Appendix 2.

5.1.3 Model setup and calibration

The numerical model MACRO (version 5.1, Larsbo *et al.*, 2005) was applied to the Estrup site covering the soil profile to a depth of 5 m b.g.s., always including the groundwater table. The model is used to simulate the water flow in the unsaturated zone during the monitoring period from July 2000 - June 2010 and to establish an annual water balance.

Compared to the setup in Rosenbom et al. (2010b), a year of validation was added to the MACRO setup for the Estrup site. The setup was subsequently calibrated for the monitoring period May 1999 June 2004 and validated for the monitoring period July 2004 June 2010. For this purpose, the following time series have been used: the observed groundwater table measured in the piezometers located in the buffer zone, measured drainage flow, and soil water content measured at two depths (25 and 40 cm b.g.s.) from the soil profile S1 (Figure 25). The TDR probes installed at the other depths yielded unreliable data with saturations far exceeding 100% and unreliable soil water dynamics with increasing soil water content during the drier summer periods (data not shown). No explanation can presently be given for the unreliable data, and they have been excluded from the analysis. The data from the soil profile S2 have also been excluded due to a problem with water ponding above the TDR probes installed at S2, as mentioned in Kjær et al. (2003). Finally, TDR-measurements at 25 cm b.g.s. in February 2010 were discarded given freezing soils (soil temperatures at or below zero degrees Celsius). The soil water content is measured with TDR based on Topp calibration (Topp et al., 1980), which will underestimate the total soil water content at the soil water freezing point as the permittivity of frozen water is much less than that of liquid water (Flerchinger et al., 2006).

Because of the erratic TDR data, calibration data are limited at this site. Data acquisition, model setup as well as results related to simulated bromide transport are described in Barlebo *et al.* (2007).

5.2 Results and discussion

5.2.1 Soil water dynamics and water balances

The model simulations were generally consistent with the observed data (which were limited compared to other PLAP sites, as noted above), indicating a good model description of the overall soil water dynamics in the unsaturated zone (Figure 26). The model provided an acceptable simulation of the overall level of the groundwater table. As in the previous hydrological year, a drop in the measured groundwater table was seen after short periods of low precipitation (Figure 26B). Also here the simulated groundwater table did not seem as sensitive to these short periods of low precipitation and tended not to drop as much as the measured values. Since the subsoil TDR data are limited, a more detailed study of soil water dynamics in these layers is difficult. However, the overall soil water saturation at 25 and 40 cm b.g.s. was captured (Figure 26D and 26E). Nothing special is noted for the groundwater table in the latest monitoring period (July 2009-June 2010). As in previous years (Rosenbom *et al.*, 2010b), the simulated groundwater table often fluctuates slightly above the drain depth during periods of drainage flow.



Figure 26. Soil water dynamics at **Estrup**: Measured precipitation and simulated percolation 0.6 m b.g.s. (A), simulated and measured groundwater level (B), simulated and measured drainage flow (C), and simulated and measured soil saturation (SW sat.) at two different soil depths (D and E). The measured data in B derive from piezometers located in the buffer zone. The measured data in D and E derive from TDR probes installed at S1 (Figure 23). The broken vertical line indicates the beginning of the validation period (July 2004-June 2010).

	Normal		Actual	Measured	Simulated	Groundwater
	precipitation ²⁾	Precipitation	evapotranspiration	drainage	drainage	recharge ³⁾
$1.7.99 - 30.6.00^{1}$	968	1173	466	_	553	154 ⁴⁾
1.7.00-30.6.01	968	887	420	356	340	111
1.7.01-30.6.02	968	1290	516	505	555	270
1.7.02-30.6.03	968	939	466	329	346	144
1.7.03-30.6.04	968	928	499	298	312	131
1.7.04-30.6.05	968	1087	476	525	468	86
1.7.05-30.6.06	968	897	441	258	341	199
1.7.06-30.6.07	968	1365	515	547	618	303
1.7.07-30.6.08	968	1045	478	521	556	46
1.7.08-30.6.09	968	1065	480	523	362	62
1.7.09-30.6.10	968	1190	531	499	522	160

Table 10. Annual water balance for **Estrup** (mm/year). Precipitation is corrected to the soil surface according to the method of Allerup and Madsen (1979).

¹⁾ Monitoring started in April 2000.

²⁾ Normal values based on time series for 1961–1990 corrected to the soil surface.

³⁾ Groundwater recharge is calculated as precipitation minus actual evapotranspiration minus measured drainage.
⁴⁾ Where drainage flow measurements are lacking, simulated drainage flow was used to calculate groundwater recharge.

The simulated drainage (Figure 26C) matched the measured drainage flow quite well. The initiation and magnitude of the spring 2010 drainage period was, however, not well captured. The period preceding this drainage period can be characterised by frozen soil and precipitation in the form of snow – a situation which MACRO has difficulties in describing. Drainage runoff over the whole monitoring period was high compared to that of the other two till sites investigated in the PLAP. This was due to a significantly lower permeability of the C-horizon than of the overlying A and B horizons (see Kjær *et al.* (2005c) for details). Due to the extreme water flow generated during the snowmelt on 28 February 2010, adequate measurement of the drainage flow as well as water sampling was not possible at the Estrup site. A similar situation occurred at the Silstrup site and further information about the technical problems is given in section 4.2.1.

The resulting water balance for Estrup for the entire monitoring period is shown in Table 10. Compared with the previous ten years, the latest hydraulic year July 2009-June 2010 was characterised by having the third-highest precipitation, the highest simulated actual evapotranspiration and medium measured drainage. Even though the model did not capture the snowmelt, there were no large differences between the measured and simulated drainage. Precipitation in this year was characterised by November-December being very wet and January-February and May-June being very dry (Appendix 4). Due to this precipitation pattern, the simulated percolation pattern of the year July 2009-June 2010 left the summer without percolation, the autumn with high percolation, and the winter with a decreasing percolation with scattered periods of both percolation and drainage runoff (Figure 26A, 26B, and 26C).



Figure 27. Bromide concentration at **Estrup**. A and B refer to suction cups located at S1 and S2, respectively. The bromide concentration is also shown for drainage runoff (C) and the horizontal monitoring well H1 (D). In September 2008, bromide measurements in the suction cups were suspended (Appendix 2). The green vertical lines indicate the dates of bromide applications.

5.2.2 Bromide leaching

Bromide has now been applied three times at Estrup. The bromide concentrations measured up to October 2005 (Figure 27 and Figure 28) relate to the bromide applied in spring 2000, as described further in Kjær *et al.* (2003) and Barlebo *et al.* (2007). In March 2009, bromide measurements in the suction cups and monitoring wells M3 and M7 were suspended.



Figure 28. Bromide concentration at **Estrup**. The data derive from the vertical monitoring wells (M2–M6). Screen depth is indicated in m b.g.s. In September 2008, monitoring wells M3 and M7 were suspended (Appendix 2). The green vertical lines indicate the dates of bromide applications.

5.2.3 Pesticide leaching

Monitoring at Estrup began in May 2000. Pesticides and degradation products monitored so far can be seen from Table 11. Pesticide application during the two most recent growing seasons (2008/2009 and 2009/2010) is shown together with precipitation and simulated precipitation in Figure 29. It should be noted that precipitation is corrected to the soil surface according to Allerup and Madsen (1979), whereas percolation (0.6 m b.g.s.) refers to accumulated percolation as simulated with the MACRO model (Section 5.2.1). Moreover, pesticides applied later than April 2010 are not evaluated in this report and hence not included in Table 11.

Table 11. Pesticides analysed at **Estrup** with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from the date of first application until the end of monitoring. 1st month perc. refers to accumulated percolation within the first month after application. C_{mean} refers to average leachate concentration in the drainage water within the first drainage season after application (See Appendix 2 for calculation methods).

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	Date	monitoring	(mm)	(mm)	Perc. (mm)	(µg/l)
Spring barley 2000						
Metsulfuron-methyl (Ally)	May 00	Apr 03	2990	1456	29	< 0.01
- triazinamin	2	1				< 0.02
Flamprop-M-isopropyl (Barnon Plus 3)	May 00	Apr 03	2914	1434	2	0.02
- flamprop (free acid)						0.01
Propiconazole (Tilt Top)	Jun 00	Apr 05	4938	2294	0	0.01
Fenpropimorph (Tilt Top)	Jun 00	Jul 02	2211	1048	0	< 0.01
- fenpropimorphic acid						< 0.02
Dimethoate (Perfekthion 500 S)	Jun 00	Jul 02	2211	1048	0	< 0.01
Pea 2001						
Glyphosate (Roundup Bio)	Oct 00	Jul 10 [†]	10484	4977	123	0.54
- AMPA						0.17
Bentazone (Basagran 480)	May 01	Jul 08	7629	3621	9	0.03
- AIBA						< 0.01
Pendimethalin (Stomp SC)	May 01	Jul 03	2208	1096	9	< 0.01
Pirimicarb (Pirimor G)	Jun 01	Jul 05	4251	1995	10	0.01
- pirimicarb-desmethyl						< 0.02
- pirimicarb-desmethyl-formamido						< 0.02
Winter wheat 2002						
Ioxynil (Oxitril CM)	Nov 01	Jul 03	1580	860	52	$0.04^{1)}$
Bromoxynil (Oxitril CM)	Nov 01	Jul 03	1580	860	52	0.011)
Amidosulfuron (Gratil 75 WG)	Apr 02	Jul 04	2148	928	8	< 0.01
MCPA (Metaxon)	May 02	Jul 04	2091	928	0	< 0.01
- 4-chlor,2-methylphenol	-					< 0.01
Propiconazole (Tilt 250 EC)	May 02	Apr 05	2920	1336	39	0.02
Pirimicarb (Pirimor G)	Jun 02	Jul 05	2982	1403	58	0.01
- pirimicarb-desmethyl						< 0.02
- pirimicarb-desmethyl-formamido		Apr 06				< 0.02

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1.

† Monitoring will continue during the following year. The values for prec. and perc. are accumulated up to July 2006.

¹⁾ Drainage runoff commenced about two and a half months prior to the application of ioxynil and bromoxynil, and the weighted concentrations refer to the period from the date of application until 1 July 2002.

The current report focuses on pesticides applied from 2008 and onwards, while leaching risk of pesticides applied before 2008 has been evaluated in previous monitoring reports (see http://pesticidvarsling.dk/publ_result/index.html).

Azoxystrobin has now been applied three times at Estrup: 22 June 2004, 29 June 2006, and 13 June 2008 (Figure 30). The last application before then was in June 1998 (Lindhardt *et al.*, 2001). Following all three applications azoxystrobin and the metabolite CyPM leached to the depth of the drainage system at the onset of drainage due to infiltration of excess rain. Concentrations in drainage water of both parent and metabolite are shown in Figure 30. At all three applications, the surface had desiccation cracks. The maximum measured concentration of azoxystrobin was 1.4 μ g/l on 24 August 2006 and 2.1 μ g/l of CyPM on 11 September 2008. The picture emerging from Figure 30 is that the leaching of the parent compound to drainage water was always highest within the year of spraying (Figure 30B). The following year concentrations would always be lower, but tended to increase with each new application. Regarding the metabolite CyPM (Figure 30C), concentrations within the year of application were, on average, higher than those of the parent compound.

Table 11 continued. Pesticides analysed at Estrup with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from the date of first application until the end of monitoring. 1st month perc. refers to accumulated percolation within the first month after application. C_{mean} refers to average leachate concentration in the drainage water within the first drainage season after application (See Appendix 2 for calculation methods).

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	Date	monitoring	(mm)	(mm)	perc. (mm)	(µg/l)
Fodder beet 2003						
Glyphosate (Roundup Bio)	Sep 02	Jul 10^{\dagger}	8289	3900	0	0.43
- AMPA						0.19
Ethofumesate (Betanal Optima)	May 03	Apr 06	2901	1371	50	0.11
Metamitron (Goltix WG)	May 03	Apr 06	2901	1371	50	1.1
- metamitron-desamino						0.21
Pirimicarb (Pirimor G)	Jul 03	Jul 05	2071	939	0	< 0.01
- pirimicarb-desmethyl		Jul 05				< 0.01
- pirimicarb-desmethyl-formamido		Apr 06				0.12
Spring barley 2004						
Fluroxypyr (Starane 180)	May 04	Jul 06	2073	1030	0	< 0.02
Azoxystrobin (Amistar)	Jun 04	Jul 08	4452	2209	38	0.12
- CvPM						0.23
Moizo 2005						
Tarbuthylazina (Inter Tarbuthylazin)	May 05	Apr 00	1217	2042	30	0.48
- desetbyl-terbytbylazine	Way 05	Apr 09	4247	2042	32	0.48
- desemvi-terbuthylazine		Jul 09	3338	1628	32	0.11
- 2-nyaroxy-rerbanyazine		$\Delta nr 09$	4247	2042	32	0.02
- desisopropyi-diruzine - 2-hydroxy-desethyl-terhythylazine		Jul 08	3338	1628	32	0.02
Bentazone (Laddok TE)	Jun 05	Jul 08	3338	1628	10	0.18
- AIBA	Juli 05	Ju 1 00	5550	1020	10	<0.10
Glyphosate (Roundup Bio)	Nov 05	Inf 10^{\dagger}	5191	2460	68	$4 04^{1}$
- AMPA	1101 05	Jui 10	5171	2100	00	0.42^{1}
Sector handler 2006						0.12
Spring barley 2006	Jun 06	J.,1 09	2442	1162	0	<0.01
florasulari (Fillius)	Juli 00	Jul 08	2442	1105	0	< 0.01
- Jiorasulam-desmelnyi	Jun 06	1,1 09	2414	1170	0	< 0.05
Azoxystrobin (Amistar)	Juli 00	Jul 08	2414	11/0	0	0.05
- Cyrm						0.15
Winter wheat 2007						a. a. (1)
Mesosulfuron-methyl (Atlantis WG)	Oct 06	Jul 08	2059	1095	63	< 0.01
- mesosulfuron					2	< 0.02
Chlormequat (Cycocel 750)	Apr 07	Jul 08	1337	603	0	<0.01
Epoxiconazole (Opus)	May 07	Jul 08	1199	600	45	0.01
Glyphosate (Roundup Bio)	Sep 07	Jul 10	3006	1393	64	$0.14^{\prime\prime}$
- AMPA						0.10"
Winter wheat 2008	0 / 07	M 10	0706	1201	50	0.021)
CL 152915	Oct 07	Mar 10	2706	1301	52	0.03°
- ULISSOIS Tehneemarcle (Felieum EC 250)	Nev 07	Mag 10	2650	1265	77	0.24°
Serving horles 2000	NOV 07	Mar 10	2038	1203	//	0.45
Diference (Equ. 480 SC)	May 00	1.110^{\dagger}	1227	520	17	0.002
bifenor acid	May 09	Jul 10	1557	320	17	0.002
- vijenos uciu - nitrofan						<pre>0.133</pre>
- nullojen Bentazone (Basagran M75)	May 00	$I_{11} I_{10}^{\dagger}$	1200	504	0	0.01
$\Delta z_{0} x_{0} x_{0} x_{0} $ (Dasagraff W1/3)	Jun 00	$J_{\rm III} 10^{\dagger}$	1290	504	0	0.03
- CvPM	Juli 07	Jul 10	1230	505	U	0.38

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1.

[†] Monitoring will continue during the following year. The values for prec. and perc. are accumulated up to July 2009. ¹⁾ Drainage runoff commenced prior to the application of pesticide and the weighted concentrations refer to the period from the date of application until 1 July the following year.



Figure 29. Application of pesticides included in the monitoring programme and precipitation (primary axis) together with simulated percolation 0.6 m b.g.s. (secondary axis) at **Estrup** in 2008/2009 (upper) and 2009/2010 (lower).

It is notable that concentrations of CyPM in the second year after spraying tended to increase even more with each new application of azoxystrobin. When looking at the groundwater there has so far been just one detection of azoxystrobin (0.011 μ g/l on 24 March 2010) in a horizontal well (data not shown). CyPM was found in the groundwater, in particular during the two last years of monitoring. There have been five detections from vertical wells, ranging between 0.014 and 0.085 μ g/l (Figure 30D and Table A5.4).

Picolinafen was applied on 30 October 2007. Concentrations of picolinafen never exceeded 0.1 μ g/l in drainage water (Figure 31B). However, its degradation product CL153815 did so in several instances (Figure 31C), reaching a maximum of 0.5 μ g/l on 6 December 2007, and was 0.016 μ g/l at its last detection on 3 April 2008. By comparing Figure 31B and Figure 31C, CL153815 can clearly be seen to be more persistent than picolinafen. Nearly a year after application of picolinafen (26 February 2009), CL153815 could be found in the drainage water at a concentration of 0.078 μ g/l.


Figure 30. Precipitation and simulated percolation 0.6 m b.g.s. (A) together with concentration of azoxystrobin (B) and CyPM (C) in the drainage runoff (DR on the secondary axis) at **Estrup**. Detections of CyPM in groundwater monitoring screen are indicated in D. Azoxystrobin was only detected once in groundwater monitoring screens (see text). The green vertical lines indicate the dates of applications. Open symbols in B and C indicate values below the detection limit of 0.01 μ g/l.

It was last detected on 26 February 2009 at 0.011 μ g/l. Neither picolinafen nor CL153815 were detected in groundwater (Table 5.4 in Appendix 5).



Figure 31. Precipitation and simulated percolation (A) together with concentration of picolinafen (B), CL153815 (C), and tebuconazole (E) in the drainage runoff (DR on the secondary axis) at **Estrup** in 2007/2010. The green vertical lines indicate the dates of applicatios. Open symbols indicate values below the detection limit of 0.01 μ g/l. While picolinafen and CL153815 were not detected in any samples from groundwater monitoring, tebuconazole was detected in five samples (see text).

Tebuconazole, applied on 22 October 2007, was seen in drainage water on several occasions and reached a maximum concentration of 2.0 μ g/l on 20 November 2008, over a year after application. Out of 80 drainage water samples analysed, 17 contained concentrations of tebuconazole above 0.1 μ g/l (Figure 31D). Tebuconazole was, detected in five groundwater samples taken more than two years after the application. Maximum concentrations found were 0.11 and 0.12 μ g/l in samples taken from two different screeens of a monitoring well on 24 March 2011 (Table A5.4 in Appendix 5).

The herbicide bifenox was used on 1 May and on 30 September 2009. Less than two weeks later 0.15 μ g/l of bifenox was detected in the drainage water (Figure 32B). Following the second application, bifenox was found twice, this time in concentrations less than 0.1 μ g/l. The metabolite bifenox acid was found in connection with a small drain flow event on 9 September 2009 at a concentration of 1.9 μ g/l (Figure 32C), whereas bifenox was below the detection limit of 0.01 μ g/l. Following the application in September there were only two detections of bifenox, both less than 0.1 μ g/l (Figure

32B), whereas eight out of ten bifenox acid detections were above 0.1 μ g/l (Figure 32C). Bifenox acid leached from the root zone to the drainage system in an average concentration of 0.153 μ g/l in the drainage water, whereas the figure for bifenox was 0.002 μ g/l (Table 11). Neither bifenox nor bifenox acid was found in the groundwater monitoring screens.

The highest concentration in drainage water of bentazone following the 14 May 2009 application was 0.16 μ g/l in connection with a small drain flow event in September 2009 (Figure 32D). None of the subsequent concentrations were above 0.1 μ g/l and the average yearly concentrations amounted to 0.05 μ g/l (Table 11). Bentazone was found four times in groundwater monitoring screens; concentrations did, however, not exceed 0.1 μ g/l (Table A5.4, Appendix 5).



Figure 32. Precipitation and simulated percolation (A) together with concentration of bifenox (B), bifenox acid (C), and bentazone (E) in the drainage runoff (DR on the secondary axis) at **Estrup** in 2004/2010. The green vertical lines indicates the dates of applications. Open symbols indicate values below the detection limit of 0.01 μ g/l. While bifenox and bifenox acid were not detected in any samples from groundwater monitoring, bentazone was detected in four samples (see text).



Figure 33. Precipitation and simulated percolation 0.6 m b.g.s. (A) together with the concentration of glyphosate (B) and AMPA (C) in the drainage runoff (DR. on the secondary axis) at **Estrup**. Data represent a nine-year period including four applications of glyphosate as indicated by the green vertical lines. Open symbols indicate values below the detection limit of 0.01 μ g/l. Detection of glyphosate and AMPA in groundwater monitoring wells is shown in D and E. In the period June 2007 until July 2010, marked with the red outlined box, analytical problems caused the concentration of glyphosate to be underestimated (see text for details).

The herbicide glyphosate has now been applied at Estrup in 2000, 2002, 2005, and 2007 (Figure 33). Following all applications, both glyphosate and AMPA could be found in the drainage water. Out of 400 drainage water samples analysed for glyphosate and AMPA in the period 31 October 2000-19 May 2010, the concentrations of glyphosate and AMPA exceeded 0.1 μ g/l in 89 and 98 samples, respectively (Figure 33B and 33C). In the same period, 708 groundwater samples were analysed for glyphosate and 712 for AMPA. During that period AMPA never exceeded 0.1 μ g/l (Figure 33E and Table A5.4 in Appendix 5), whereas glyphosate did so in three samples, of which two were taken on 7 July 2005 from two different wells, concentrations being 0.67 and 0.59 μ g/l, and

one on 13 January 2010 from a third well with a concentration of 0.17 μ g/l (Figure 33D and Table A5.4 in Appendix 5). In Figure 33, the period June 2007 to July 2010 has been put inside a red box to indicate that within this period, analytical problems caused glyphosate to be underestimated. Results from the external quality assurance reveal that in the period June 2007 to July 2010 the concentrataion of glyphosate may have been underestimated by a factor of up to ~2 as compared to previous periods (See section 7.2.2.).

When comparing the three-year periods following the application of glyphosate in September 2002 and September 2007, a pattern of longevity/persistence seems to emerge, in particular for the metabolite AMPA. Three years following spraying with glyphosate there will still be a leaching of AMPA, whereas that of glyphosate is much less (Figure 33B and Figure 33C). This long-term leaching of AMPA may indicate that AMPA is retained within the soil and gradually released over a very long time, as described in Kjær *et al.* (2005a), or that glyphosate is retained within the soil and then gradually degraded into AMPA. With an increased detection of glyphosate in the groundwater samples at Estrup following high precipitation events in September 2005 (nearly three years after latest application), September 2008 and January 2010 (one and two and a half years after the latest application, respectively), evidence of the latter pattern seems also to be recognizable.

It is remarkable that detections of particularly glyphosate in groundwater monitoring wells seem to increase over the years (Figure 33D). Within the first four years, detections of glyphosate were scarce, and AMPA is not found at all (Figure 33E). Following these four years there is a gradual increase, particularly in detections and concentrations of glyphosate. A similar tendency was observed for AMPA, although detections are less frequent and the concentrations comparatively lower. In this respect it should be noted that there had been no application of glyphosate for at least seven years prior to the 2000 application (Lindhardt *et al.*, 2001).

Pesticide leaching at Estrup is mostly confined to the depth of the drainage system. Apart from AMPA, CyPM, bentazone, desethyl-terbuthylazine, deisopropylatrazine, and glyphosate having been detected in 8, 9, 16, 7, 27, and 39 groundwater samples, respectively, pesticides have only sporadically been detected in groundwater monitoring screens below the depth of the drainage system (Appendix 5, Table A5.4). Due to decreased hydraulic conductivity and a lower degree of preferential flow, transport of water and solutes at Estrup is much slower beneath the drainage system than above it. Slow transport may allow for dispersion, dilution, sorption and degradation, thereby further reducing the deep transport. Compared to the other loamy soils investigated, the retention characteristics at Estrup suggest that the C-horizon (situated beneath the drainage depth) is less permeable with a lower degree of preferential flow occurring through macropores (see Kjær *et al.*, 2005c, for details). An indication thereof are the long periods with groundwater table beyond drainage depth in which an increasing lateral transport to the drainage system and decreased leaching to the deeper groundwater will occur.

6 Pesticide leaching at Faardrup

6.1 Materials and methods

6.1.1 Site description and monitoring design

Faardrup is located in southern Zealand (Figure 1). The test field covers a cultivated area of 2.3 ha (150 x 160 m). The terrain slopes gently to the west by $1-3^{\circ}$ (Figure 34). Based on three profiles in the buffer zone bordering the field, the soil was classified as Haplic Vermudoll, Oxyaquic Hapludoll and Oxyaquic Argiudoll (Soil Survey Staff, 1999). The topsoil is characterised as sandy loam with 14–15% clay and 1.4% organic carbon. Within the upper 1.5 m numerous desiccation cracks coated with clay are present. The test field contains glacial deposits dominated by sandy till to a depth of about 1.5 m overlying a clayey till. The geological description shows that small channels or basins filled with meltwater clay and sand occur both interbedded in the till and as a large structure crossing the test field (Lindhardt *et al.*, 2001). The calcareous matrix and the reduced matrix begin at 1.5 m and 4.2 m b.g.s., respectively.

The dominant direction of groundwater flow is towards the west in the upper part of the aquifer (Figure 34). During the monitoring period the groundwater table was located 1-2 and 2-3 m b.g.s. in the lower and upper parts of the area, respectively. During fieldwork within the 5 m deep test pit it was observed that most of the water entering the pit came from an intensely horizontally-fractured zone in the till at a depth of 1.8-2.5 m. The intensely fractured zone could very well be hydraulically connected to the sand fill in the deep channel, which might facilitate parts of the applied bromide reached the vertical monitoring well (M6) located in the sand-filled basin (Figure 35 and Figure 38), thus indicating that hydraulic contact with the surface in the "basin" does not differ from that in other parts of the test field, and that the basin is a small pond filled with sediments from local sources.

A brief description of the sampling procedure is provided in Appendix 2. The monitoring design and test site are described in detail in Lindhardt *et al.* (2001) and the analysis methods in Kjær *et al.* (2002).



Figure 34. Overview of the **Faardrup** site. The innermost white area indicates the cultivated land, while the grey area indicates the surrounding buffer zone. The positions of the various installations are indicated, as is the direction of groundwater flow (by an arrow). Pesticide monitoring is conducted weekly from the drainage system (during period of continuous drainage runoff) and monthly and half-yearly from selected groundwater monitoring wells as described in Table A2.1 in Appendix 2.



Figure 35. Geological description of Faardrup (Lindhardt et al., 2001).

6.1.2 Agricultural management

Management practice during the two recent growing seasons is briefly summarized below and detailed in Appendix 3 (Table A3.5). For information about management practice during the previous monitoring periods, see previous monitoring reports available on http://pesticidvarsling.dk/publ_result/index.html.

On 26 August 2008, 30 kg/ha of KBr was applied as a tracer. Ploughing of the field took place 1 December 2008. On 5 April 2009 a crop of sugar beets (cv. Palace) was sown, emerging on 16 April. The first weed spraying was done on 24 April, when the first leaf was visible (pinhead-size) and the cotyledons horizontally unfurled, using phenmedipham and metamitron. On 30 April, when the first pair of beet leaves were visible, but not yet unfurled (pea-size), weeds were sprayed with triflusulfuron, metamitron, ethofumesate and phenmedipham. The latter was not included in the monitoring, however. On 11 May, where the plants had four leaves unfurled, weeds were again sprayed with triflusulfuron, metamitron, ethofumesate and phenmedipham, and again the latter was not included in the monitoring. Due to problems with couch grass (Agropyrum repens, L.), cycloxydim was used twice: on 14 May where five leaves had unfurled, and on 17 June when the beets covered from 10-40% of the area. Cycloxydim was not included in the monitoring programme. The sugar beets were harvested on 6 October, yielding 348.23 hkg/ha of beets and 189.3 hkg/ha of top (fresh weight). The top was ploughed in on 1 November 2009.

On 22 April 2010 the field was sown with a mixture of spring barley varieties. The barley was undersown with red fescue (cv. Maximum). When four to six tillers were detectable on the barley it was sprayed with the herbicide bentazone. The fungicide azoxystrobin was applied on 2 July, but not included in the monitoring. The barley was harvested on 21 August yielding 58.5 hkg/ha of grain (85% DM) and approximately 27 hkg/ha of straw (100% DM).

6.1.3 Model setup and calibration

The numerical model MACRO (version 5.1) was applied to the Faardrup site covering the soil profile to a depth of 5 m b.g.s., always including the groundwater table. The model was used to simulate the water flow in the unsaturated zone during the full monitoring period September 1999-June 2010 and to establish an annual water balance.

Compared to the setup in Rosenbom *et al.* (2010b), a year of validation was added to the MACRO setup for the Faardrup site. The setup was accordingly calibrated for the monitoring period May 1999-June 2004 and validated for the monitoring period July 2004-June 2010. For this purpose, the following time series were used: observed groundwater table measured in the piezometers located in the buffer zone, water content measured at three depths (25, 60, and 110 cm b.g.s.) from the two profiles S1 and S2 (Figure 34) and measured drainage flow. Data acquisition and model setup are described in Barlebo *et al.* (2007).

	Normal	D	Actual	Measured	Simulated	Groundwater
	precipitation /	Precipitation	evapotranspiration	drainage	drainage	recharge
1.7.99-30.6.00	626	715	572	192	152	-50
1.7.00-30.6.01	626	639	383	50	35	206
1.7.01-30.6.02	626	810	514	197	201	99
1.7.02-30.6.03	626	636	480	49	72	107
1.7.03-30.6.04	626	685	505	36	19	144
1.7.04-30.6.05	626	671	469	131	55	72
1.7.05-30.6.06	626	557	372	28	16	158
1.7.06-30.6.07	626	796	518	202	212	77
1.7.07-30.6.08	626	645	522	111	65	12
1.7.08-30.6.09	626	713	463	46	21	204
1.7.09-30.6.10	626	624	445	54	48	125

Table 12. Annual water balance for **Faardrup** (mm/year). Precipitation is corrected to the soil surface according to the method of Allerup and Madsen (1979).

¹⁾ Normal values based on time series for 1961–1990.

²⁾ For July 1999-June 2002, July 2003-June 2004, in January and February of both 2005 and 2006, and July 2006-June 2007, measured at the DIAS Flakkebjerg meteorological station located 3 km from the test site (see detailed text above).
 ³⁾ Groundwater recharge is calculated as precipitation minus actual evapotranspiration minus measured drainage.

Due to electronic problems, precipitation measured at Flakkebjerg located 3 km east of Faardrup was used for the monitoring periods: July 1999-June 2002, July 2003-June 2004, January and February of both 2005 and 2006, and July 2006-June 2007. Precipitation measured locally at Faardrup was used for the rest of the monitoring period.

6.2 Results and discussion

6.2.1 Soil water dynamics and water balances

The level and dynamics of the soil water saturation in all three horizons in the hydraulic year July 2009-June 2010 were generally well described by the model (Figure 36D, 36E, and 36F). However, for the summer period 2010 the model underestimated the drop in the measured groundwater table (Figure 36B).

The resulting water balance for Faardrup for the 11 monitoring periods is shown in Table 12. Compared with the previous ten years, the latest hydraulic year July 2009-June 2010 was characterised by having the second-lowest precipitation, the third-lowest simulated actual evapotranspiration, and the sixth-lowest measured and fifth-lowest simulated drainage. Precipitation in this year was characterised by August, January, and February being very dry and November and May being very wet (Appendix 4). Due to this precipitation pattern, the duration of the simulated percolation period of the year July 2009-June 2010 was represented by continuous percolation throughout the period November - June (Figure 36A). Compared to the other years, the climate this year gave rise to a short period, where the groundwater table was a bit higher than the drainage level, causing a low short-term contribution to the drains (Figure 36B and 36C).



Figure 36. Soil water dynamics at **Faardrup**: Measured precipitation and simulated percolation 1 m b.g.s. (A), simulated and measured groundwater level GWT (B), simulated and measured drainage flow (C) and simulated and measured soil water saturation (SW sat.) at three different soil depths (D, E, and F). The measured data in B derive from piezometers located in the buffer zone. The measured data in D, E and F derive from TDR probes installed at S1 and S2 (Figure 32). The broken vertical line indicates the beginning of the validation period (July 2004-June 2010).



Figure 37. Bromide concentrations at **Faardrup**. A and B refer to suction cups located at S1 and S2. The bromide concentration is also shown for drainage runoff (C) and the horizontal monitoring wells (D). In September 2008, bromide measurements in the suction cups were suspended. The green vertical lines indicate the dates of bromide applications.

6.2.2 Bromide leaching

The bromide concentration shown in Figure 37 and Figure 38 relates primarily to the bromide applied in May 2000, as described further in Kjær *et al.* (2003), and further evaluated in Barlebo *et al.* (2007). In August 2008, 30 kg/ha potassium bromide was applied for the second time. In September 2008, bromide measurements in the suction cups and monitoring wells M2 and M7 were suspended. A drastic increase in bromide concentration in M4 and M5 was detected in May-June 2009 (Figure 38).



Figure 38. Bromide concentrations at **Faardrup**. The data derive from the vertical monitoring wells (M2–M7). Screen depth is indicated in m b.g.s. In September 2008, monitoring wells M2 and M7 were suspended (Appendix 2). The green vertical line indicates the dates of bromide applications.



Figure 39. Application of pesticides included in the monitoring programme and precipitation (primary axis) together with simulated percolation (secondary axis) at **Faardrup** in 2008/09 (upper) 2009/2010 (lower).

6.2.3 Pesticide leaching

Monitoring at Faardrup began in September 1999 and presently encompasses several pesticides and their degradation products, as indicated in Table 13. The application time of the pesticides included in the monitoring during the two most recent growing seasons is shown together with precipitation and simulated precipitation in Figure 39. It should be noted that precipitation is corrected to the soil surface according to Allerup and Madsen (1979), whereas percolation (1 m b.g.s.) refers to accumulated values as simulated with the MACRO model. It should also be noted that as e.g. tribenuronmethyl (applied as Express) degrades rapidly, the leaching risk is more associated with its degradation product, triazinamin-methyl. For the same reason it is the degradation product and not the parent compounds that is monitored in the PLAP (Table 13).

Table 13. Pesticides analysed at Faardrup with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from the date of first application (app. date) until the end of monitoring. 1st month perc. refers to accumulated percolation within the first month after application. Cmean refers to average leachate concentration in the drainage water the first drainage season after application (See Appendix 2 for calculation methods). .

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	Date	monitoring	(mm)	(mm)	perc. (mm)	(µg/l)
Winter wheat 1999						
Glyphosate (Roundup 2000)	Aug 99	Apr 03	2526	947	0	< 0.01
- AMPA						< 0.01
Bromoxynil (Briotril)	Oct 99	Apr 02	1738	751	35	< 0.01
Ioxynil (Briotril)	Oct 99	Apr 02	1738	751	35	< 0.01
Fluroxypyr (Starane 180)	Apr 00	Apr 02	1408	494	7	< 0.01
Propiconazole (Tilt Top)	May 00	Jul 03	2151	669	0	< 0.01
Fenpropimorph (Tilt Top)	May 00	Jul 02	1518	491	0	< 0.01
- fenpropimorphic acid						< 0.01
Pirimicarb (Pirimor G)	Jun 00	Jul 03	2066	684	0	< 0.01
- pirimicarb-desmethyl						< 0.01
- pirimicarb-desmethyl-formamido						< 0.02
Sugar beet 2001						
Glyphosate (Roundup 2000)	Oct 00	Jul 03	1747	709	0	< 0.01
- AMPA						0.01
Metamitron (Goltix WG)	May 01	Jul 03	1512	507	4	0.01
- metamitron-desamino	-					0.01
Ethofumesate (Betanal Optima)	May 01	Jul 03	1512	507	4	0.06
Desmedipham (Betanal Optima)	May 01	Jul 03	1512	507	4	< 0.01
- EHPC						< 0.02
Phenmedipham (Betanal Optima)	May 01	Jul 03	1512	507	4	< 0.01
- MHPC						< 0.02
Fluazifop-P-butyl (Fusilade X-tra)	Jun 01	Jul 03	1460	503	0	< 0.01
- fluazifop-P (free acid)						0.02
Pirimicarb (Pirimor G)	Jul 01	Jul 03	1460	503	1	< 0.01
- pirimicarb-desmethyl						< 0.01
- pirimicarb-desmethyl-formamido						< 0.02
Spring barley 2002						
Flamprop-M-isopropyl (Barnon Plus 3)	May 02	Jul 04	1337	333	0	< 0.01
- flamprop-M (free acid)	5					< 0.01
MCPA (Metaxon)	May 02	Jul 04	1358	337	4	< 0.01
- 4-chlor-2-methylphenol	2					< 0.02
- triazinamin-methyl ¹ (Express)	May 02	Jul 04	1358	337	4	< 0.02
Dimethoate (Perfekthion 500 S)	Jun 02	Jul 04	1328	333	0	< 0.01
Propiconazole (Tilt 250 EC)	Jun 02	Jul 04	1328	333	0	< 0.01

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1. ¹⁾ Degradation product of tribenuron-methyl. The parent compound degrades too rapidly to be detected by monitoring.

The current report focuses on the pesticides applied from 2008 and onwards, while the leaching risk of pesticides applied before 2008 has been evaluated in previous monitoring reports (see http://pesticidvarsling.dk/publ_result/index.html).

Table 13 continued. Pesticides analysed at Faardrup with the product used shown in parentheses. Degradation products are in italics. Precipitation (prec.) and percolation (perc.) are accumulated from the date of first application (app. date) until the end of monitoring. 1st month perc. refers to accumulated percolation within the first month after application. C_{mean} refers to average leachate concentration in the drainage water the first drainage season after application (See Appendix 2 for calculation methods). The number of pesticide-positive samples is indicated in parentheses.

Crop and analysed pesticides	Application	End of	Prec.	Perc.	1 st month	C _{mean}
	Date	monitoring	(mm)	(mm)	perc. (mm)	(µg/l)
Winter rape 2003						
Clomazone (Command CS)	Aug 02	Apr 05	1761	509	4	< 0.02
- FMC65317 (propanamide-clomazon)						< 0.02
Winter wheat 2004						
Prosulfocarb (Boxer EC)	Oct 03	Apr 06	1542	454	0	< 0.01
MCPA (Metaxon)	Jun 04	Jul 06	1307	331	0	< 0.01
- 4-chlor,2-methylphenol						< 0.01
Azoxystrobin (Amistar)	Jun 04	Jul 07	2098	636	0	< 0.01
- CyPM						< 0.01
Maize 2005						
Terbuthylazine (Inter-Terbutylazin)	May 05	Jul 08	2078	666	4	0.67
- desethyl-terbuthylazine	May 05	Jul 08	2078	666		0.59
- 2-hydroxy-terbuthylazine	May 05	Jul 08	2078	666		0.04
- desisopropyl-atrazine	May 05	Jul 08	2078	666		0.03
- 2- hydroxy-desethyl-terbuthylazine	May 05	Jul 07	1428	465	4	0.07
Bentazone (Laddok TE)	May 05	Jul 07	1408	464	6	2.82
- AIBA						<0.01
Spring barley 2006						
Fluroxypyr (Starane 180 S)	May 06	Jul 08	1496	524	17	< 0.02
Epoxiconazole (Opus)	Jun 06	Jul 08	1441	507	3	< 0.01
Winter Rape 2007						
Thiamethoxam (Cruiser RAPS)	Aug 06	Jul 08	1304	505	27	< 0.01
- CGA 322704						< 0.02
Propyzamide (Kerb 500 SC)	Feb 07	Apr 09	1476	375	46	0.138^{1}
- RH-24644						$< 0.01^{1/2}$
- RH-24580						$< 0.01^{1/2}$
- КП-24033						<0.01
Winter wheat 2008						
Pendimethalin (Stomp)	Oct 07	Dec 09	1462	451	24	< 0.01
Tebuconazole (Folicur EC 250)	Nov 07	Dec 09	1405	413	56	< 0.01
Sugar beet 2009						
Triflusulfuron (Safari)	Apr 09	Jul 10 [†]	769	210	2	< 0.01
-IN-D8526						< 0.01
-IN-E7710						< 0.01
-IN-M/222	A 00	T 1 10 [†]	7(0)	010	2	< 0.01
Ethorumesate (Ethosan)	Apr 09	Jul 10°	/69	210	2	< 0.01
Metalling (Gollath)	A		140	2111	· •	<0.01

Systematic chemical nomenclature for the analysed pesticides is given in Appendix 1.

[†] Monitoring will continue during the following year. The values for prec. and perc. are accumulated up to July 2009. ¹⁾ Drainage runoff commenced prior to the application of propyzamide and the weighted concentrations refer to the period from the date of application (Feb 07) until 1 July 2007.

Tebuconazole and pendimethalin were applied in 2007 and these pesticides have until now been detected in five and two samples (data not shown, see Rosenbom et al., 2010b), respectively. Measured concentrations, however, never exceeded 0.1 µg/l. Triasulfuron-methyl was applied in April 2009, but neither the parent compound nor its degradations products have so far been detected.

Metamitron, ethofumesate and triflusulfuron were applied April 2009 (Figure 39) and so far neither of the pesticides nor their metabolites have been detected in water samples from Faardrup.

7 Pesticide analysis quality assurance

Reliable results and scientifically valid methods of analysis are essential for the integrity of the present monitoring programme. Consequently, the field monitoring work has been supported by intensive quality assurance entailing continuous evaluation of the analyses employed. Two types of sample are used in the quality control – samples with known pesticide composition and concentration are used for *internal monitoring* of the laboratory method, while *externally spiked samples* are used to incorporate additional procedures such as sample handling, transport and storage. Pesticide analysis quality assurance (QA) data for the period July 2009 to June 2010 are presented below, while those for the preceding monitoring periods are given in previous monitoring reports (available on http://pesticidvarsling.dk/publ_result/index.html).

7.1 Materials and methods

All pesticide analyses were carried out at commercial laboratories selected on the basis of a competitive tender. In order to assure the quality of the analyses, the call for tenders included requirements as to the laboratory's quality assurance (QA) system comprising both an internal and an external control procedure. In addition to specific quality control under the PLAP, the laboratory takes part in the proficiency test scheme employed by the Danish Environmental Protection Agency when approving laboratories for the Nationwide Monitoring and Assessment Programme for the Aquatic and Terrestrial Environments (NOVANA).

7.1.1 Internal QA

With each batch of samples the laboratory analysed one or two control samples prepared at each laboratory as part of their standard method of analysis. The pesticide concentration in the internal QA samples ranged between 0.03–0.13 µg/l. Using these data it was possible to calculate and separate the analytical standard deviation into within-day (S_w), between-day (S_b) and total standard deviation (S_t). Total standard deviation was calculated using the following formula (Wilson 1970, Danish EPA 1997): $s_t = \sqrt{s_w^2 + s_b^2}$

7.1.2 External QA

Every four months, two external control samples were analysed at the laboratories along with the various water samples from the five test sites. Two stock solutions of different concentrations were prepared from two standard mixtures in ampoules prepared by Dr. Ehrenstorfer, Germany (Table 14). Fresh ampoules were used for each set of standard solutions. The standard solutions were prepared two days before a sampling day and stored in darkness and cold until use. For the preparation of stock solutions 150 μ l (low level) or 350 μ l (high level) of the pesticide mixture was pipetted into a preparation glass containing 10 ml of ultrapure water. The glass was closed and shaken thoroughly

and shipped to the staff collecting the samples. The staff finished the preparation of control samples in the field by quantitatively transferring the standard solution to a 3.01 measuring flask. The standard solution was diluted and adjusted to the mark with groundwater from a groundwater well. In the present report period the final concentrations correspond to 50 and 117 μ g/l in the final solution for low and high spike levels, respectively. After a thorough mixing, the control sample was transferred to a sample bottle and transported to the laboratories together with the regular samples. As water sample supply was occasionally limiting at Faardrup, all volumes were reduced by a factor of three for this location, keeping the concentrations in the final control samples identical to the other locations.

The pesticide concentration in the solution is indicated in Table 14. Blank samples consisting of HPLC water were also included in the external QA procedure every month. All samples included in the control sample were labelled with coded reference numbers, so that the laboratory was unaware of which samples were controls and which were blanks.

 Table 14. Pesticide concentrations in both the original ampoules and in the resulting high-level and low-level external control samples.

Commenced	A		II: -h local contact	T and land a setual
Compound	Ampoule		High-level control	Low-level control
	Concentration (mg/l)	Ampoule	(µg/l)	(µg/l)
AMPA	1.000	2	0.117	0.050
CyPM	1.000	1	0.117	0.050
Bentazone	1.000	1	0.117	0.050
Bifenox (free acid)	1.000	1	0.117	0.050
CL153815	1.000	1	0.117	0.050
Epoxiconazole	1.000	1	0.117	0.050
Ethofumesate	1.000	1	0.117	0.050
Glyphosate	1.000	2	0.117	0.050
IN70941 (PPU)	1.000	1	0.117	0.050
IN-M7222	1.000	1	0.117	0.050
Metsulfuron-methyl	1.000	1	0.117	0.050
Metamirton	1.000	1	0.117	0.050
Pendimethalin	1.000	1	0.117	0.050
Tebuconazole	1.000	1	0.117	0.050
TFMP	1.000	1	0.117	0.050

7.2 **Results and discussion**

7.2.1 Internal QA

Ideally, the analytical procedure provides precise and accurate results. However, in the real world results from analysis are subject to a certain standard deviation. Such standard deviation may be the combined result of several contributing factors. Overall, the accuracy of an analytical result reflects two types of error: Random errors related to precision and systematic errors relating to bias. In a programme like PLAP it is relevant to consider possible changes in analytical "reliability over time". As these errors may change over time it is relevant to distinguish between standard deviations resulting from *within-day* variation as opposed to those associated with *between-day* variation in the analytical result. To this end, control samples are included in the analytical process as described above. Thus, by means of statistical analysis of the internal QA data it is possible to separate and estimate the different causes of the analytical variation in two categories: day-to-day variation and within-day variation (Miller *et al.*, 2000; Funk *et al.*, 1995). This kind of analysis can provide an indication of the reliability of the

analytical results used in the PLAP. The statistical tool used is an analysis of variance (ANOVA) and encompasses all duplicate pesticide analyses, single analyses being excluded. The analysis can be divided into three stages:

- 1. **Normality**: An initial test for normality is made as this is an underlying assumption for the one-way ANOVA.
- 2. Between-day contribution: Explained simply, this test will reveal any day-today contribution to the variance in the measurements. If there is none, the total standard deviation can be considered to be attributable to the within-day error of the analysis. For this purpose an ANOVA-based test is used to determine if the between-day standard deviation (S_b) differs significantly from 0 (this test is made as an F-test with the H_0 : between-day mean square = within-day mean square).
- 3. Calculating standard deviations: If the F-test described above reveals a contribution from the between-day standard deviation (S_b) , it is relevant to calculate three values: The within-day standard deviation S_w , the between-day standard deviation S_b , and the total standard deviation S_t .

As the error associated with the analytical result is likely to be highly dependent on the compound analysed, the QA applied is pesticide-specific. The results of the internal QA statistical analysis for each pesticide are presented in Table 15. For reference, estimated S_b values are listed for all pesticides, including those for which the between-day variance is not significantly greater than the within-day variance. ANOVA details and variance estimates are also included, even for pesticides where the requirement for normality is not fulfilled. Such data should obviously be interpreted with caution.

As a rule of thumb, the between-day standard deviation should be no more than double the within-day standard deviation. From Table 15 it can be seen that S_b/S_w ratios greater than two were observed for several compounds. For these compounds, the results indicate that day-to-day variation makes a significant contribution. Among the compounds meeting the normality requirement, the S_b/Sw ratio is highest for PPU, desethyl-terbuthylazine and desisopropyl-atrazine. When all compounds are considered, a particularly high S_b/S_w ratio is apparent for desethyl-terbuthylazine and triazinamine. Such relatively high ratios can be caused by very low within-day standard deviations, i.e. within each laboratory day, the variation on the analysis is small compared to the other compounds, whereas the variation between days is comparable to the other compounds analysed. This is the case for desethyl-terbuthylazine. Thus, low values of Sw rather than critical values of Sb caused the high ratios, as reflected by a reasonably low St. In contrast, for the compound triazinamine and others it is apparent that it is the between-day (S_b) contribution that is causing a high S_b/Sw ratio. As reflected by the data in Table 15, the three compounds with the highest observed between-day contribution were triazinamine, bifenox acid and picolinafen.

The total standard deviations (S_t) of the various analyses of pesticides and degradation products lie within the range 0.005-0.652 µg/l, the highest value being observed for triazinamin (max S_t for compounds other than triazinamine was 0.044 µg/l). In general, the data suggest that the analytical procedure used for the quantification of triazinamine may benefit from a critical review. Excluding the triazinamine data, the overall mean S_t was 0.014 µg/l.

Table 15. Internal QA of pesticide analyses carried out in the period 1.7.2008-30.6.2009. Results of the test for normality, one-way analysis of variance (ANOVA), the estimated values of standard deviations (w: within-day, b: between-day, t: total – see text for details), pesticide concentration in internal QA sample (Conc.) and number of duplicate samples (n) are given for each pesticide. For test the P value α =0.05 was used.

Compound	Normal distribution $\alpha = 0.05$	Significant Sb Between day contribution ANOVA	Sw (µg/l)	Sb (µg/l)	St (µg/l)	Ratio Sb/Sw	N	Conc. (µg/l)
AMPA *		u=0.05	0.016	0.006	0.017	0.38	27	0.03
Azoxystrohin	Ves	Ves	0.010	0.000	0.0017	2.93	27 48	0.05
Bentazone	yes	yes	0.002	0.000	0.000	5.83	48	0.05
Bifenox		yes	0.001	0.005	0.005	1.43	34	0.05
Bifenox acid*		yes	0.007	0.010	0.012	2 33	30	0.05
CI 153815*		yes	0.017	0.079	0.031	2.33	26	0.10
Desethyl-terbuthylazine*		yes	0.0012	0.027	0.051	17.28	20	0.15
Epoxiconazole	Ves	yes	0.001	0.007	0.008	2 47	7	0.05
Glyphosate	yes	yes	0.000	0.007	0.011	0.56	27	0.03
IN70941*	Ves	Ves	0.010	0.003	0.011	4 88	$\frac{27}{24}$	0.05
IN70942*	yes	yes	0.003	0.004	0.004	4.00 6.78	$\frac{24}{24}$	0.05
Introp-12	Ves	yes	0.001	0.000	0.000	0.70	2 - 5	0.05
Mesosulfuron*	yes	Ves	0.003	0.000	0.005	2 49	5	0.05
Mesosulfuron-methyl	yes	yes	0.003	0.000	0.000	1 70	6	0.10
Metribuzin-diketo*	yes	yes	0.000	0.014	0.017	3 54	12	0.05
Metsulfuron_methyl*	yes	yes	0.002	0.000	0.000	0.72	5	0.05
Nitrofen*	VAS	VAS	0.010	0.012	0.020	1 30	34	0.05
Dandimethalin	yes	yes	0.002	0.010	0.010	4.39	14	0.05
Picelinefen		Noc	0.009	0.000	0.011	1.76	14 26	0.05
Tehuconazola		yes	0.010	0.029	0.000	1.70	20 29	0.11
		yes	0.003	0.008	0.008	5.00	38	0.05
1 Hazinamin*		yes	0.051	0.050	0.052	12.73	15	0.11

*Degradation product.

7.2.2 External QA

As part of the quality control a set of blanks are analysed to evaluate the possibility of false positive findings in the programme. From these results it can be concluded that contamination of samples during collection, storage and analysis is not likely to occur. In a total of 26 blank samples consisting of HPLC water a trace of glyphosate (< 0.02 μ g/l well below the residue limit of 0.1 μ g/l and close to the detection limit) was found in a single sample. Samples found to contain pesticides or their degradation product are thus regarded as true positive findings.

Table 16 provides an overview of the recovery of all externally spiked samples. As the results for each field site in Table 16 are based on only a few observations for each concentration level (high/low), the data should not be interpreted too rigorously.

.	Tyls	strup	Jynd	evad	Sils	trup	Est	rup	Faar	drup	Average	$n_{\rm low}/n$
	Low	High	Low	High	Low	High	Low	High	Low	High		high
	LOW	mgn	LOW	mgn	LOW	mgn		111g11 06	LOW	mgn	74	2/2
AMPA*	100	100	100	100			02	100	10(111	/4	3/3 10/10
Bentazone	102	102	100	102			110	102	126	111	105	10/10
Bifenox acid*				68				60			64	[0/5]
CL153815*			72	70			78	75			75	3/3
CyPM*	87	90					67	64			78	6/6
Epoxiconazole			174	153							164	1/1
Ethofumesate					99	99			104	102	101	5/6
Glyphosate							58	54			57	3/3
IN70941(PPU)*	34	26	61	41							42	4/6
IN-M7222*					130	122			116	108	120	5/6
Metamitron					74	74			87	79	78	5/6
Metsulfuron-methyl*					67	68					68	3/3
Pendimethalin	84	82							100	102	92	2/2
Tebuconazole	113	111	135	123			104	102	118	119	115	7/7
TFMP*					157	133					146	3/3

Table 16. Externally spiked samples. Average recovery (%) of the nominal concentration at low/high concentration level indicated for each site. n_{low} and n_{high} refer to the total number of samples being spiked at low and high concentrations, respectively. Bold font is used for recoveries outside the range 70% to 120%.

*Degradation produkt.

Whereas the recovery of the most spiked compounds in the samples is generally good (i.e. in the range 70% to 120%), the broad range of average recoveries indicates that for some compounds there may be reason for concern. In the current reporting period one exception has been made concerning spiking, as traces of Bifenox acid, CyPM and bentazone were found in the groundwater used for spiking at Silstrup, and data from this site have been omitted due to this bias. The somewhat low levels of recovery found for bifenox acid at Jyndevad and Estrup indicate that the the QA needs to be reevaluated for this compound when more results are available (at present the QA is based on five samples). Also, the detection limit for Bifenox acid at 0.05 makes the QA spiking less informative at this level compared to the other compounds in the programme that have detection limits around 0.01 and 0.02 Considering the low recovery of glyphosate, identified in previous reports, analytical procedures have now been optimised and implemented in the programme with effect from 1 July 2010 and will thus be reflected in the next reporting period. However, it is anticipated that the QA data in the reports to follow will verify the improved methodologies for this compound. In the programme a shift from GC/MS to LC/MS analytical procedure was made in June 2007. The low recoveries relate to the analytical LC/MS procedure used in the programme during the the period June 2007 to July 2010. During this period the concentration of glyphosate may have been underestimated. Due to the analytical cause of the problem, it is not possible to give an exact value for such a possible underestimation. However, a preliminary estimate has been made using all the QA data available for glyphosate and AMPA, i.e. all spiked QA samples analysed for these two compounds in the period October 2007 to June 2010. For glyphosate the mean recovery was 86% (n=73, std dev 24%) compared to 40% (n=18, std dev 16%) using LC/MS. For AMPA the GC/MS recoveries were 76% (n=31, std dev 24%) compared to 68% (n=16, std dev 15%) using LC/MS. A simple t-test indicates that there is a difference in the recoveries for glyphosate (P < 0.001), whereas no significant difference was found for the AMPA recovery. A rough estimate based on the mean recovery obtained using the two glyphosate methods would indicate that analytical results reported in the period June 2007 to July 2010 may have underestimated the concentration of glyphosate by a factor of ~2 as compared to previours result based on a GC/MS analysis.

All the compounds included in the spiking procedure (Table 14) were detected in the laboratory. Additionally, a number of compounds were measured at the threshold of detection of the analytical procedure (i.e. close to 0.01 µg/l). The occurrence of a limited number of false positives is expected when analysing environmental matrices, and these findings do not cause a general concern in relation to the reliability of the analytical procedures used in the programme. Also, there were 12 reportings with a mean content of 0.05 µg/l for the compound PPU-desamino (IN70942), which was not included in the spiking ampoule. The compound PPU -desamino (IN70942) is a degradation product of PPU (IN70941), which was included in the spiking, and the presence of a degradation product indicates that the estimated leaching of PPU (IN70941) may be underestimated as described in Rosenborn et al. (2009). The same aspect relates to 11 reportings of metamitron-desamino, a degradation product of metamitron that was included in the spike solution. Thus, these compounds may be formed from compounds present in the spiking solution, In general, since the levels were low and in the range of the detection limit and well below the residue limit, these findings do not cause concern for the overall quality of the programme.

During the 2009/2010 monitoring period a total of five pesticides and nine degradation products were detected in samples from the experimental fields, and the external and internal QA data relating to these particular pesticides/degradation products are of special interest. These data (when available) are therefore illustrated in Appendix 6.

7.3 Summary and concluding remarks

The QA system showed that:

- The internal QA indicates that the reproducibility of the pesticide analyses was good with total standard deviation (St) in the range 0.005-0.044 μ g/l, except for triazinamin with a large St of 0.652 caused in particular by Sb.
- As demonstrated by the external QA, recovery was generally good in externally spiked samples. Low recovery of glyphosate was, however, observed in all samples, but a revision of the analytical procedure being implemented in parallel with the current reporting period is anticipated to solve these issues in the PLAP-programme.
- Contamination of samples during collection, storage and analysis is not likely to occur. In a single sample out of a total of 26 blank samples a trace of glyphosate was reported. No other pesticides or pesticide degradation products were detected.

8 Summary of monitoring results

This section summarizes the monitoring data from the entire monitoring period, i.e. both data from the two most recent monitoring years (detailed in this report) and data from the previous monitoring years (detailed in previous reports available on http://pesticidvarsling.dk/publ_result/index.html). Pesticide detections in samples from the drainage systems, suction cups and monitoring wells are detailed in Appendix 5. The monitoring data in 1 m b.g.s. (collected in drains and suction cups) reveal that the applied pesticides exhibit three different leaching patterns – no leaching, slight leaching and pronounced leaching (Table 17). Pronounced leaching in 1 m b.g.s. is defined as root zone leaching (1 m b.g.s.) exceeding an average concentration of 0.1 µg/l within the first season after application. On sandy and loamy soils, leaching is determined as the weighted average concentration in soil water and drainage water, respectively (Appendix 2). The monitoring data from the groundwater monitoring screens is divided into three categories: no detection of the pesticide (or its degradation products), detections of the pesticide (or its degradation products) not exceeding 0.1 µg/l, and detections of the pesticide (or its degradation products) exceeding 0.1 µg/l (Table 19). It should be noted, though, that the present evaluation of the leaching risk of some of these pesticides is still preliminary as their potential leaching period extends beyond the current monitoring period. This applies to those pesticides marked with a single asterisk in Table 17 and 19. Fourteen of the applied pesticides (or their degradation products) exhibited pronounced root zone leaching and 12 of these were also detected in the groundwater monitoring screens in concentrations exceeding $0.1 \,\mu g/l$.

• Azoxystrobin, and in particular its degradation product CyPM, leached from the root zone (1 m b.g.s.) in high average concentrations at the loamy sites Silstrup and Estrup. CyPM leached into the drainage water in average concentrations exceeding 0.1 µg/l at both the Silstrup and Estrup sites, while azoxystrobin only leached in concentrations exceeding 0.1 µg/l at Estrup (Table 17 and 18). At both sites, leaching of azoxystrobin and CyPM has hitherto mostly been confined to the depth of the drainage system, and they have rarely been detected in monitoring screens situated below drainage depth (Tables 19 and 20). However, detection of CyPM in groundwater monitoring wells has gradually increased over time with highest numbers of detection found after the latest applications (2009 at Silstrup, Figure 24 and 2008 at Estrup, Figure 30). Apart from one sample, however, concentrations detected in only four samples from the drainage water, and in no samples from the sandy Jyndevad site (Appendix 5).

Table 17. Root zone leaching (**1 m b.g.s.**) of pesticides or their degradation products at the five PLAP sites. An asterisk indicates pesticides that have been included in the monitoring programme for less than two years. The colours indicate the degree of leaching and the letters H, F, I, and GR indicate the type of pesticide: herbicide, fungicide, insecticide and growth retardant, respectively. Pesticides applied in spring 2010 are not included in the table.

Azoxystrobin (F) Bentazone (H) Bifenox (H) Ethofumesate (H) Fluazifop-P-butyl (H) Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	(Sandy soil) 2) *	(Sandy soil) * 2) 1) 2) 2)	(Loamy soil) * * * *	(Loamy soil) * *	(Loamy soil) 2)
Azoxystrobin (F) Bentazone (H) Bifenox (H) Ethofumesate (H) Fluazifop-P-butyl (H) Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	2)	* 2) 1) 2)	*	*	2)
Bentazone (H) Bifenox (H) Ethofumesate (H) Fluazifop-P-butyl (H) Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	2)	* 2) 1) 2)	*	*	2)
Bifenox (H) Ethofumesate (H) Fluazifop-P-butyl (H) Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	2)	* 2) 1) 2)	*	*	2)
Ethofumesate (H) Fluazifop-P-butyl (H) Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	2)	2) 1) 2)		*	2)
Fluazifop-P-butyl (H) Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	2)	2) 1) 2)		*	2)
Glyphosate (H) Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	*	ı) 		*	
Metamitron (H) Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	*	1) 			
Metribuzin (H) Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)	*	i) 2)			
Picolinafen (H) Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)		2)			
Pirimicarb (I) Propyzamide (H) Rimsulfuron (H)		2)			
Propyzamide (H) Rimsulfuron (H)		2)			
Rimsulfuron (H)		2)			
		2)			
Terbuthylazine (H)		2)			
Tebuconazole (F)		2)			
Amidosulfuron (H)				2)	
Bromoxynil (H)					
Clomazone (H)			•		
Dimethoate (I)					
Epoxiconazole (F)					
Flamprop-M-isopropyl (H)					
Fluroxypyr (H)					
Ioxvnil (H)					
Mancozeb(F)					
MCPA (H)					
Mesosulfuron-methyl (H)					
Pendimethalin (H)					
Phenmedipham (H)					
Propiconazole (F)					
Prosulfocarb (H)					
Pyridate (H)					
Triflusulfuron (H)					*
Chlormequat (GR)					
Clopyralid (H)					
Desmedipham (H)					
Fenpropimorph (F)					
Florasulam (H)					
Iodosulfuron-methyl-sodium (H)					
Linuron (H)					
Metsulfuron-methyl (H)					
Thiamethoxam (I)					
Tribenuron-methyl (H)					
Triasulfuron (H)					
Linuron (H) Metsulfuron-methyl (H) Thiamethoxam (I) Tribenuron-methyl (H) Triasulfuron (H)					

* Potential leaching period extends beyond the current monitoring period.

¹⁾ Derived from application before May 1999 (see Kjær *et al.*, 2002).

²⁾ Degradation products are not monitored (see text).

Pesticide (or its degradation products) leached 1 m b.g.s. in average concentrations exceeding 0.1 μ g/l within the first season after application.

Pesticide (or its degradation products) was detected in either several (more than three) consecutive samples or in a single sample in concentrations exceeding 0.1 μ g/l; average concentration (1 m b.g.s.) below 0.1 μ g/l within the first season after application.

Pesticide either not detected or only detected in very few samples in concentrations below 0.1 µg/l.

Table 18. Number of samples from 1 m b.g.s. in which the various pesticides and their degradation products were detected at each site with the maximum concentration $(\mu g/l)$ in parentheses. The table only encompasses those pesticides/degradation products detected in either several (more than three) consecutive samples or in a single sample in concentrations exceeding 0.1 µg/l. Degradation products are indicated in italics. Pesticides applied in spring 2010 are not included.

	Tylstrup	Jyndevad	Silstrup	Estrup	Faardrup
Azoxystrobin	0	0	10(0.034)	100(1.4)	0
-CyPM	0	0	64(0.34)	184(2.1)	4(0.059)
Bentazone	1(0.012)	39(1.6)	45(6.4)	145(20)	18(43)
-AIBA	0	2(0.034)	0	1(0.06)	1(0.057)
Bifenox		2(0.036)	1(0.027)	3(0.15)	
-Bifenox acid		1(0.1)	13(4.2)	11(1.9)	
-Nitrofen		0	1(0.023)		
Ethofumesate			15(0.227)	35(3.362)	13(12)
-fluazifop- $P^{2)}$	0	0	0	0	8(3.8)
$-TFMP^{2}$			21(0.52)		
Glyphosate		0	67(4.7)	251(31)	4(0.093)
-AMPA		1(0.014)	122(0.35)	356(1.6)	10(0.11)
Metamitron			31(0.315)	42(26.369)	12(1.7)
-metamitron-desamino			46(0.399)	49(5.549)	16(2.5)
Metribuzin	2(0.024)	0			
-metribuzin-desamino-diketo	81(2.1)	0			
-metribuzin-diketo	242(0.69)	3(0.088)			
Picolinafen		1(0.015)		17(0.07)	
-CL153815	_	0		31(0.5)	
Pirimicarb	0	0	14(0.054)	39(0.077)	7(0.056)
-pirimicarb-desmethyl	0	1(0.011)	1(0.052)	0	6(0.053)
-pirimicarb-desmethyl-	0	0	0	26(0.379)	3(0.039)
Propyzamide	0		23(1.6)		4(0.51)
-RH24580	0		2(0.016)		0
-RH24644	0		15(0.051)		4(0.022)
- <i>RH24</i> 635	$0 \\ 100(0, 15)$	154(0.20)	0		1(0.017)
$-PPU^{s_{j}}$	122(0.15)	154(0.29)			
-PPU-aesamino"	35(0.042)	89(0.13)	(0(1 55))	111/11)	(1)(10)
I erbuthylazine	0	19(0.05()	60(1.55)	111(11) 145(9,2)	41(10)
-aesetnyiterbutnyiazine	2(0.012) 17(0.042)	18(0.050)	108(1.08)	145(8.2)	89(8.3)
- desisopropyl-alrazine	1/(0.042) 5(0.016)		43(0.041)	71(0.44)	23(0.50)
-2-hydroxy-desethyl-	5(0.010) 1(0.04)		28(0.11) $26(0.020)^{*}$	80(0.3)	$\delta(1)$
-2-nyaroxy-terbutnytazin Tebuconazole	1(0.04)	0	20(0.039)	$\frac{87(0.99)}{41(2)}$	21(0.38) 4(0.045)
Amidosulfuron	0	$\frac{0}{3(0,11)}$			4(0.043)
Bromoxynil	0	0		3(0,6)	0
Clomazone	Ő	0		5(0.0)	1(028)
-FMC65317(Propanamide-clomazone)	ŏ				0
Dimethoate	ŏ	0	1(1.417)	0	ŏ
Epoxiconazole	Õ	Õ	0	13(0.39)	Õ
ETU ¹⁾	7(0.038)				
Flamprop-M-isopropyl	0		12(0.109)	20(0.069)	1(0.037)
-flamprop (free acid)	0		7(0.096)	13(0.031)	1(0.089)
Fluroxypyr	0	0	0	3(0.025)	1(0.19)
Ioxynil	0	0		20(0.25)	1(0.011)
MCPA		0	0	11(3.894)	2(0.28)
-4-chlor-2-methylphenol		0	0	1(0.046)	1(0.24)
Mesosulfuron-methyl		0		13(0.059)	
Pendimethalin	0	0	14(0.064)		2(0.041)
Phenmedipham			0		0
-MHPC			0		2(0.19)
Propiconazole	0	0	6(0.033)	25(0.862)	0
Prosulfocarb			5(0.18)		0
Pyridate		0	0		
-PHCP			4(2.69)		
Triflusulfuron			0		0
-IN-E7710			5(0.014)		0

¹Degradation product of mancozeb. ²⁾Degradation product of fluazifop-P-butyl ³⁾Degradation product of rimsulfuron.

*Included in the monitoring at Silstrup from February 2003, eight months after application of terbuthylazine.

Table 19. Detections of pesticides and their degradation products in water samples from the groundwater monitoring screens at the five PLAP sites (see table 20 for details). An asterisk indicates pesticides that have been included in the monitoring programme for less than two years. The colours indicate the level of detection (see below) and the letters H, F, I, and GR indicate the type of pesticide: herbicide, fungicide, insecticide and growth retardant, respectively. Pesticides applied in spring 2010 are not included in the table.

	Tylstrup	Jyndevad	Silstrup	Estrup	Faardrup
	(Sandy soil)	(Sandy soil)	(Loamy soil)	(Loamy soil)	(Loamy soil)
Azoxystrobin (F)					
Bentazone (H)					
Bifenox		*	*	*	
Ethofumesate (H)					
Fluazifop-P-butyl (H)	2)	2)			2)
Glyphosate (H)					
Metamitron (H)					
Metribuzin (H)		1)			
Picolinafen (H)					
Pirimicarb (I)					
Propyzamide (H)					
Rimsulfuron (H)					
Terbuthylazine (H)					
Tebuconazole (F)					
Amidosulfuron (H)		2)		2)	
Bromoxynil (H)					
Clomazone (H)					
Dimethoate (I)					
Epoxiconazole (F)					
Flamprop-M-isopropyl (H)					
Fluroxypyr (H)					
Ioxynil (H)					
Mancozeb (F)					
MCPA (H)					
Mesosulfuron-methyl (H)					
Pendimethalin (H)					
Phenmedipham (H)					
Propiconazole (F)					
Prosulfocarb (H)					
Pyridate (H)					
Triflusulfuron (H)					*
Chlormequat (GR)					
Clopyralid (H)					
Desmedipham (H)					
Fenpropimorph (F)					
Florasulam (H)					
Iodosulfuron-methyl-sodium (H)					
Linuron (H)					
Metsulfuron-methyl (H)					
Thiamethoxam (I)					
Tribenuron-methyl (H)					
Triasulfuron (H)					

Potential leaching period extends beyond the current monitoring period. * 1)

Derived from application before May 1999 (see Kjær et al., 2002).

2) Degradation products are not monitored (see text).

Pesticide (or its degradation products) detected in water samples from groundwater monitoring screens in concentrations exceeding $0.1 \,\mu g/l$.

Pesticide (or its degradation products) detected in water samples from groundwater monitoring screens in concentrations not exceeding $0.1 \,\mu$ g/l.

Pesticide (or its degradation products) not detected in water samples from the groundwater monitoring screens.

Table 20. Number of samples from the groundwater monitoring screens in which the various pesticides and/or their degradation products were detected at each site with the maximum concentration (µg/l) in parentheses (see Appendix 5 for further details). Degradation products are indicated in italics. Pesticides applied in spring 2010 are not included.

	Tylstrup	Jyndevad	Silstrup	Estrup	Faardrup
Azoxystrobin	0	0	0	1(0.011)	0
-CyPM	0	0	28(0.1)	9(0.085)	0
Bentazone	0	0	29(0.44)	16(0.022)	11(0.6)
-AIBA	0	0	0	1(0.026)	0
Bifenox		2(0.05)	5(0.1)	0	
-Bifenox acid		0	13(3.1)	0	
-nitrofen		0	0	0	
Ethofumesate			5(0.038)	0	31(1.4)
-fluazifop- P^{2}	0	0	1(0.072)	0	6(0.17)
$-TFMP^{2}$			48(0.29)		
Glyphosate		0	4(0.031)	39(0.67)	3(0.017)
-AMPA		2(0.022)	15(0.08)	8(0.07)	2(0.029)
Metamitron			29(0.168)	0	24(0.63)
-metamitron-desamino		_	30(0.19)	0	48(1.3)
Metribuzin	1(0.014)	0			
-metribuzin-desamino-diketo	236(0.204)	20(1.831)			
-metribuzin-diketo	453(0.554)	26(1.372)			
Picolinafen		0		0	
-CL153815		0		0	
Pirimicarb	0	0	3(0.011)	1(0.015)	2(0.035)
-pirimicarb-desmethyl	0	0	0	0	3(0.042)
-pirimicarb-desmethyl-	0	0	0	0	2(0.076)
Propyzamide	0		9(0.14)		1(0.033)
-RH24580	0		0		0
-RH24644	0		2(0.032)		0
-RH24655	0	004/011	0		0
$-PPU^{(3)}$	2(0.045)	284(0.11)			
-PPU-desamnino"	0	71(0.028)	26(0.124)	1(0,000)	51(1,0)
lerbuthylazine	0	0	36(0.124)	1(0.022)	51(1.9)
-aesethyl-terbuthylazine		24(0.023)	161(0.143)	7(0.053)	66(0.94)
-aesisopropyl-atrazine	1(0.014) 1(0.026)		4(0.047)*	27(0.034)	50(0.04)
-2-hydroxy-desethyl-	1(0.026)		1(0.010)*		7(0.092)
-2-nyaroxy-terbutnylazin	U 1(0,011)	1/0 01 4)	0*	5(0, 12)	34(0.009)
	1(0.011)	1(0.014)			1(0.01)
Amidosulluron Bromovunil	0	0		0	0
Clomazone	0	0		0	0
$EMC65317_{(2)}$	0				0
Dimethoate	0	0	1(0.085)	0	0
$-FTI^{l}$	2(0,024)	0	1(0.005)	0	0
Enoviconazole	2(0.024)	1(0.011)			
Flampron-M-isopropyl	0	1(0.011)	1(0.024)	0	0
-flamprop (free acid)	Ő		0	ŏ	Ő
Fluroxypyr	Ő	0	Ő	1(0 058)	1(0.072)
Ioxvnil	ŏ	ŏ	0	0	1(0.01)
MCPA	Ũ	ŏ	0	1(0.019)	0
-4-chlor-2-methylphenol		ŏ	Ő	0	ŏ
Mesosulfuron-methyl		Õ	Ū.	Õ	Ū.
Pendimethalin	0	Õ	0	-	0
Phenmedipham			0		2(0.025)
-MHPC			0		1(0.053)
Propiconazole	0	0	0	2(0.022)	1(0.035)
Prosulfocarb			1(0.027)		0
Pyridate		0	0		
-PHCP		0	14(0.309)		
Triflusulfuron			0		
-IN-M7222			1(0.052)		
Desmedipham			1(0.033)		0
Fenpropimorph	0	1(0.029)	0	0	1(0.015)
-fenpropimorph-acid	0	0	0	0	0
Metsulfuron-methyl				0	
-triazinamin				1(0.042)	

-triazinamin ⁻¹Degradation product of mancozeb. ²Degradation product of fluazifop-P-butyl. ³Degradation product of rimsulfuron.

*Included in the monitoring at Silstrup from February 2003, eight months after application of terbuthylazine.

- Bentazone leached through the root zone (1 m b.g.s.) in average concentrations • exceeding 0.1 µg/l in the drainage system at the loamy sites of Silstrup, Estrup, and Faardrup. Moreover, bentazone was frequently detected in the monitoring screens situated beneath the drainage system at Silstrup and Faardrup (Table 19 and 20). Apart from eight samples, however, concentrations detected were all below 0.1 µg/l. At Estrup leaching was mostly confined to the depth of the drainage system and rarely detected in deeper monitoring screens (Appendix 5). On the sandy soils, bentazone leached at Jyndevad, but was only detected once 1 m b.g.s. at Tylstrup. At Jyndevad high concentrations (exceeding 0.1 μ g/l) were detected in the soil water samples from suction cups 1 m b.g.s. four months after application. Thereafter, leaching diminished and bentazone was not subsequently detected in the monitoring wells. Although leached in high average concentrations (>0.1 μ g/l) at four sites, bentazone was generally leached within a short period of time. Initial concentrations of bentazone were usually very high, but then decreased rapidly. In general, concentrations exceeding 0.1µg/l were only found within a period of one to four months following the application. The degradation product AIBA was detected twice in the vadose zone at Jyndevad, once in drainage water at Estrup and Faardrup (Table 18), and once in water from a horizontal well at Estrup (Table 20).
- Bifenox acid (degradation product of bifenox) leached through the root zone and entered the drainage water system in average concentrations exceeding 0.1 μ g/l at the loamy sites of both Silstrup and Estrup. While leaching at Estrup seems to be confined to the depth of the drainage system, leaching to groundwater monitoring wells situated beneath the drainage system was observed at Silstrup, where concentrations exceeding 0.1 μ g/l were observed up to six months after application.. Similar evidence of pronounced leaching was **not** observed on the coarse sandy soil as there was only a single detection of bifenox acid in soil water, whereas bifenox was detected very sporadically in soil and groundwater, concentrations always less than 0.1 μ g/l.
- In the loamy soil of Estrup, ethofumesate, metamitron, and its degradation product • metamitron-desamino leached through the root zone (1 m b.g.s.) into the drainage water in average concentrations exceeding 0.1 μ g/l (Table 17). The compounds have not been detected in deeper monitoring screens. These compounds also leached 1 m b.g.s. at the Silstrup and Faardrup sites, reaching both the drainage system (Table 17 and 18) and groundwater monitoring screens (Table 19 and 20). Average concentrations in drainage water were not as high as at Estrup, although concentrations exceeding 0.1 µg/l were observed in both drainage water and groundwater monitoring screens during a 1-6-month period at both Silstrup and Faardrup (see Kjær et al., 2002 and Kjær et al., 2004 for details). The above leaching was observed following an application of 345 g/ha of ethofumesate and 2,100 g/ha of metamitron in 2000 and 2003. Since then, ethofumesate has been regulated and the leaching risk related to the new admissible dose of 70 g/ha was evaluated with the two recent applications (2008 at Silstrup and 2009 at Faardrup). Although metamitron has not been regulated, a reduced dose of 1400 g/ha was applied at the two recent applications. The leaching following these recent applications (2008 at Silstrup and 2009 at Faardrup) with reduced dose of both ethofumesate and metamitron was minor. Apart from a few samples from the drainage system and groundwater monitoring wells containing less than 0.1 µg/l,

neither ethofumesate nor metamitron was found in any of the analysed water samples (see section 4.2.3 and 6.2.3).

- Fluazifop-P-butyl has several times been included in the monitoring programme at Jyndevad, Tylstrup, Silstrup, and Faardrup. As fluazifop-P-butyl rapidly degrades, until July 2008 monitoring has focused only on its degradation product fluazifop-P (free acid). Except for one detection below 0.1 μ g/l in groundwater at Silstrup and 17 detections with eight exceeding 0.1 μ g/l (four drains, three vadose zone, one groundwater, Table 18 and 20) at Faardrup, leaching was not evident. TFMP, the degradation product of fluazifop-P-butyl, was included in the monitoring programme at Silstrup in July 2008 following an application of fluazifop-P-butyl. After approximately one month, TFMP was detected in the groundwater monitoring wells, where concentrations at or above 0.1 μ g/l were detected within a ten-month period following application (Figure 22, Tables 19 and 20). At the onset of drainage flow in September, TFMP was detected in all the drainage water samples at concentrations exceeding 0.1 μ g/l (Figure 22). The leaching pattern of TFMP indicates pronounced preferential flow also in periods with a relatively dry vadose zone.
- Glyphosate and AMPA was found to leach through the root zone at high average • concentrations on loamy soils. At the loamy sites Silstrup and Estrup, glyphosate has been applied two (in 2001 and 2003) and four (in 2000, 2002, 2005, and 2007) times within the monitoring period, respectively. All six autumn applications have resulted in detectable leaching of glyphosate and AMPA from the upper meter into the drainage water, often at concentrations exceeding 0.1 µg/l several months after application. Higher leaching levels of glyphosate and AMPA have mainly been confined to the depth of the drainage system and have rarely been detected in monitoring screens located below the depth of the drainage systems, although it should be noted that detections of particularly glyphosate in groundwater monitoring wells at Estrup seem to increase over the years (Figure 33D). This increase may be underestimated for the period June 2007 to July 2010 as external quality assurance of analytical methods in this period indicates that the true concentration of glyphosate may be underestimated (see section 7.2.2). On two occations heavy rain events and snowmelt triggered leaching to the groundwater monitoring wells in concentrations exceeding 0.1 µg/l more than two year after the application (Figure 33D). Numbers of detections exceeding 0.1 μ g/l in groundwater monitoring wells are, however, very limited (only three samples). Glyphosate and AMPA were also detected in drainage water at the loamy site of Faardrup (as well as at the now discontinued Slaeggerup site), but in low concentrations (Kjær et al., 2004). Evidence of glyphosate leaching was only seen on loamy soils, whereas the leaching risk was negligible on the coarse sandy soil of Jyndevad. Here, infiltrating water passed through a matrix rich in aluminium and iron, thereby providing good conditions for sorption and degradation (see Kjær et al., 2005a for details).
- Two degradation products of metribuzin metribuzin-diketo and metribuzindesamino-diketo – leached 1 m b.g.s. at average concentrations exceeding $0.1 \,\mu g/l$ in the sandy soil at Tylstrup. Both degradation products appear to be relatively stable and leached for a long period of time. Average concentrations reaching $0.1 \,\mu g/l$ were seen as late as three years after application (Table 17). Evidence was also found that their degradation products might be present in the groundwater several

years after application, meaning that metribuzin and its degradation products have long-term sorption and dissipation characteristics (Rosenbom *et al.*, 2009). At both sandy sites (Tylstrup and Jyndevad), previous applications of metribuzin has caused marked groundwater contamination with its degradation products (Kjær *et al.*, 2005b).

- At Estrup, CL153815 (degradation product of picolinafen) leached through the root zone upper meter into the drainage water in average concentrations exceeding 0.1 µg/l (Appendix 5). CL153815 has not been detected in deeper monitoring screens (Table 20). Leaching of CL153815 have not been observed on the sandy soil at Jyndevad, (Table 17, Table 20, and Appendix 5).
- Pirimicarb together with its two degradation products pirimicarb-desmethyl and pirimicarb-desmethyl-formamido has been included in the monitoring programme for all five sites. All of the three compounds have been detected, but only pirimicarb-desmethyl-formamido leached through the root zone (1 m b.g.s.) entering the drainage system in average concentrations exceeding 0.1 μ g/l (Table 17) at Estrup. Comparable high levels of leaching of pirimicarb-desmethyl-formamido have not been observed with any of the previous applications of pirimicarb at the other PLAP sites (Table 17 and Kjær *et al.*, 2004). Both degradation products (pirimicarb-desmethyl and pirimicarb-desmethyl-formamido) have been detected in deeper monitoring screens at Faardrup (Table 19 and 20).
- Propyzamide leached through the root zone (1 m b.g.s.) at the loamy Silstrup and Faardrup sites, entering the drainage system at average concentrations exceeding 0.1 µg/l (Table 17 and 18). Propyzamide was also detected in the monitoring screens situated beneath the drainage system. Apart from a few samples at Silstrup, the concentrations in the groundwater from the screens were always less than 0.1 µg/l (Appendix 5, Table 19 and 20).
- One degradation product of rimsulfuron PPU leached from the root zone (1 m • b.g.s.) in average concentrations reaching 0.10-0.13 µg/l at the sandy soil site at Jyndevad. Minor leaching of PPU was also seen at the sandy site Tylstrup, where low concentrations (0.021-0.11 μ g/l) were detected in the soil water sampled 1 and 2 m b.g.s (Tables 17 and 18). In groundwater PPU was occasionally detected and twice exceeded 0.1 µg/l at Jyndevad, whereas it was only detected once (and at a low concentration) at Tylstrup (Table 19 and 20). At both sites, PPU was relatively stable and persisted in the soil water for several years, with relatively little further degradation into PPU-desamino. E.g. average leaching concentrations reaching 0.1 μ g/l were seen as much as three years after application at Jyndevad. With an overall transport time of about four years, PPU reached the downstream monitoring screens. Thus, the concentration of PPU-desamino was much lower and apart from four samples at Jyndevad, never exceeded 0.1 µg/l. It should be noted that the concentration of PPU is likely to be underestimated by up to 22-47%. Results from the field-spiked samples thus indicate that PPU is unstable and may have further degraded to PPU-desamino during analysis (Rosenbom et al., 2010a).
- Terbuthylazine as well as its degradation products leached through the root zone (1 m b.g.s.) at high average concentrations on loamy soils. At the three loamy soil sites Silstrup, Estrup, and Faardrup, desethyl-terbuthylazine leached from the upper meter

entering the drainage water in average concentrations exceeding 0.1 µg/l (Table 17 and 18). Four years after application at Estrup, both terbuthylazine and desethylterbuthylazine were detected in drainage water, but not exceeding 0.1 µg/l. At Silstrup (Kjær et al., 2007) and Faardrup (Kjær et al., 2009), desethyl-terbuthylazine was frequently detected in the monitoring screens situated beneath the drainage system (Table 19 and 20) at concentrations exceeding 0.1 µg/l during a 2- and 24month period, respectively. Leaching at Estrup (Kjær et al., 2007) was confined to the drainage depth, however. Minor leaching of desethyl-terbuthylazine was also seen at the two sandy sites Jyndevad and Tylstrup, where desethyl-terbuthylazine was detected in low concentrations ($<0.1 \mu g/l$) in the soil water sampled 1 m b.g.s. While desethyl-terbuthylazine was not detected in the groundwater monitoring screens at Tylstrup, it was frequently detected in low concentration ($< 0.1 \mu g/l$) at Jyndevad (Table 20, Kjær et al., 2004). Marked leaching of terbuthylazine was also seen at two of the three loamy sites (Estrup and Faardrup), the leaching pattern being similar to that of desethyl-terbuthylazine. 2-hydroxy-desethyl-terbuthylazine and 2hydroxy-terbuthylazine leached at both Faardrup and Estrup and at the latter site the average drainage concentration exceeded 0.1 µg/l. Leaching of these two degradation products was at both sites confined to the drainage system. None of the two degradation products were detected in groundwater monitoring screen at Estrup, whereas at Faardrup both were found, but at low frequencies of detection and concentrations.

• Tebuconazole has been applied in autumn 2007 at Tylstrup, Jyndevad, Estrup and Faardrup. Only on the loamy soil of Estrup did it leach through the root zone (1 m b.g.s.) and into the drainage water in average concentrations exceeding 0.1 μ g/l (Table 17 and 18). Leaching was mainly confined to the depth of the drainage system, although the snow melt occurring in March 2011 (more than two years after application) induced leaching of tebuconazole to the groundwater monitoring well in concentrations exceeding 0.1 μ g/l (Tables 19 and 20). None of the applications at the three other PLAP sites caused tebuconazole to be detected in similarly high concentrations in the vadose zone, though concentrations below 0.1 μ g/l have been detected in a few samples from the groundwater monitoring screens (Table 19 and 20).

The monitoring data also indicate leaching 1 m b.g.s. of a further 17 pesticides (or their degradation products), but often in low concentrations. Although the concentrations detected 1 m b.g.s. exceeded 0.1 μ g/l in several samples, the average leaching concentration (1 m b.g.s.) did not. This is summarized in Table 18, showing the number of samples in which the various pesticides were detected on each site as well as the maximum concentration. Apart from slight leaching of ETU (Kjær *et al.*, 2002) and amidosulfuron, within this group of 16 pesticides (or their degradation products) leaching from 1 meter was only observed at the loamy soil sites, where it was associated with pronounced macropore transport, resulting in very rapid movement of pesticides through the vadose zone. It should be noted that the findings regarding amidosulfuron are of very limited use since the degradation products – with which the leaching risk is probably mainly associated – are not included, as methods for their analysis are not yet available.

Eleven of the 42 pesticides applied – about 26% – did not leach at all from 1 m b.g.s. during the monitoring period (Table 17). Four of the 11 were, however, detected in the

groundwater monitoring screens (Table 19). The group of 11 includes the three different sulfonylureas – metsulfuron-methyl, triasulfuron, iodosulfuron-methyl-sodium and tribenuron-methyl applied at several sites. For example, tribenuron-methyl was applied at four different sites under different hydrological conditions, with percolation (1 m b.g.s.) during the first month after application ranging from 0 to 114 mm. The monitoring results give no indication of leaching for any of the compounds or their degradation products. It should, however, be noted, that the leaching risk associated with an autumn application of tribenuron-methyl, where preferential transport is likely to occur, has not yet been evaluated for the loamy soils.

The leaching patterns of the sandy and loamy sites are further illustrated in Figure 40 and 41, showing the frequency of detection in samples collected 1 m b.g.s. (suction cups on sandy soils and drainage systems on loamy soils) and the deeper located groundwater monitoring screens.



Figure 40. Frequency of detection in samples from the suction cups (left) and groundwater monitoring screens located deeper than the suction cups (right) at the sandy soil sites: **Tylstrup** (A, B) and **Jyndevad** (C, D). Frequency is estimated for the entire monitoring period and the length of time that the different pesticides have been included in the programme and the number of analysed samples thus varies considerably among the different pesticides. The figure only includes the fifteen most frequently detected pesticides. Pesticides monitored for less than two years are indicated by an asterisk and pesticides monitored for less than one year are not included.



Figure 41. Frequency of detection in samples from the drainage system (left) and groundwater monitoring screens located deeper than the drainage system (right) at the loamy soil sites: **Estrup** (A, B), **Silstrup** (C, D), and **Faardrup** (E, F). Frequency is estimated for the entire monitoring period and the time that the different pesticides have been included in the programme and the number of analysed samples thus varies considerably among the different pesticides. The figure only includes the 15 most frequently detected pesticides. Pesticides monitored for less than two years are indicated by an asterisk and pesticides monitored for less than one year are not included.

On the sandy soils the number of leached pesticides as well as the frequency of detection was much lower than on loamy soils (Figure 40 and 41), the exceptions being the mobile and persistent degradation products of rimsulfuron and metribuzin, frequently found in both suction cups and groundwater monitoring wells. This difference was mainly due to the different flow patterns characterising the two different soil types. On the sandy soils infiltrating water mainly passed through the matrix, thereby providing good conditions for sorption and degradation. Pesticides being leached in the sandy soils were thus restricted to mobile as well as persistent pesticides.

On the loamy soils pronounced macropore transport resulted in the pesticides moving very rapidly through the unsaturated zone. Compared to the sandy soils residence time was much lower on the structured, loamy soils. As a result of this, various types of pesticides, even those being strongly sorbed, were prone to leaching on the loamy types of soil.

At the loamy sites pronounced leaching was generally confined to the depth of the drainage system. Several pesticides were often detected in the drainage system, whereas the frequency of detection in the monitoring screens situated beneath the drainage system was lower and varied considerably between the three sites (Figure 41). These differences should be seen in relation to the different sampling procedures applied. Frequent, integrated water samples can be provided from a drainage system that continuously captures water infiltrating throughout the drainage runoff season. However, although the monitoring screens situated beneath the drainage systems were sampled less frequently (on a monthly basis from a limited number of the monitoring screens (Appendix 2), pesticides were frequently found in selected screens at Faardrup and Silstrup. Hitherto, at the Estrup site, leaching of pesticides has mainly been confined to the depth of the drainage system. Apart from 39, 27 and 16 samples containing glyphosate, desisopropyl-atrazine, and bentazone respectively, pesticides have only occasionally been detected in the screens beneath the drainage system (Appendix 5). The differences are, however, largely attributable to the hydrological conditions. Compared to the Silstrup and Faardrup sites, the C horizon (situated beneath the drainage depth) at the Estrup site is less permeable with less preferential flow through macropores (se Kjær et al. 2005c for details). The movement of water and solute may therefore be slower at Estrup, allowing for dispersion, dilution, sorption and degradation and thereby reducing the risk of transport to deeper soil layers. An indication thereof are the long periods with groundwater table beyond drainage depth in which an increasing lateral transport to the drainage system and decreased leaching to the deeper groundwater will occur.

Comparing the loamy sites, the number of drainage water samples containing pesticides/degradation products was markedly higher at Silstrup and Estrup than at Faardrup, which is largely attributable to the differences in the hydrological conditions. The occurrence of precipitation and subsequent percolation within the first month after application were, generally, higher at Silstrup and Estrup than at Faardrup (Table 9, Table 11, and Table 13).
9 References

Allerup, P., and Madsen, H. (1979): Accuracy of point precipitation measurements. Danish Meteorological Institute, *Climatological Papers No. 5*, Copenhagen, 84 pp.

Barlebo, H.C., Rosenbom, A., and Kjær, J. (2007): Evaluation of Pesticide Scenarios for the Registration Procedure; no. 1178; Danish Environmental Protection Agency. Available on http://www2.mst.dk/common/Udgivramme/Frame.asp? http://www2.mst.dk/Udgiv/publications/2007/978-87-7052-540-/html/default_eng.htm.

Boesten, J. J. T. I. (2000): From laboratory to field: uses and limitations of pesticide behaviour models for the soil/plant system. *Weed Res.*, 40, 123–138.

Danish EPA (1997): Bekendtgørelse nr. 637 af 30. juni 1997, Miljø- og Energiministeriet 1997.

EEC (1971): Directive 71/250/EEC of 15 June 1971 establishing Community methods of analysis for the official control of feeding-stuffs, *Official Journal L 155, 12/07/1971 pp. 0013–0037.* Information is also available on: http://www.seas.upenn.edu/courses/belab/LabProjects/1997/BE210S97R4R01.htm

Funk, W., Dammann, V., and Donnevert, G. (1995): Quality assurance in analytical chemistry, VCH Verlagsgesellschaft GmbH, Weinheim.

Flerchinger, G.N., Seyfried, M.S. and Hardegree, S.P. (2006): Using soil freezing characteristics to model multi-season soil water dynamics. Vadose Zone Journal, 5, 1143-1153.

Juhler, R.K., Henriksen, T., Rosenbom, A.E., and Kjaer, J. (2010): Fate and transport of chlormequat in subsurface environments. *Environ. Sci. Pollut. Res.*, 17(6), 1245-1256.

Kjær, J., Ullum, M., Olsen, P., Sjelborg, P., Helweg, A., Mogensen, B., Plauborg, F., Jørgensen, J. O., Iversen, B. O., Fomsgaard, I., and Lindhardt, B. (2002): The Danish Pesticide Leaching Assessment Programme: Monitoring results, May 1999–June 2001, Geological Survey of Denmark and Greenland, 2002.

Kjær, J., Ullum, M., Olsen, P., Sjelborg, P., Helweg, A., Mogensen, B., Plauborg, F., Grant, R., Fomsgaard, I., and Brüsch, W. (2003): The Danish Pesticide Leaching Assessment Programme: Monitoring results, May 1999–June 2002, Geological Survey of Denmark and Greenland, 2003.

Kjær, J., Olsen, P., Barlebo, H. C., Juhler, R. K., Plauborg, F., Grant, R., Gudmundsson, L., and Brüsch, W. (2004): The Danish Pesticide Leaching Assessment Programme: Monitoring results, May 1999–June 2003, Geological Survey of Denmark and Greenland, 2004.

Kjær, J., Olsen, P., Ullum, M., and Grant, R. (2005a): Leaching of glyphosate and amino-methylphosphonic acid from Danish agricultural field sites, *J. Environ. Qual.*, 34, 608–630.

Kjær, J., Olsen, P., Henriksen, T. and Ullum, M. (2005b): Leaching of metribuzin metabolites and the associated contamination of a sandy Danish aquifer, *Environ. Sci. Technol.* 39, 8374-8381.

Kjær, J., Olsen, P., Barlebo, H. C., Juhler, R. K., Henriksen, T., Plauborg, F., Grant, R., Nyegaard P., and Gudmundsson L. (2005c): The Danish Pesticide Leaching Assessment Programme: Monitoring results, May 1999–June 2004, Geological Survey of Denmark and Greenland, 2005.

Kjær, J., Olsen, P., Barlebo, H. C., Henriksen T., Plauborg, F., Grant, R., Nyegaard P. Gudmundsson, L., and Rosenbom, A. (2007): The Danish Pesticide Leaching Assessment Programme: Monitoring results, May 1999–June 2006, Geological Survey of Denmark and Greenland, 2007.

Kjær, J., Rosenbom, A., Olsen, P., Juhler, R.K., Plauborg, F., Grant, R., Nyegaard, P, Gudmundsson, L., and Brüsch, W. (2008): The Danish pesticide Leaching Assessment Programme: Monitoring results May 1999 - June 2007. Geological Survey of Denmark and Greenland. Copenhagen, 92 pp. + 6 Appendices.

Kjær, J., Rosenbom, A., Olsen, P., Ernstsen, V., Plauborg, F., Grant, R., Nyegaard, P, Gudmundsson, L., and Brüsch, W. (2009): The Danish pesticide Leaching Assessment Programme: Monitoring results May 1999 - June 2008. Geological Survey of Denmark and Greenland. Copenhagen, 85 pp. + 6 Appendices.

Kördel, von W. (1997): Feldversuche zum Austrag von Pflanzenschutzmitteln über Drainage – Abschätzung der Belastung aquatischer Ökosysteme, *Gesunde Pflanzen, 49* (5): 163–170 (in German).

Larsbo, M., Roulier, S., Stenemo, F., Kasteel, R., and Jarvis, N. (2005): An improved dual-permeability model of water flow and solute transport in the vadose zone. *Vadose Zone Journal* 4(2), 398-406.

Lindhardt, B., Abildtrup, C., Vosgerau, H., Olsen, P., Torp, S., Iversen, B. V., Jørgensen J. O., Plauborg, F., Rasmussen, P., and Gravesen, P. (2001): The Danish Pesticide Leaching Assessment Programme: Site characterization and monitoring design, Geological Survey of Denmark and Greenland, 2001.

Miller, J. N., and Miller, J. C. (2000): Statistics and chemometrics for analytical chemistry, Pearson, Essex.

Plantedirektoratet (2009): Vejledning om gødsknings- og harmoniregler. Planperioden 1. august 2008 til 31. juli 2009. Ministeriet for Fødevare, Landbrug og Fiskeri. Revideret oktober 2008. 105 pp. ISBN 978-87-7083-238-0. (Web) http://www.netpublikationer.dk/FVM/9114/pdf/Vejledning_GR_HR_2008-09_rev_udg.pdf. Plantedirektoratet (2010): Vejledning om gødsknings- og harmoniregler. Planperioden 1. august 2009 til 31. juli 2010. Ministeriet for Fødevarer, Landbrug og Fiskeri. Revideret udgave 14. September 2009. http://www.netpublikationer.dk/FVM/978-87-7083-418-6/index.htm.

Topp, G.C., Davis, J.L. and Annan, A.P. (1980). Electromagnetic determination of soil water content: Measurements in coaxial transmission lines. Water Resources Research, 16, 574-582.

Rosenbom, A.E., Kjær, J., Henriksen, T., Ullum, M., and Olsen, P. (2009): Ability of the MACRO Model to Predict Long-Term Leaching of Metribuzin and Diketometribuzin, *Environ. Sci. Technol.*, 43(9), 3221-3226.

Rosenbom, A.E., Kjær, J., and Olsen, P. (2010a): Long-term leaching of rimsulfuron degradation product through sandy agricultural soils, *Chemosphere*, 79, 830-838.

Rosenbom A., Brüsch, W. Juhler, R. K., Ernstsen, V., Gudmundsson, L., Plauborg, F., Grant, R. and Olsen, P., (2010b): The Danish Pesticide Leaching Assessment Programme: Monitoring results May 1999 - June 2009. Geological Survey of Denmark and Greenland. Copenhagen (available on www.pesticidvarsling.dk).

Thorling, L. (ed.) (2010): Grundvand. Status og udvikling 1989-2008. GEUS 2010. ISBN978-87-7871-274-5. http://www.geus.dk/publications/grundvandsovervaagning/index.htm.

U.S. Environmental Protection Agency (1998): Guidance for prospective groundwater monitoring studies. Environmental Fate and Effects Division, Office of Pesticide Programs, U.S. Environmental Protection Agency, September 16, 1998.

Wilson, A. L. (1970): The performance characteristics of analytical methods II, *Talanta* 17(1), 31–44.

Parameter Systematic chemical nomenclature Anidosulfuron N:IIII(1-6 dimethory-2-pyrimidin-1-yloxy]phenyl]-N- methylmethanesulfonamide Aroxystrobin Methyl (Tp-2) 21 (6-(2-yamophenoxy)-4-pyrimidin-4-yloxy]phenyl]-3- methoxyacrylate - CyPM E-24216 (c-2-yamophenoxy)-4-pyrimidin-4-yloxy]phenyl]-3- methoxyacrylate - CyPM E-24216 (c-2-yamophenoxy)-2-pyrimidin-4-yloxy]phenyl]-3- methoxyacrylate Bentazone 3 (1-methylethyl)-11-2,1,3-benzotialazin 4(3H)-one 2.2 dioxide AllAn 2-mino-N-isopropyl-barzizmid Bronoxyrill So-bichlorophyl-4-mirophyl-1-3-isovazolidone Nitrofm 2-4-dichlorophenoyl-1-sinvazolidone Choravacia 2-chlorophenolyl-mirophyl-1-sinvazolidone Comzone 2-(Ca-thorpheno)Imethyl]-3-isovazolidone Comzone 2-(Ca-thorpheno)Imethyl]-1-3-isovazolidone Comzone 2-(Ca-thorpheno)Imethyl]-1-siovazolidone Comzone 3-6 Dichloropyridne-2-carboxylic acid Desmedipham Ethyl 3-cphenylca-anoylnextyl-phosphorodithioate Fluaziop-1 ²⁺¹ S-trifluoromethyl-pyridne-2-adimethyloxyphenyl-ethyl-phosphorodithioate Fluaziop-1 ²⁺¹ C-2(C-athoxypropyl)-2.3-epncy-2-(4-fluorophenyl)-2.6-imethylnorpholine Flampto-M-isopropyl C	2	~
Amidosulfaron N:[[][(4.6-dimethory-2-pyrimidimy]Damino[carbony]1-amino[bulfomy]1-N-methy]methanesulfonamide Azoxystrobin Methyl (E) -2[-2] (6 (-2-yanophenoxy) - pyrimidim -4-yloxy]-phenyl] - 3-methoxyacrylic acid -CyPM E-2-(2-16-(2-yanophenoxy)-pyrimidim -4-yloxy]-phenyl] - 3-methoxyacrylic acid Bentazone 3-(1-methylethyl) - 11-2.1-5-benzohidaitadia: (3H) one 2.2 dioxide -AIM 2-amino.N-isopropyl-benzamid Bronox ynil 3-dibtorophenoxy)-2-nitrobenzoic acid -AIMA 2-amino.N-isopropyl-benzamid Bronox acid 5-(2,4-dichtorophenoxy)-2-nitrobenzoic acid -Nitrofen 2-4/dichtorophenoxy)-2-nitrobenzoic acid -Nitrofen 2-dichtorophenyl - 1-isopphenyl-ether Chlormequat 2-chlorobylytimethyl[methyl]-14-dimethyl-3-isoxazolidione -EMCOS 177 (Propanamide (N-12-chlorophenyl)methyl-phenyl-achamate -Gongradid 36-Dichtorophyridine-2-carboxylic acid Desmedipham Ebhyl 3-(phenylcarbanonylnextyl-prindinyl)oxylphenoxy-propunoic acid -FIMC 517 (Propanamide (-2-d-4iox/synpus)final-phonyloxylic acid Desmedipham Ebhyl 3-(chenylcarbanonylin-23-dimethylBrenotruns-5, 1-methoylarophonyl-2-dimethyl phonyl)-1-methyl 2-methylpropyl-12.6-imethylmorpholine FIMC 517 (Propanamide (-2-d-4iox/synpus)-phenyl)-2-1-methylpropyl-2.6-imethylmorphol	Parameter	Systematic chemical nomenclature
methylmethanesulfonanide Axoxystrbin Methyl (C)-212-(6-C-2xophenoxy)-4-pyrimidin-4-ytoxylphenyl)-3-methoxyacrylac - CyPM E2-42-16-(e-xymophenoxy)-pyrimidin-4-ytoxylphenyl)-3-methoxyacrylic acid Bentazone 3-(1-methylethyl)-HE-21,3-benzohiadiazin-4(3H)-one 2.2 dioxide ABA 2-amino. Nisoporph-Benzamid Bronoxynil 3.5 dibrono-4-hydroxybenzonicile Brienox methyl 5-(2-4-dichorphenoxy)-2-nitrobenzoate Bronox acid 5-(2-4-dichorphenyl-nitrobenzoate - Bilenox acid 5-(2-4-dichorphenyl-nitrobenzoate - FileGS171 (Propanamide 2-(2-chorbenyl-nitrobenzoate (Chorophenyl-Anthylophenyl)-ethylpenyl-1-3-systroxy-2.2-dimethylophenyl-ethylpenyl 2-(2-chorophylpenyl-mylpenyl-ethyl-anamet (Copyrald 3.6-Dichloropyridine-2-carboxylic acid Desmethylacthanate (Copyrald 3.6-Dichloropyridine-2-carboxylic acid Desmethylacthanate (FIRCS)17 (Propanamide (C)-dimethyl 5-methylacthanate Desmethylacthanate (Copyrald 3.6-Dichloropyridine-2-carboxylic acid Desmethylacthanate (Darbating) 3.6-Dichloropyridine-2-carboxylic acid Desmethylacthanate (Darbating) 3.6-Dichlorophethyl)-sphenyl-12-site Dimathy	Amidosulfuron	N-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]-amino]sulfonyl]-N-
Azoxystrobin Methyl (B) 2-[2](6-(2-cynophenoxy)-pyrimidin-4-yloxy]-phenyl) - 3-methoxyacrylia caid - CyPM E>2-(2)-6-cynophenoxy)-pyrimidin-4-yloxy]-phenyl) - 3-methoxyacrylic acid - AlBA 2-amino-N-isopropyl-benzanid Bromzone 3-(1-methylethyl)-HE-12-3-benzohiaduzin-4(3H)-one 2.2 dioxide - AlBA 2-amino-N-isopropyl-benzanid Bromzone 3-(2-adchlorophenoxy)-2-mirobenzoate - Bifenox acid 5-(2-4-dichlorophenoxy)-2-mirobenzoate - Bifenox acid 5-(2-4-dichlorophenoxy)-2-mirobenzoate - Bifenox acid 5-(2-dichlorophenoxy)-2-mirobenzoate - Choroethyltrimethylammonium chloride Choroethyltrimethylammonium chloride Clomazone -2(2-chlorophenyl)-mtyl)-4-4-dimethyl-3-isozazolidione - FMCC5517 (Propanaide (D-2-chloroyphenyl)-ehyl ster Desnediphan Ehbyl 5-methylcarbamosylnethyl-posphorodinioate - EHDC Carbamic acid, (3-hydroxyphenyl)-2-grinityl)(xylphenoxy-yropanoic acid - FMD ²⁺² S-rifluoromethyl-pyrtyl-phyl-2-grinityl)(xylphenoxy-yropanoic acid - FMD ²⁺⁷ S-rifluoromethyl-pyrtyl-phyl-2-sethylpropyl)-2.6-imethylmorpholine - Fauszign-P ²⁺² S-rifluorophenylb-2-sethylp-1-3-methylpropyl)-2.6-imethylmorpholine - Fauszign-P ²⁺² S-rifluorophenylb-2-sethylp-1-2-methylpropyl-2		methylmethanesulfonamide
$ \begin{array}{ll} \label{eq:product} \end{tabular} \e$	Azoxystrohin	Methyl (E)-2-{2-[(6-(2-cyanophenoxy)-4-pyrimidin-4-yloxy]phenyl}-3-
$-CyPM$ $\mathbb{P}_{2}^{2} < \mathbb{P}_{2}^{2} < < \mathbb{P}_{2}^{2} < \mathbb{P}_{2}^{2} < \mathbb{P}_{2}^{2} < < \mathbb{P}$		methowacrulate
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	C DM	$E_{0}(2) \left($
Bentazone 3-(1-methylethy)-1H-2.1,3-benzanid AIBA 2-mino.N-isopropyl-benzanid Bromoxynil 3,5-dibromo-4-hydroxybenzonitrile Bifenox methyl 5-(2,4-dichlorophenoxy)-2-nitrobenzoate - Bifenox 2-dichlorophenoxyn-2-nitrobenzoate - Nitrofen 2-dichlorophenoxyn-2-nitrobenzoate - Nitrofen 2-dichlorophenoxyn-2-nitrobenzoate - Nitrofen 2-dichlorophenyl-Nitrobyl-A-dimethyl-3-isoxazolioine - FMC65317 (Propanamide- Chorochethyltrimethylammonium chloride Clopyrald 3-6-Dichloropyridine-2-carboxylic acid Desmedipham Ehtyl 3-(phenylcarbanoyloxyphenyl-carbanate - EHPC Carbanic acid, (3-hydroxyphenyl)-thyl ester Dimethoate C)-2-dimethyl 5-xypridinyloxylphenosolruna-syl-methanesulfonate - Fluazifop-P ³⁺ (R)-2-(4-((5-(trifluoromethyl)-phenyl)-2-aphyrodithioate Cib-4-13-44-(1,2-dimethylethyl)-phenyl)-2-amethylpropyl]-2.6-imethylmorpholine Fempropinorph Cis-4-13-44-(1,2-dimethylethyl)-phenyl)-2-methylpropyl]-2.6-imethylmorpholine Flamprop-M-isopropyl N-benzoyl-N-3-chitoro-4-flourophenyl)-D-alaninate Prosulan 2, 6-3 kriftuoro 5-methyl-1, 3-5-tirazine Flamprop-M-isopropyl N-c(2, 6-difluorophenyl)-14-(1, 4-dimethystyl)-D-alaninate </td <td>- CyPM</td> <td>E-2-(2-[6-cyanophenoxy)-pyrimidin-4-yloxy]-phenyl) – 3-methoxyacrylic acid</td>	- CyPM	E-2-(2-[6-cyanophenoxy)-pyrimidin-4-yloxy]-phenyl) – 3-methoxyacrylic acid
- AIBA 2-amino-N-isopropyl-benzanirile Bromoxynil 3.5-dibrohenzany2-2-nitrobenzoate Bifenox acid 5:(2.4-dichlorophenoxy)-2-nitrobenzoate Bifenox acid 5:(2.4-dichlorophenoxy)-2-nitrobenzoate Othornequat 2-chlorobyl/inelyl-a-dichlorophenoxy)-2.nitrobenzoate Chornequat 2-chlorobyl/inelyl-4-4-dimethyl-3-isoxazolidione Clopyznild 3-6-Dichloropyridine-2-carboxylic acid Desmedipham Ethyl 3-chlorobyl/acarbanoxylnexyl-phosphorodithioate Clopyznild 3-6-Dichloropyridine-2-carboxylic acid Desmedipham Ethyl 3-chlorobyl-acbanomyloxylphenyl-phosphorodithioate Ethoromesate (+)-2-ethoxy-2,3-dihydro-3.3-dimethyl-benzylphenzyl-propanosic acid FIME Curbanic acid, (3-hydroxyphenyl)-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4+Triazol Cix-4-(13-4-(1,-2-ethlorophenyl))-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4+Triazol Cix-4-(13-4-(1,-2-ethlorophenyl))-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4+Triazol Cix-4-(13-4-(1,-2-ethlorophenyl))-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4+Triazol Cix-4-(13-4-(1,-2-ethlorophenyl))-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4+Triazol Cix-4-(13-4-(1,-2-ethlorophenyl))-2.3-epoxy-2-(4-fluorophenyl)propyl)-2.6- Ferraresulam-desmethyl	Bentazone	3-(1-methylethyl)-1H-2,1,3-benzothiadiazin-4(3H)-one 2,2 dioxide
Bromoxynil 3.5-dibromo-f-hydroxyberzonitrile Bifenox methyl 5-(2.4-dichtorophenoxy)-2-nitrobenzoate - Bifenox acid 5-(2.4-dichtorophenoxy)-2-nitrobenzoate - Bifenox acid 5-(2.4-dichtorophenoxy)-2-nitrobenzoate - Chlornequat 2-ehloroethyltrimethylammonium chloride Chornequat 2-ehloroethyltrimethyl-3-hydroxy-2.2-dimethyl propanamide Copyralid 3-6-Dichtoropyroline-2-carboxylic acid Desmedipham Ehl/PC Carbanic acid, (3-hydroxypheny)-exhlop lestr Dimethoate O,O-dimethyl S-methylcarbannoylmethyl-phosphorodithioate Ehl/PC Carbanic acid, (3-hydroxyphenyl)-exhlop lestr Dimethoate (h)-2-(4-(15-(rifluoromethyl)-p-2-pyridinyl)oxyphenoxy-propanoic acid - Filmazi[ga-P ⁻¹⁾ (R)-2-(4-(15-(rifluoromethyl)-phenzyl-2-a-methylpropyl)-2,6-imethylmorpholine Exproprimorph Cis 4-13-14-(2-carboxypropyl)-phenyl)-2-methylpropyl)-2,6-imethylmorpholine Fampropinnorphic acid N-benzyl-N-(3-chloro-4-flourophenyl)-D-alaninate Florasulam-desmethyl N-c2,6-difluorophenyl-N-2-shydroxyl1.2,4-litiazolo (1,5-c)pyrimidine-2-sulfonanilide Florasulam-desmethyl N-c2,6-difluorophenyl-N-3-shydroxyl1.2,4-litiazolo (1,5-c)pyrimidine-2-sulfonanilide Francopinnorphic aci	- AIBA	2-amino-N-isopropyl-benzamid
Biffonx methyl 5-(2,4-dichlorophenoxy)-2-nitrobenzoate Biffonx acid 5-(2,4-dichlorophenoxy)-2-nitrobenzoate Biffonx acid 2,4-dichlorophenoxy)-2-nitrobenzoate Chornequat 2,-dichlorophenyl +-nitrophenyl ether Clomazone 2,-dichlorophenyl +-nitrophenyl ether Comazone 2,-dichlorophenyl +-nitrophenyl ether Clopyralid 3,6-Dichloropyridine-2-carboxylic acid Desmedipham Ethyl 3-(phenylcarbanov)loxy)phenyl-phosphorodithioate Elboritonesate (D)-2-dindy-3,3-dinkyto-3,3-dinethyl-phosphorodithioate Elboritonesate (D)-2-dindy-3,2-dinkyto-3,3-dinethyl-phosphorodithioate Elboritonesate (E)-2-ethoxy-2,3-dinkyto-3,3-dinethyl-phosphorodithioate Elboritonesate (E)-2-ethoxy-2,3-dinkyto-3,3-dinethyl-phosphorodithioate Elboritonazole (ZRS, 33R)-1-(2,-2-chlorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4-triazol 1,2,4-triazol (Z-d-chlorophenyl)-2,-binylpropyl]-2,6-imethylmorpholine Fenpropinorphic Cis-4-(3,1-4(-1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine Fenpropinorphic acid Cis-4-(3,1-4(-1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine Fenpropinorphic acid Cis-4-(3,1-4(-1,1-dimethylethyl)-2-methylpropyl]-2,6-imethylfino	Bromoxvnil	3.5-dibromo-4-hydroxybenzonitrile
Bifencx acid $5(2,4-dichlorophenoxy)-2-nitrobenzoic acidNitrofen2,4-dichlorophenyl -4-nitrophenyl etherChlormequat2-dichlorophyllrimethyl ammonium chlorideClonzacone2(-2chlorphenyl)methyl] -3-kyarazolidioneClonzingen(N=2-chlorophyllrimethyl phyl] -3-kyarazolidioneClonzingen3.6-Dichloropytidine-2-carboxylic acidClonzingen3.6-Dichloropytidine-2-carboxylic acidDimethoate0, 0-dimethyl 3-methylcarbanateCharbine acid, (3-hydroxylnenyl-chlyl esterDimethoate(D, -2-chtoxy, -2, -4)(-6, -2-chtoxy, -propanoic acidFluzzifop-P21(N=2-chtoxy, -2, -4)(-6, -2-chtoxy, -propanoic acidFluzzifop-P21(N=2-chtoxy, -2, -4)(-6, -2-chtoxy, -propanoic acidFluzzifop-P21(N=2-chtoxy, -2, -4)(-6, -2-chtoxy, -propanoic acidFruzifop-P21(N=2-chtoxy, -2)(-4)(-6, -2-chtoxy, -propanoic acidFentropinnorphic acid(N=2+chtoxy, -2)(-4)(-6, -2-chtoxy, -2)(-4)(-6, -2-chtoxy, -2)(-4)(-6, -2)(-6)(-6)(-6)(-6)(-6)(-6)(-6)(-6)(-6)(-6$	Bifenox	methyl 5-(2 4-dichlorophenoxy)-2-nitrobenzoate
- Dynometal data 2-4-dischorphenyl 4-nitrophoenyl data - Nitrofen 2-4-dischorphenyl 4-nitrophenyl data Chormequat 2-chlorophenyl 4-nitrophenyl data Chormequat 2-(12-chlorophenyl)-4-dimethyl-3-isoxazolidione Clopzynid 3-(12-chlorophenyl)-4-dimethyl-3-isoxazolidione Clopzynid 3-(12-chlorophenyl)-4-dimethyl-3-isoxazolidione Clopzynid 3-(12-chlorophenyl)-4-dimethyl-3-isoxazolidione Clopzynid 3-(12-chlorophenyl)-4-signamate Desmediphan Ethyl 3-(phenylcarbanoyloxy)phenyl-arbanate - Fluzzifop-P ²) (R)-2-(4-((3-cfliftoromethyl)-2-pyridinyl)oxy)phenoxy-propanoic acid TFTMP ²) Striftoromethyl-pyridin-2-di Expositonazole (28, 358)-1-(2-(2-cl)-chlorophenyl)-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 12,4 +tizazol 1-24-4tizazol - Fluzzifop-P ²) Ethylenethiourea Fenzopinomphic acid Cis4-13+(4-(1, -dimethylethyl)-phenyl]-2-methylpropyl]-2.6-imethylImorpholine Isopropyl N-benzyl-N-(3-chloro-4-flourophenyl)-D-alaninate N-(2-6-difluorophenyl)-N-3-laninate Florasulam 2.6-3/chloro-6-fluoro-2-pyridinyl)oxylacetic acid Florasulam-desmethyl N-(2-6-difluorophenyl)-1.3,5-triazine-2-sulfonaniide	Diference and	5 (2.4 diablorophonovy) 2 nitrobonzoia acid
 Altrofen 2.4-dichlorophenyl 4-nitropienyl ether Chlormequat Chlormequat Chlorotenyl miterhyl 4-dimethyl 5-isosazolidione 2.[2chlorophenyl)methyl] 4.4-dimethyl-3-isosazolidione 2.[2chlorophenyl)methyl] 4.4-dimethyl-3-isosazolidione Clopyralid 3.6-Dichloropyridime-2-carboxylic acid Desmedipham Ethyl 3 (chenylcarbamoyloxy)phenylcarbamate Carbanic acid, (3-hydroxy)phenyl-ethyl ester Dimethoate C.Odimethyl 5-methylcarbamoylimethyl-phosphorodithioate Chuz(por, P⁻¹) C.Jdetoxy. 2.3-dihydro-3.3-dimethylcenbamate Fluzz(por, P⁻¹) S.Trifluoromethyl.'P.yridin-2.ol Epoxiconazole Cis-4-13-14-(1.1-dimethylethyl-phenyl)-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H-1,2.4-triazol Ethyl enchioarea Fenpropimorph Cis-4-13-14-(1.1-dimethylethyl-phenyl)-2.3-epoxy-2.6-(a-fluorophenyl)propyl)-2.6-intethylmorpholine Fenpropimorph Cis-4-13-14-(1.1-dimethylethyl-phenyl)-2.3-epoxy-2.6-(a-fluorophenyl)propyl)-2.6-intethylmorpholine Fenpropimorph Cis-4-13-14-(1.1-dimethylethyl)-phenyl)-2-methylpropyl]-2.6-intethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate Florasulam-desmethyl N-benzoyl-N-(3-chloro-6-fluro-2-pyridinyl)oxylactic acid Glyphosate N-(phosphonomethyl2glycine AMPA Amino-3.5-dichloro-6-fluro-2-pyridinyl)oxylactic acid Glyphosate N-(phosphonomethyl2glycine Addichlorophyll-1.3-striazine-2-yl)amino]=carbonyl]amino]-sulfonyl]henzoia Jacianamic Addichlorophyll-1.4-dicarboxyr-6-methyl-1.3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]henzoia Addichlorophyll-1.2	- Dijenox ucia	$3-(2,4-\alpha)$
Chlormequat 2-chlorotehyllrak-dimethyl-3-isozazolidione 2-l(2-chlorophyn)methyll-4-dimethyl-3-isozazolidione 2-l(2-chlorophyn)methyll-3-dydroxy-2,2-dimethyl propanamide clonacome) 3.6-Dichloropyrdine-2-carboxylic acid 2-byralid 3.6-Dichloropyrdine-2-carboxylic acid 2-byralid 2-chlorotehyll-athyll-athyll-athyll-3-dydroxy-2,2-dimethyl propanamide clonacome) 3.6-Dichloropyrdine-2-carboxylic acid 2-byralid 2-chlorotehyll-athyll-a	- Nitrofen	2,4-dichlorophenyl 4'-nitrophenyl ether
Clomazone 2-[(2-chlorpheny])methy]I-4/-dimethyl-3-isozazolidione - FMC65317 (Propanamide- clomazone) (N-[2-chlorpheno])methyl] -3-hydroxy-2.2- dimethyl propanamide Clopyralid 3.6-Dichloropyrdime-2-carboxylic acid Desmedipham Ethyl 3-(phenylcarbamoyloxy)phenyl-ethyl ester Dimethoate (D-0-dimethyl 5-methylcarbamoylowthyl-phosphorodithioate Ethoftmesate (P)-2-ethoxy-2.3-dihydro-3.3-dimethylbenzofuran-5-yl-methansulfonate - Fluactjop-P ⁻² (R):2-4(-(5.ctrifluoromethyl)-2-pyrdinyl)/2.3-epoxy-2-(4-fluorophenyl)popyl)-1H- 1.2,4+triazol - ETU ⁻¹ Ethylenethiourea Fenpropimorph Cis-4-[3-4(-1]dimethylethyl)-phenyl]-2-methylpropyl]-2.6- dimethylmorpholine Fenpropimorph Cis-4-[3-4(-2.carboxypropyl)-phenyl]-2-methylpropyl]-2.6- dimethylmorpholine FlampropM-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Florasulam-desmethyl 2.6, %: Trifluoro-5-methoxy-s-triazolo [1.5-c]pyrinidine-2-sulfonanitide - Florasulam-desmethyl N-(2.6-difluorophenyl)-N-(1.1, 2.4)triazolo[1.5-c]pyrinidine-2-sulfonanitide - Florasulam-desmethyl N-(2.6-difluorophenyl-1.3, 5-triazine - MIPA Amino-methylphosphonic acid Idyphosate N-(2.6-difluorophenyl-1.3, 5-triazine - Metsulfuron-methyl-sodium sodium sait of meth	Chlormequat	2-chloroethyltrimethylammonium chloride
- FMC65317 (Propanamide clomacone) (N-[2- chlorophenol)methyl] -3-hydroxy-2,2- dimethyl propanamide clomacone) Clopyralid 3,6-Dichloropyridine-2-carboxylic acid Desmedipham Ehtyl 3-(phenylcarbamoylocylphenylcarbamate - EHPC Carbanic acid, (3-hydroxyhenyl-phenyl ehtyl ester Dimethotate (D)-dimethyl S-methylcarbamoylmethyl-phosphorodithioate Ethofumesate (E)-2-(4+((5-(trifluoromethyl)-2)-syridinyl)oxylphenoxy-propanoic acid - FHm2 ³¹ S-trifluoromethyl-pyridin-2-ol Epoxiconazole Ciss.4-[3-[4-(1]-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-inethylmorpholine - Forpopinorph Ciss.4-[3-[4-(1]-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-inethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Flamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Flamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Florasulam-desmethyl N-(5, c-fifuorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonamide Glyphosate N-(6, c-fifuorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonamide - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl-sodium Iodosulfuron-methyl-sodium sodium salt of methyl + 1-(3,5-triazine-2-y)amino]=carbonyl]amio]-sulfony	Clomazone	2-[(2-chlorphenyl)methyl]-4,4-dimethyl-3-isoxazolidione
clonecome) 3.6-Dichloropyridine-2-carboxylic acid Desmedipham Ethyl 3-(phenylcarbamoyloxy)phenylcarbamate - EHPC Carbanic acid, (3-bydroxyphenyl)-ethyl ester Dimethoate (D,O-dimethyl 5-methylcarbamoylmethyl-phosphorodithioate Ethofumesate (E)-2-ethoxy-3.3-dimydro-3.3-dimethylbenzofuran-5-yl-methasulfonate - Fluacifyop-P ⁻² (R)-2-(4-((S-trifluoromethyl)-2yridinyl)oxy)phenoxy-propanoic acid - TFMP ⁻² S-trifluoromethyl-pyridin-2-of Epoxiconazole (2RS, SR)-1-(2-(2-carboxypropyl)-phenyl)-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1.2,4-triazol - ETU ⁻¹ Ethylenethiourea Fenpropimorph Cis-4-[3-[4-(4]-(1-dimethylenyl)-phenyl]-2-methylpropyl]-2.6- dimethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Plorasulan 2-6, Septimizazolo [1.5-c]pyrimidine-2-sulfonanilide - Florasulan-desmethyl N-(2.6-difluorophenyl)-8-fluoro-5-hydroxy11.2,4ltriazolo[1.5-c]pyrimidine-2- sulfonamide - Plorasulan-desmethyl N-(2.6-difluorophenyl)-1-methyl-phyloylacetic acid Glyphosate N-(phosphonomethyl)[pyrimidin-2-ylumino]-acarbonyl]amino]sulfonyl]benzote - Triazinamine 2-amino-4-methoxy-6-methyl-1,3.5-triazine - Metrolylinon-0. S-d	- FMC65317 (Propanamide-	(N-[2- chlorophenol)methyl] -3-hydroxy-2,2- dimethyl propanamide
Clopyrald 3,6-Dichloropyridine-2-carboxylic acid Desmedipham Ethyl 3-(phenylcarbamoyloxy)phenylcarbamate - EHPC Carbamic acid (3-hydroxy)henyl)-ethyl ester Dimethoate (D,O-dimethyl S-methylcarbamoylmethyl-phosphorodithioate Ethofumesate (E)-2-ethoxy-2.3-dihydro-3.3-dimethylbenzofurma-5-yl-methanesulfonate - Fluar [7] S-trifturonmethyl-2-pyridinyloxy)phenoxy-propanoic acid - TFUP 1 Explored (C)-2-c-thorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H- 12,4-triazol Explored (C)-2-c-thorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H- 12,4-triazol Explored (C)-2-c-thorophenyl)-2-methylpropyl]-2,6-imethylmorpholine Fenpropimorphic acid Cis-4-[3-[4-(1, 1, -dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Florasulam 2', 6', 8-Trifluoro-5-methoxy-s-triazolo [1, 5-c]pyrimidine-2-sulfonanilide Florasulam-desmethyl Nc(2, 6-difluorophenyl)-B-fluo-2-lyridinyl)oxylacetic acid Glyphosate N. (phosphonomethyl)glycine - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl Methyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-y])amino]=carbonyl]amino]-sulfonyl]benzoate - Metsulfuron-methyl Methyl2-[[[(4-methoxy-6-methyl-1,3,5-triazi	clomazone)	
Copyman 2),0-Dramopyriane 2),0-Dramopyriane 2),0-Dramate - EHPC Carbamic acid, (3-hydroxyphenyl)-ethyl ester Dimethoate C),O-dimethyl 3-emethylcarbamoyloxy/phenyloexphenylonexphenyloxyphenyl-propanoic acid - Fluazifgor-P ² / (R)-2-(4-((5-(trifluoromethyl)-2-pyridinyl)oxy)phenoxy-propanoic acid - TTMP ² / 5-trifluoromethyl-pyridin-2-0 Epoxiconazole (2RS, 3SR)-1-(2-(2-chlorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H-1,2,4-triazol - ETU ^{1/1} Ethylenethiourea Fenpropimorphic acid Cis-4-[3-[4-(2-carboxypropyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine - Fenpropimorphic acid Cis-4-[3-[4-(2-carboxypropyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine - Fenpropimorphic acid Cis-4-[3-[4-(2-carboxypropyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate Florasulam 2, 6: 3: Frifluoro-5-methoysstrizolo [1, 5-clpyrimidine-2-sulfonanilide Florasulam-desmethyl N-(2, 6-difluorophenyl)-8-fluro-2-pyridinyl)oxylacetic acid Glyphosate N-(phosphonomethyl)glycine - AMPA Amino-3-fichhoro-6-fluro-2-pyridinyl)oxylacetic acid Iodosulfuron-methyl sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]-carbonyl]amino]sulfonyl]benzoia	Clopyralid	3.6 Dichloropyriding 2 carboxylic acid
Desincipation Ethyl 3-(pneny(carbanoy(0xy)pneny(carbanate EHPC Carbanic acid (3 hydroxy)heny)(-ehyl ester Dimethoate O,O-dimethyl S-methylcarbanoylmethyl-phosphorodirhioate Ethofumesate (±)-2-ethoxy-2.3-dihydro-3.3-dimethylbenzofuran-5-yl-methanesulfonate Plauzifyp-P ²⁾ (R)-2-(4-((5-(rifluoromethyl))-2-pyridinyl)oxy)phenoxy-propanoic acid Strifluoromethyl-pyridin-2-ol Epoxiconazole Epoxiconazole (2RS, 3SR)-1-(2.2)-cehtorophenyl)-2.3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1.2,4-triazol ErTU ¹⁰ Ethylenethiourea Fenpropimorph Cis-4-[3-[4-(2,-carboxyproyp))-phenyl]-2-methylpropyl]-2,6- dimethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Florasulan 2, 6, 8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide Florasulan-desmethyl N-benzoyl-N-(3-chloro-6-fluro-2-pyridinyl)oxylacetic acid Glyphosate N-(phosphonomethyl-lylmino)sulfonyl]benzoate - AMPA Amino-methylphosphonic acid Idovsulfuron-methyl-sodium Sodi-dobe-2-methyl-1.3,5-triazine-2-ylamino]=carbonyl]amino]-sulfonylbenzoate - Triacimamine 2-amino-4-methoxy-6-methyl-1.3,5-triazine-2-ylamino]=carbonyl]amino]-sulfonylbenzoate - Advita Adino-A-sdihydo-3-methyl-6-phenyl-1.2,4-triazine-5-one		5,0-Dicinolopying ine-2-carboxyine actu
- EHPC Carbanic acid, (3-hydroxyphenyl)-ethyl ester Dimethoate Q.O-dimethyl S-methylcarbanoynlmethyl-phosphorodithioate Ethofumesate (1)-2-ethoxy-2,3-dihydro-3,3-dimethylbenzofuran-5-yl-methanesulfonate - Fluazifgor-P ²) (R)-2 (4-((5-(trifluoromethyl))-2-pyridinyl)oxy)phenoxy-propanoic acid TFMP ³) 5-trifluoromethyl-pyridin-2-3 Epoxiconazole (2RS, 3SR)-1-(2-(2-chlorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H-1,2,4-triazol - ETU ^{1/1} Ethylenethiourea Fenpropimorphic acid Cis-4-1314-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine - Fenpropimorphic acid Cis-4-1314-(2-carboxypropyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine - Fenpropimorphic acid Strifluoro-5-methoxy-6-trizolo [1,5-clpyrimidine-2-sulfonanilide Florasulam 2, 6: Strifluoro-5-methoxy-6-trizolo [1,5-clpyrimidine-2-sulfonanilide Florasulam-desmethyl N-(2,6-difluorophenyl)-8-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethyl)glycine - AMPA Amino-3-5-dichloro-6-fluro-2-pyridinyl)oxylacetic acid Odosulfuron-methyl-sodium sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2-y]/painino]-carbonyl]amino]-sulfonyl[benzoia acid Oxynil 4-hydroxy-3,5-diiodobenzonitile Limoro 3-(3,4-dichloroph	Desmedipham	Etnyl 3-(pnenylcarbamoyloxy)pnenylcarbamate
Dimethoate O.O-dimethyl S-methylcarbamoylmethyl-phosphorodithoate Ethofumesate (±)-2-ethoxy-5.3-dividydo-3.	- EHPC	Carbamic acid, (3-hydroxyphenyl)-ethyl ester
Ethofumesate (±)-2-cthoxy-2,3-dihydro-3,3-dimethylbenzofuran-5-yl-methanesulfonate - Fluazifop-P ⁻¹ Strifluoromethyl)-2-yridinyl)oxy)phenoxy-propanoic acid TFMP ⁻¹ Strifluoromethyl-pyridin-2-ol Epoxiconazole (2RS, 3SR)-1-(2-(2-cholorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1.2,4-triazol - ETU ⁻¹ Ethylenethiourea Fenpropinorphic Cis-4-[3-[4-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine - Fenpropinorphic acid Cis-4-[3-[4-(2-carboxypropyl]-phenyl]-2-methylpropyl]-2,6-imethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Florasulan 2, 6, 8-Trifluoro-5-methoxy-s-triazolo [1, 5-c]pyrimidine-2-sulfonanilide - Florasulan-desmethyl N-(2, 6-difluorophenyl)-8-fluoro-5-hydroxy[1, 2, 4]triazolo[1, 5-c]pyrimidine-2-sulfonanilide - Florasulan-desmethyl N-(2, 6-difluorophenyl)-8-fluoro-2-pyridinyl)oxylacetic acid Glyphosate N-(phosphonomethyl]elycine - AMPA Amino-methyl-bosphonic acid Iodosulfuron-methyl-sodium sodium salt of methyl -1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoite acid - Triazinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonylhenzoit acid NCPA (4-chloro-2-methylphenzyl-2,4-triazine-5-one Met	Dimethoate	O,O-dimethyl S-methylcarbamoylmethyl-phosphorodithioate
- Fluazifop-P^2i(R)-2-(4-((5-(trifluoromethyl)-2-pyridinyl)oxy)phenoxy-propanoic acid 5-trifluoromethyl-pyridin-2-ol Epoxiconazole- TFMP 2i5-trifluoromethyl-pyridin-2-ol (R)-2-(2-chlorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4-triazol- ETU 1iEthylenethioureaFenpropimorphCis-4-[3-[4-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6- dimethylmorpholineFlamprop M-isopropylIsopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninateFlorasulam2, 6-(8-Trifluoro-5-methoxy-s-trizzolo [1,5-c]pyrimidine-2-sulfonanilideFlorasulam2, 6-(8-Trifluoro-5-methoxy-s-trizzolo [1,5-c]pyrimidine-2-sulfonanilideFluroxypyr(4-anino-3,5-dichloro-6-fluro-2-pyridinyl)oxylacetic acidGlyphosateN-chosphonomethylglycine- AMPAAmino-methylphosphonic acidIodosulfuron-methylsodium salt of methyl 4-iodo-2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]lenzio acidIoxynil4-hydroxy-3,5-diiodobenzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoa acidMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenoMcPA(4-chloro-2-methylphenoxy)acetic acidMcPA(4-chloro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Hetribuzin-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metribuzin-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metribuzin-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metribuzin-desamino4,5-dihydro-3-methyl-1,4,5-	Ethofumesate	(±)-2-ethoxy-2.3-dihydro-3.3-dimethylbenzofuran-5-yl-methanesulfonate
$\begin{aligned} & TrMP^{3} \\ TrMP^{3} \\ & TrMP^{3} \\ & S-trifluoromethyl-pyridin-2-01 \\ & Spring \\ &$	- Eluariton- $P^{(2)}$	$(\mathbf{R})_2 - (4 - ((5 - (trifluoromethyl)_2 - nyridinyl)_0xy) nhenoxy-pronanoic acid$
- HWP 5-influoromethyl-pyrdint-2-0i Epoxiconazole (2RS, 3SR)-1-(2-(2-chorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H- 1,2,4-triazol Erporionazole (2RS, 3SR)-1-(2-(2-chorophenyl)-2methylpropyl]-2,6-imethylmorpholine Fempropimorph Cis-4-[3-[4-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6- dimethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Fiamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Florasulam 2, 6, 8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide Fluroxypyr (4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxylacetic acid Glyphosate N-cphosphonomethylglycine - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl-sodium sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2- yl)amino]carbonyl]amino]sulfonyl]benzoate - Triacinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine - Metsulfuron-methyl Methyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoia acid Ioxynil 4-hydroxy-3,5-diiodobenzonitrile Linuron 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea MCPA 4-chlor-2-methylphenoxyacetic acid - 4-chlor-2-methylphenol 4-chlor-2-methylphenoxyacetic acid <td>$TEMD^{2}$</td> <td>5 to fly a new steal new dig 2 sl</td>	$TEMD^{2}$	5 to fly a new steal new dig 2 sl
Epoxiconazole(2RS, S3R)-1-(2-(2-chlorophenyl)-2,3-epoxy-2-(4-thuorophenyl)propyl)-1H- 1,2,4-triazol- ETU $^{(1)}$ EthylenethioureaFenpropimorphCis-4-13-[4-(2,-carboxypropyl)-phenyl]-2-methylpropyl]-2,6- dimethylmorpholine- Fenpropimorphic acidCis-4-13-[4-(2,-carboxypropyl)-phenyl]-2-methylpropyl]-2,6- dimethylmorpholineFlamprop-M-isopropylIsopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate- Flamprop (free acid)N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanineFlorasulam2, 6, 3 Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide- Florasulam-desmethylN-(2,6-difluorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2- sulfonanideFluroxypyr(4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acidGlyphosateN-(phosphonomethylg/grine- AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2- yl)amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCCPA(4-chloro-2-methylphenolMetsulfuron-methylMethyl 2-13-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonanidomethylbylpenzoate- 4-chlor-2-methylphenol4-chlor-2-methylphenolMetsulfuron-methylMethyl 2-13-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methylemplenolMetaulfuron-methylMethyl 2-[[((-de-dimethoxy-6-methyl-1,2,4-triaz	- IFMP	5-trilluorometnyi-pyridin-2-oi
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Epoxiconazole	(2RS, 3SR)-1-(2-(2-chlorophenyl)-2,3-epoxy-2-(4-fluorophenyl)propyl)-1H-
- ETU ^{1/j} Ethylenethiourea Fenpropimorphi Cis-4-[3-[4-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine - Fenpropimorphic acid Cis-4-[3-[4-(2,-carboxypropyl)-phenyl]-2-methylpropyl]-2,6-idimethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine - Flamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine - Florasulam 2, 6, 37 Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanlide - Florasulam-desmethyl N-(2,6-difluorophenyl)-8-fluoro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonanlide Fluroxypyr (4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethylglycine - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl-sodium sodium salt of methyl 1-3,5-triazine - Metsulfuron-methyl Methyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoite Ionynil 4-hydroxy-3,5-diiodobenzonitrile Linuron 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea MCPA (4-chloro-2-methylphenol Metanitron-desamino 4-chlor-2-methylphenol Metanitron-desamino 4-chlor-2-methylphenol Metanitron-desamino 4-chlor-2-methylphenol		1,2,4-triazol
FenpropimorphicCis-4-[3-[4-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholine- Fenpropimorphic acidCis-4-[3-[4-(2-carboxypropyl)-phenyl]-2-methylpropyl]-2,6-imethylmorpholineFlamprop M-isopropylIsopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate- Flamprop (free acid)N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanineFlorasulam2,6.38-Trifluoro-5-methoy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilde- Florasulam-desmethylN-(2,6-difluoro-fs-methoy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilde- Florasulam-desmethylN-(2,6-difluoro-fs-methoy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilde- AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-fl[((4-methoxy-6-methyl-1,3,5-triazin-2-y)lamino]earbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diidobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoty)-a-methylureaMCPA(4-chloro-2-methylphenoty)-1-wethoxy-1-methylureaMCPA(4-chloro-2-methylphenoty)-1-2,4-triazine-5-one- Metanitron4-amino-6-tert-butyl-4,5-dihydro-3-methyltio-1,2,4-triazine-5-one- Metanitron-desamino6-(1,1-dimethylethyl)-3-(methyltio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methyltio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methyltio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-(4-methoxy-6-methyl-1,3,5-triazine-5-one- M	- $ETU^{(1)}$	Ethylenethiourea
- Fenpropinorphic acid Cis-4-[3]4-(2-carboxypropt)-phenyl]-2-methylpropyl]-2.6- dimethylmorpholine Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Flamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Florasulam 2, 6', 8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide - Florasulam-desmethyl N-(2,6-diffluorophenyl)-8-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethyl)lglycine - AMPA Amino-methyl phosphonic acid Jodosulfuron-methyl-sodium sodium salt of methyl-1,3,5-triazine - Triazinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine - Metsulfuron-methyl Methyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfornyl]benzoic acid Ioxynil 4-hydroxy-3,5-diiodobenzonitrile Juminol 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea MCPA (4-chloro-2-methylphenol) Methyl 2-[3-(4,6-dimethoxypyl-benyl-1,2,4-triazine-5-one Metamitron 4-amino-6-tert-butyl-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one Metamitron-desamino 4-5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one Metamitron-desamino 6-tert-butyl-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-dine Metribuzin-desamino 6-	Fenpropimorph	Cis-4-[3-[4-(1,1-dimethylethyl)-phenyl]-2-methylpropyl]-2.6-imethylmorpholine
Perproprintorpart at fail Classify (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (2) (4/2) (- Fannronimorphic acid	$Cis - A_[3_[A_(2_carboxyprony])_phenyl]_2_methylpronyl]_2_6_$
Flamprop-M-isopropyl Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate - Flamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Plorasulam 2', 6', 8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide - Florasulam-desmethyl N-(2,6-difluorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonanilide Fluroxypyr (4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethyl)glycine - AMPA Amino-methylphosphonic acid Idodsulfuron-methyl-sodium sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]sulfonyl]benzoate - Triazinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine - Methyl[zmon-methyl Methyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid Ioxynil 4-hydroxy-3,5-diiodobenzonitrile Linuron 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea MCPA (4-chlor-2-methylphenol 4-chlor-2-methylphenol 4-chlor-3-methyl-1,2,4-triazine-5-one Metamitron 4-anino-4,5-dihydro-3-methyl-1,2,4-triazine-5-one Metamitron 4-anino-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one Metribuzin-desamino 6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5,dione		dimethylmomboline
Flamprop-M-isopropylIsopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine- Flamprop (free acid)N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanineFlorasulam2', 6', 8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilde- Florasulam-desmethylN-(2,6-difluorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonanildeFluroxypyr(4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acidGlyphosateN-(phosphonomethyl)glycine- AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-fl[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]earbonyl]amino]sulfonyl]benzoate- Triazinamine2-anino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxyl-1-methylureaMCPA(4-chloro-2-methylphenoslMetsulfuron-methylMethyl2-[3(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-4.5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metribuzin4-amino-6-tert-butyl-4.5-dihydro-3-methylthio-1,2,4-triazin-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3(methylthio)-1,2,4-triazin-5-die- Metribuzin-desamino6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazin-5-die- Metribuzin-desamino6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazin-5-die- Metribuzin-desamino6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazin-5-die- Metribuzin-diseto		
- Flamprop (free acid) N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine Florasulam 2', 6', 8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide - Florasulam-desmethyl N-(2, 6-difluorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonanilde Fluroxypyr (4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethyl)glycine - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl-sodium sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2-y])amino]acarbonyl]amino]sulfonyl]benzoate - Triazinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine - Metsulfuron-methyl Methyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-y])amino]=carbonyl]amino]-sulfonyl]benzoia caid Ioxynil 4-hydroxy-3,5-diiodobenzonitrile Linuron 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea MCPA (4-chlor-2-methylphenox)acetic acid Mcbrad 4-chlor-2-methylphenoxy acetic acid Mcthor-2-methylphenol 4-chlor-2-methylphenoxy-6-methyl-1,2,4-triazin-5-one Metamitron 4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one Metribuzin 4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one Metribuzin-desamino-diketo 6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-5,dion	Flamprop-M-1sopropyl	Isopropyl N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alaninate
Florasulam 2',6',8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilde - Florasulam-desmethyl N-(2,6-difluorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonaniide Fluroxypyr (4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethyl)glycine - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl-sodium sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2-y])amino]carbonyl]amino]sulfonyl]benzoate - Triazinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]sulfonyl]benzoic acid Ioxynil 4-hydroxy-3,5-diiodobenzonitrile Silionyl]benzoic acid 3-(3.4-dichlorophenyl)-1-methoxy-1-methylurea MCPA (4-chloro-2-methylphenoly)-1-methoxy-1-methylurea MCPA (4-chloro-2-methylphenoly) Metsulfuron-methyl Methyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4-methanesulfonanidomethylbenzoate Metamitron 4-amino-4,5-dihydro-3-methyl-1,2,4-triazin-5-one - Metribuzin 4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazin-5-one - Metribuzin-desamino 6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazin-5-one - Metribuzin 4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazin-5-one - Metribuzin-desamino 6-(1,1	- Flamprop (free acid)	N-benzoyl-N-(3-chloro-4-flourophenyl)-D-alanine
- Florasulam-desmethyl N-(2,6-difluorophenyl)-8-fluro-5-hydroxy[1,2,4]triazolo[1,5-c]pyrimidine-2-sulfonamide Fluroxypyr (4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acid Glyphosate N-(phosphonomethyl)glycine - AMPA Amino-methylphosphonic acid Iodosulfuron-methyl-sodium sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2- yl)amino]carbonyl]amino]sulfonyl]benzoate - Triazinamine 2-amino-4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]sulfonyl]benzoic acid Ioxynil 4-hydroxy-3,5-dicidobenzonitrile Linuron 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea MCPA (4-chloro-2-methylpheno) Messoulfuron-methyl Methyl 2-[[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoate Metamitron 4-amino-4,5-dihydro-3-methyl-1,2,4-triazin-5-one - Metribuzin 4-amino-6-tert-butyl-4,5-dihydro-3-methylhoi-1,2,4-triazin-5-one - Metribuzin-desamino 6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one - Metribuzin-desamino-diketo 6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one - Metribuzin-desamino-diketo 6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dione - Metribuzin-desamino-diketo 6-tert-butyl-4,5-dihydro-3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid <	Florasulam	2',6',8-Trifluoro-5-methoxy-s-triazolo [1,5-c]pyrimidine-2-sulfonanilide
FluroxypyrsulfonamideFluroxypyr(4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acidGlyphosateN-(phosphonomethyl)glycine-AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIonynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenoxy4-chlor-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenoxy)	- Florasulam-desmethyl	N-(2.6-difluorophenyl)-8-fluro-5-hydroxy[1.2.4]triazolo[1.5-c]pyrimidine-2-
Fluroxypyr(4-amino-3,5-dichloro-6-fluro-2-pyridinyl)oxy]acetic acidGlyphosateN-(phosphonomethyl)glycine- AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2-yl)amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-Ioxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenolMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-sanino-6,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metamitron4-sanino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-t		sulfonamide
HutoxypyCreatmics_j.suchthologram(GlyphosateN-(phosphonmethyl/glycine- AMPAAmino-methyl/phosphonic acidlodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenolxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-schlor-2-methylphenolMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo10-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione<	Flurovypyr	(A amino 3.5 dichloro 6 fluro 2 pyridinyl)ovylacetic acid
ChypnosateN-(phosphonomethyl)glycine- AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2- yl)amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chlor-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenolMethyl 2-[3(-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-6-tert-butyl-4,5-dihydro-3-methyl-1,2,4-triazin-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo9-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Methyl2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]hethyl]benzoic acid- Metribuzin-diketo9-(1-ethyl)-2,6-(initro-3,4-xym	Thuroxypyr	(4-anino-5,5-dicinoro-o-nuro-2-pyndinyr)oxy jacetic acid
- AMPAAmino-methylphosphonic acidIodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2- yl)amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodbenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chloro-2-methylphenolMesosulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2,yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl]amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl]benzoic acid- Methyl2-[[[[(4-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl]benzoic acid- Methyl2-[[[[10,6-dimethoxy-2-pyrimidinyl]benzoic acid- Methyl2-[[[[11,6-dimethoxy-2-pyrimidinyl]amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl]benzoic acid- Methyl2-[11,4-6-dimethoxy-2-pyrimidinyl]benzoic acid- Methyl2-[[[11,4,6-dime	Glyphosate	N-(phosphonomethyl)glycine
Iodosulfuron-methyl-sodiumsodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2- yl)amino]carbonyl]amino]sulfonyl]benzoate- Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine- Metsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chloro-2-methylphenoxy)acetic acidMetsulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-6-tert-butyl-6-phenyl-1,2,4-triazin-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-dione- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Methylfuron(4-chlory-2-pyrimidinyl)amino]carbonyl]amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[(4,6-dimethoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[(4,6-dimethoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Methylloron2-[[[(4,6-dimethoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Methylloron2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl	- AMPA	Amino-methylphosphonic acid
$\begin{tabular}{lllllllllllllllllllllllllllllllllll$	Iodosulfuron-methyl-sodium	sodium salt of methyl 4-iodo-2-[[[[(4-methoxy-6-methyl-1,3,5-triazin-2-
Triazinamine2-amino-4-methoxy-6-methyl-1,3,5-triazine $Metsulfuron-methyl$ Methyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenolMesosulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5,-dione- Metribuzin-desamino6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMethyl2-[[[((4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[((4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ^{3/} 3-phenyl-4-hydroxy-6-chloropyridazine		vl)amino]carbonvl]amino]sulfonvl]benzoate
Metsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxyacetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenolMetamitron4-chlor-2-methylphenolMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-oneMetamitron4-amino-6-tert-butyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino-diketo6-(ert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-desamino-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Methylloron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acid- Methyl-103-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- MHPCMethyl-N-(3-hydoxyphenyl-carbamate- PHCP 3 3-phenyl-4-hydroxy-6-chloropyridazine	- Triazinamine	2-amino-4-methoxy-6-methyl-1 3 5-triazine
Methylar birdingMethylar birdingIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chlor-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenoxy)acetic acidMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-desamino6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMethyl2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acid- Menomethylam3-((methoxycarbonyl)amino]henyl(3-methylphenyl)carbamate- MHPCMethyl-(3-hydoxyphenyl)-carbamate- MHPC1-amino-3-hydroxybenzene- PHCP $^{3'}$ 3-phenyl-4-hydroxy-6-chloropyridazine	- Matsulfuron-mathyl	Methyl2-[[[[(A_methovy_6_methyl_1 3 5_triazine_2_v])amino]=carbonyl]amino]
sulfonyljbenzoic acidIoxynil4-hydroxy-3,5-diiodobenzonitrileLinuron3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenolMesosulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4- methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metamitron desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2,yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[((4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [((methylsulfonyl)amino]methyl]benzoic acid- MetholinN-(1-ethyl)-2,6-dinitro-3,4-xynilePendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- PHPC3-phenvl-4-hydroxy-6-chloropyridazine	- Meisuljuron-meinyi	Methyl2-[[[(4-methoxy-0-methyl-1,5,5-thazme-2-yl]ammo]-caroonyl]ammo]-
loxynil4-hydroxy-3,5-diiodobenzonitrileLinuron $3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenoy]Mesosulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4-methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-2,1)amino]=carbonyl]amino]-sulfonyl]benzoic acid- Mesosulfuron2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-[(methylsulfonyl)amino]methyl]benzoic acidPhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenzene- 3$		sulfonyl]benzoic acid
Linuron $3-(3,4-dichlorophenyl)-1-methoxy-1-methylureaMCPA(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chloro-2-methylphenoxy)acetic acidMesosulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4-methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)- 1,2,4-triazine-3,5-dione- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid- Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-[[(methylsulfonyl)amino]methyl]benzoic acid- Menedipham3-((methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxybenzene- 3-aminophenol1-amino-3-hydroxybenzene- PHCP 313-phenyl-4-hydroxy-6-chloropyridazine$	loxynil	4-hydroxy-3,5-diiodobenzonitrile
MCPA $(4-chloro-2-methylphenoxy)acetic acid- 4-chlor-2-methylphenol4-chlor-2-methylphenolMesosulfuron-methylMethyl 2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4-methanesulfonamidomethylbenzoateMetamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-oneMetribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-3-methylthio)-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[(4,6-dimethoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid- Mesosulfuron2-[[[((4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-[[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhemedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N.(3-hydoxyphenyl)-carbamate- MHPC1-amino-3-hydroxybenzene- 3-aminophenol1-amino-3-hydroxybenzene- PHCP 3j3-phenyl-4-hydroxy-6-chloropyridazine$	Linuron	3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea
$\begin{array}{llllllllllllllllllllllllllllllllllll$	MCPA	(4-chloro-2-methylphenoxy)acetic acid
Mesosulfuron-methylMethyl $2-[3-(4,6-dimethoxypyrimidin-2-yl)ureidosulfonyl]-4-$ methanesulfonamidomethylbenzoateMetamitron $4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one$ - Metamitron-desamino $4,5$ -dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one- Metribuzin $4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metsulfuron-methylMethyl2-[[[(4,6-dimethoxy-2-pyrimidinyl]amino]=carbonyl]amino]=carbonyl]amino]-sulfonyl]benzoic acid- Mesosulfuron2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-[[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ^{3j}3-phenyl-4-hydroxy-6-chloropyridazine$	- 4-chlor-2-methylphenol	4-chlor-2-methylphenol
Metasyl 2-[5-(4,6-dimethoxyp)]methanesulfonamientoxyp)methane	Mesosulfuron methyl	Methyl 2 [3 (A 6 dimethownwrimidin 2 yd)ureidosulfonyl] A
Metamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-oneMetribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)- 1,2,4-triazine-5-(4H)-one- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-[[(methylsulfonyl)amino]methyl]benzoic acidN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- PHCP ³ 3-phenyl-4-hydroxy-6-chloropyridazine	Wesosulfulon-methyl	methonoculfonomidomethylbonzoete
Metamitron4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one- Metamitron-desamino4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-oneMetribuzin4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino $6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-3,5-dione- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-PendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- MHPC3-aminophenol- PHCP ^{3)}3-phenyl-4-hydroxy-6-chloropyridazine$		
- Metamitron-desamino $4,5$ -dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-oneMetribuzin 4 -amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino 6 -(1,1-dimethylethyl)-3-(methylthio)- 1,2,4-triazine-5-(4H)-one- Metribuzin-desamino-diketo 6 -tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo 4 -amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione- Metribuzin-diketo 4 -amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[((4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid 2 -[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-PendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham 3 -[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- PHCP 3 3 -phenyl-4-hydroxy-6-chloropyridazine	Metamitron	4-amino-4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazin-5-one
Metribuzin $4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one- Metribuzin-desamino6-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazine-5-(4H)-one- Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[((4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-sulfonyl]benzoic acid2-[[[((4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4-PendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- PHCP ^{3}3-phenyl-4-hydroxy-6-chloropyridazine$	- Metamitron-desamino	4,5-dihydro-3-methyl-6-phenyl-1,2,4-triazine-5-one
 Metribuzin-desamino Metribuzin-desamino-diketo Metribuzin-diketo Metribuzin-diketo Metribuzin-diketo Metsulfuron-methyl G-(1,1-dimethylethyl)-3-(methylthio)-1,2,4-triazin-3,5-dione 4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dione Methyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid Mesosulfuron 2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acid N-(1-ethyl)-2,6-dinitro-3,4-xynile Phenmedipham -[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate MHPC Methyl-N-(3-hydoxyphenyl)-carbamate - Aminophenol - PHCP³ 3-phenyl-4-hydroxy-6-chloropyridazine 	Metribuzin	4-amino-6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-5-one
• Metribuzin-desamino-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dione• Metribuzin-diketo6-tert-butyl-4,5-dihydro-3-methylthio-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid• Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate• MHPCMethyl-N-(3-hydoxyphenyl)-carbamate• PHCP ³ 3-phenyl-4-hydroxy-6-chloropyridazine	- Metribuzin-desamino	6-(1,1-dimethylethyl)-3-(methylthio)- 1.2.4-triazin-5-(4H)-one
Metributin distantio-diretoOtercodi/1-4,5-dihydro-3-metriydro-3,5-dione- Metribuzin-diketo4-amino-6-tert-butyl-4,5-dihydro-1,2,4-triazine-3,5-dioneMetsulfuron-methylMethyl2-[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³ 3-phenyl-4-hydroxy-6-chloropyridazine	- Metribuzin-desamino-diketo	6-tert-butyl-4 5-dibydro-3-methylthio-1 ? 4-triazine-3 5-dione
- Metriolizin-aiketo4-amino-6-tert-butyl-4,5-dinydro-1,2,4-triazine-5,5-dioneMetsulfuron-methylMethyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³ 3-phenyl-4-hydroxy-6-chloropyridazine	Metribusin dilata	A suring C tast hutsil 4.5 dihadra 1.2.4 trianing 2.5 diana
Metnyl2-[[[[(4-methoxy-6-methyl-1,3,5-triazine-2-yl)amino]=carbonyl]amino]- sulfonyl]benzoic acid- Mesosulfuron2-[[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³)3-phenyl-4-hydroxy-6-chloropyridazine	- meiribuzin-aiketo	4-anno-o-tert-buty1-4,5-dinydro-1,2,4-triazine-5,5-dione
sulfonyl]benzoic acid- Mesosulfuron2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP 3)3-phenyl-4-hydroxy-6-chloropyridazine	Metsulfuron-methyl	Methy12-[[[[(4-methoxy-6-methy1-1,3,5-triazine-2-yl)amino]=carbonyl]amino]-
- Mesosulfuron2-[[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]amino]sulfonyl]-4- [[(methylsulfonyl)amino]methyl]benzoic acidPendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³⁾ 3-phenyl-4-hydroxy-6-chloropyridazine		sulfonyl]benzoic acid
Increasing a contract of the second part of the second	- Mesosulfuron	2-[[[[(4 6-dimethoxy-2-pyrimidinyl)aminolcarbonyllaminolsulfonyll_4
PendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³⁾ 3-phenyl-4-hydroxy-6-chloropyridazine	11C30301ju1011	2 [[[(¬,o-unionoxy-2-pyrindunyr/aninojcaroonyrjaninojsunonyrj-4-
PendimethalinN-(1-ethyl)-2,6-dinitro-3,4-xynilePhenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³⁾ 3-phenyl-4-hydroxy-6-chloropyridazine		[[(ineutyisuitonyi)aminojmetnyi]benzoic acid
Phenmedipham3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP ³⁾ 3-phenyl-4-hydroxy-6-chloropyridazine	Pendimethalin	N-(1-ethyl)-2,6-dinitro-3,4-xynile
- MHPCMethyl-N-(3-hydoxyphenyl)-carbamate- 3-aminophenol1-amino-3-hydroxybenzene- PHCP 3)3-phenyl-4-hydroxy-6-chloropyridazine	Phenmedipham	3-[(methoxycarbonyl)amino]phenyl (3-methylphenyl)carbamate
- <i>3-aminophenol</i> - <i>PHCP</i> ³ 1-amino-3-hydroxybenzene 3-phenyl-4-hydroxy-6-chloropyridazine	- MHPC	Methyl-N-(3-hydoxyphenyl)-carbamate
- PHCP ³ 3-phenyl-4-hydroxy-6-chloropyridazine	- 3-aminophenol	1-amino-3-hydroxybenzene
	- PHCP ³⁾	3-phenyl-4-hydroxy-6-chloropyridazine

 Table A1.1 Systematic chemical nomenclature for the pesticides and degradation products encompassed by the PLAP.

Parameter	Systematic chemical nomenclature
Picolinafen	4'-fluoro-6-(α,α,α-trifluoro-m-tolyloxy)pyridine-2-carboxanilide
- CL153815	6-(3-trifluoromethylphenoxy)-2-pyridine carboxylic acid
Pirimicarb	2-(dimethylamino)-5,6-dimethyl-4-pyrimidinyldimethylcarbamate
- Pirimicarb-desmethyl	2-(dimethylamino)-5,6-dimethyl-4-pyrimidinylmethylcarbamate
- Pirimicarb-desmethyl-	2-methylformamido-5,6-dimethylpyrimidine-4-yl dimethylcarbamate
formamido	
Propiconazole	1-[[2-(2,4-dichlorophenyl)-4-propyl-1,3-dioxolan-2-yl]methyl]-1H-1,2,4-triazole
Propyzamide	3,5-dichloro-N-(1,1-dimethylprop-2-ynyl)benzamide
- RH-24644	2-(3,5-dichlorophenyl)-4,4-dimethyl-5-methylene-oxalzoline
- RH-24580	N-(1,1-dimethylacetonyl)-3,5-dichlorobenzamide
- RH-24655	3,5-dichloro-N-(1,1-dimethylpropenyl)benzamide
Prosulfocarb	N-[[3-(4-methoxy-6-methyl-1,3,5-triazin-2-yl)-3-[2-(3,3,3,-
	trifluro=propyl)phenylsulfonyl]urea
Rimsulfuron	N-[[(4,6-dimethoxy-2-pyrimidinyl)amino]carbonyl]-3-(ethylsulfonyl)-2-
	pyridinesulfonamide
- PPU	N-(4,6-dimethoxy-2-pyrimidinyl-N-((3-ethylsulfonyl)-2-pyridinyl)urea (IN70941)
- PPU-desamino	N-((3-(ethylsulfonyl)-2-pyridyl)-4,6 dimethoxy-2 pyrimidinamine (IN70942)
Terbuthylazine	6-chloro-N-(1,1-dimethylethyl)-N-ethyl-1,3,5,triazine-2,4-diamine
- Desethyl-terbuthylazine	6-chloro-N-(1,1-dimethylethyl)-1,3,5,triazine-2,4-diamine
- Desisopropyl-atrazine	6-chloro-N-ethyl-1,3,5,triazine-2,4-diamine
- 2-hydroxy-desethyl-	6-hydroxy-N-(1,1-dimethylethyl)-1,3,5,triazine-2,4-diamine
terbuthylazine	
- 2-hydroxy-terbuthylazine	6-hydroxy-N-(1,1-dimethylethyl)-N´-ethyl-1,3,5,triazine-2,4-diamine
Tebuconazole	a-[2-(4-chlorophenyl)ethyl]-a-(1,1-dimethylethyl)-1H-1,2,4-triazole-1-ethanol
Thiamethoxam	3-(2-cholro-thiazol-5-ylmethyl)-5-methyl[1,3,5]oxadiazinan-4ylidene-N-nitroamine
- CGA 322704	[C(E)]-N-[(2-chloro-5-thiazolyl)methyl]-N'-methyl-N'-nitroguanidine
- Triazinamin-methyl ⁴⁾	4-methoxy-6-methyl-1,3,5-triazin-methylamine
Triflusulfuron	methyl 2-[4-dimethylamino-6-(2,2,2-trifluoroethoxy)-1,3,5-triazin-2-
	ylcarbamoylsulfamoyl]-m-toluate
- IN-E7710	N-methyl-6-(2,2,2-trifluoroethoxy)-1,3,5-triazine-2,4-diamine
- IN-D8526	N,N-dimethyl-6-(2,2,2-trifluoroethoxy)-1,3,5-triazine-2,4-diamine
- IN-M7222	6-(2,2,2-trifluoroethoxy)-1,3,5-triazine-2,4-diamine
Triasulfuron	1-[2-(2-chloroethoxy)phenylsulfonyl]-2-(4-methoxy-6-methyl-1,3,5-triazine-2-yl)-urea
¹ Degradation product of ma	incozeb.

 Table A1.1 (continued) Systematic chemical nomenclature for the pesticides and degradation products encompassed by the PLAP.

²⁾Degradation product of mancozep.
 ²⁾Degradation product of fluazifop-P-butyl.
 ³⁾Degradation product of pyridate.
 ⁴⁾Degradation product of Tribenuron-methyl.

From each of the PLAP sites, samples were collected of groundwater, drainage water and soil water in the unsaturated zone. A full description of the monitoring design and sampling procedure is provided in Lindhardt *et al.* (2001) and Kjær *et al.* (2003), respectively.

Until March 2002, pesticide analysis was performed monthly on water samples from the suction cups located both 1 m b.g.s. and 2 m b.g.s., from two screens of the horizontal monitoring wells and from two of the downstream vertical monitoring wells. In addition, more intensive monitoring encompassing all four groups of suction cups, six screens of the horizontal monitoring wells and five monitoring wells was performed every four months (Kjær *et al.*, 2002). At the loamy sites, the pesticide analysis was also performed on drainage water samples.

The monitoring programme was revised in March 2002 and the number of pesticide analyses was reduced. At the loamy sites, pesticide analysis of water sampled from the suction cups was ceased, and the monthly monitoring was restricted to just one monitoring well. At Jyndevad, pesticide analysis of the suction cups located 2 m b.g.s. was ceased and the interval for the intensive monitoring encompassing the larger number of monitoring screens was extended to six months, except for the suction cups 2 m b.g.s. at Tylstrup, where the four-month interval was retained (Kjær *et al.*, 2003).

On the sandy soils, the analysis of a number of pesticides in water from the monitoring wells had to be further reduced, due to economical constraints imposed by the high prices on pesticide analysis. This reduction was based on results from the suction cups implying that leaching risk of certain pesticides was negligible, why analysis of a limited number of groundwater samples would be reasonable (see Table A5.1 and Table A5.2 in Appendix 5).

In March 2008, a new revision of the monitoring programme was completed resulting in an optimization of the programme including an additional reduction in the sampling programme (Table A2.1). On the loamy sites, sampling from the suction cups for inorganic analysis, from one-two monitoring wells per site, and one horizontal well at Silstrup (H2) and Faardrup (H1) was suspended. On the sandy sites, only sampling from the monitoring well M6 at Tylstrup has been suspended (see Rosenbom *et al.*, 2010b for details).

Table A2.1. Pesticide monitoring programme in suction cups (S), horizontal monitoring wells (H) and vertical
monitoring wells (M) as of March 2009. Water sampling places (S, H, and M) from where sampling stopped in 2008
and 2009 are given in bold. Well M10 at Silstrup was included in the programme on 5 February 2009.

e given in colar wen hi	to a photop was more and in the program	
Monthly monitoring	Half-yearly monitoring	Not
(Intensive)	(Extensive)	Monitored
M4, M5, S1a, S2a	M1, M3, M4, M5, S1a , S2a, S1b [*] ,	M2, M6 , M7
	$S2b^*$	
M1, M4, S1a, S2a	M1, M2, M4, M5, M7, S1a, S2a	M3, M6 , S1b, S2b
M5, H1.2	M5, M9, M10, M12, H1.1, H1.2, H1.3	M1, M2, M4, M6, M8, M7, M11, M13,
		H2.1, H2.2, H2.3
M4, H1.2	M1, M4, M5, M6, H1.1, H1.2, H1.3	M2, M3 , M7 ,
M4, M5, H2.3	M4, M5, M6, H2.1, H2.3, H2.5	M1, M2, M3, M7, H1.1, H1.2, H1.3
	Monthly monitoring (Intensive) M4, M5, S1a, S2a M1, M4, S1a, S2a M5, H1.2 M4, H1.2 M4, M5, H2.3	Monthly monitoring (Intensive) Half-yearly monitoring (Extensive) M4, M5, S1a, S2a M1, M3, M4, M5, S1a , S2a, S1b [*] , S2b [*] M1, M4, S1a, S2a M1, M2, M4, M5, M7, S1a, S2a M5, H1.2 M5, M9, M10, M12, H1.1, H1.2, H1.3 M4, M5, H2.3 M4, M5, M6, H2.1, H2.3, H2.5

S1a and S1b refer to suction cups installed 1 and 2 m b.g.s., respectively, at location S1, whereas S2a and S2b refer to suction cups installed 1 and 2 m b.g.s., respectively, at location S2.^{*} At Tylstrup suctions cups installed 2 m b.g.s are monitored four times a year(see text).

Until July 2004, pesticide analyses were performed weekly on water sampled timeproportionally from the drainage system. Moreover, during storm events additional samples (sampled flow-proportionally over 1-2 days) were also analysed for pesticides. In June 2004 the drainage monitoring programme was revised. From July 2004 and onwards pesticide analyses were done weekly on water sampled flow- proportionally from the drainage water system. See Kjær *et al.* 2003 for further details on the methods of flow-proportional sampling. The weighted average concentration of pesticides in the drainage water was calculated according to the following equation:

$$C = \frac{\sum_{i=1}^{n} M_i}{\sum_{i=1}^{n} V_i}$$

 $M_i = C_i \cdot V_i$

where:

n = Number of weeks within the period of continuous drainage runoff.

 V_i = Weekly accumulated drainage runoff (mm/week).

 C_i = Pesticide concentration collected by means of the flow-proportional sampler (µg/l).

Until July 2004 where both time and flow-proportional sampling was applied the numbers were:

 $M_i = Ct_i \cdot V_i$ If no flow event occurs within the i'th week $M_i = Cf_i \cdot Vf_i$ If a flow event occurs within the i'th week and if $Cf_i \cdot Vf_i > Ct_i \cdot V_i$

where:

n = Number of weeks within the period of continuous drainage runoff.

 V_i = Weekly accumulated drainage runoff (mm/week).

 $Vf_i = Drainage runoff accumulated during a "flow event" (mm/storm event).$

 Cf_i = Pesticide concentration in the "event samples" collected by means of the flow-proportional sampler (µg/l).

 Ct_i = Pesticide concentration in the weekly samples collected by means of the time-proportional sampler (µg/l).

Tables 9, 11, and 13 report the weighted average leachate concentration in the drainage water within the first drainage season after application. In these tables this calculation period is defined as the period from the date of application until 1 July the following year, as pesticides are usually present in the first drainage runoff occurring after application of pesticide.

On the sandy soils the weighted average concentration of pesticides leached to the suction cups situated 1 m b.g.s. was estimated using the measured pesticide concentration and estimated percolation on a monthly basis. Pesticide concentrations measured in suction cups S1 and S2 were assumed to be representative for each sample period. Moreover, accumulated percolation rates deriving from the MACRO model were assumed to be representative for both suction cups S1 and S2. For each of the measured concentrations, the corresponding percolation (Perc.) was estimated according to the equation:

$$P_i = \sum_{t_1}^{t_2} P_t$$

where $t = sampling date; t_1 = 0.5(t_{i-1}+t_i); t_2=0.5(t_i+t_{i+1})$ $P_t = daily percolation at 1 m b.g.s.$ as estimated by the MACRO model (mm) The average concentration was estimated according to the equation:

$$C = \frac{\sum C_i \cdot P_i}{\sum P_i}$$

where

 C_i = measured pesticide concentration in the suction cups located 1 m b.g.s.

Tables 3 and 6 report the weighted average leachate concentration. In these tables this calculation period is defined as the period from the date of first detection until 1 July the following year. On sandy soils the transport of pesticides down to the suction cups situated at 1 m depth may take some time. In most cases the first detection of pesticides occurs around 1 July, why the reported concentration represents the yearly average concentration. In a few cases the first detection of pesticides occurs later, but this later occurrence does not affect the weighted average calculation. E.g. the reported average concentration using a calculation period from the first detection until 1 July the following year is equal to that using a calculation period of a year (1 July – 30 June) the following year). Unless noted the concentrations. In the few cases where reported concentrations are either not representative for an annual average concentration or not representative for the given leaching pattern (leaching increases the second or third year after application) a note is inserted in the table.

Appendix 2. Pesticide monitoring programme - Sampling procedure

- mere no promoto	
Date	Management practice
09.02.07	Herbicide - 1.0 l/ha Kerb 500 SC (propyzamide)
27.03.07	Herbicide - 0.8 l/ha Matrigon (clopyralid)
08.06.07	Irrigation 30 mm
01.08.07	Direct harvest and simultaneous shredding of straw (seed yield 24.5 hkg/ha 91% DM, straw yield)
03.08.07	Rotary cultivated - depth 3.0 cm (straw incorporation)
07.09.07	Rotary cultivated - depth 7.0 cm (straw incorporation)
12.09.07	Ploughed - 22 cm depth
12.09.07	Winter wheat sown - cv. Smuggler.
18.10.07	Herbicide - 5.0 l/ha Stomp (pendimethalin)
16.11.07	Herbicide - 1.0 l/ha Folicur EC250 (tebuconazole)
22.05.08	Irrigation - 32 mm
29.05.08	Irrigation - 32 mm
05.06.08	Irrigation - 32 mm
13.06.08	Irrigation - 30 mm
17.06.08	Fungicide – 1.0 l/ha Amistar (azoxystrobin)
18.08.08	Winter wheat harvested (seed yield 92.1 hkg/ha 85% DM)
31.08.08	Straw yield (18.5 hkg/ha 100% DM)
10.04.09	Ploughed - 24 cm depth
10.04.09	Rolled with a concrete roller
14.04.09	Spring barley sown - cv. Keops
15.05.09	Herbicide - 1.5 l/ha Basagran M75 (bentazone + MCPA)
23.06.09	Fungicide - 1.0 l/ha Amistar (azoxystrobin) – fungi
29.06.09	Irrigation - 26 mm
08.07.09	Mavrik (tau-fluvalinate) - pests - 0.1 l/ha (not analysed)
08.07.09	Irrigation – 27 mm
20.08.09	Harvest of spring barley. Grain yield 53.4 hkg/ha, 85% DM
28.08.09	Straw removed, yield 17.4 hkg/ha 100% DM
04.04.10	Ploughed - 24 cm depth
26.04.10	Rolled with concrete roller
04.05.10	Seedbed preparation - 10 cm depth
06.05.10	Planting of potatoes - cv. Kuras
17.05.10	Ridging
26.05.10	Herbicides - 1.0 l/ha Fenix (aclonifen) + 10 g/ha Titus WSB (rimsulfuron)
08.06.10	Herbicide - 20 g/ha Titus WSB (rimsulfuron)
15.06.10	Fungicide - 0.2 l/ha Ranman (cyazofamid)
24.06.10	Fungicide - 0.2 l/ha Ranman (cyazofamid)
01.07.10	Fungicide - 0.2 l/ha Ranman (cyazofamid)

Table A3.1 Management practice at **Tylstrup** during the 2007 to 2010 growing seasons. The active ingredients of the various pesticides are indicated in parentheses.

the various pest	leides are indicated in parentileses.
Date	Management practice
13.04.07	Plant growth inhibitor – 1.0 l/ha Cycocel 750 (Chlormequat-chloride)
27.04.07	Irrigation - 27 mm
07.05.07	Herbicide - 1.0 l/ha Opus (epoxiconazole)
05.06.07	Irrigation - 27 mm
07.08.07	Harvest of triticale (seed yield 38.7 hkg/ha 85% DM, straw yield 38.3 hkg/ha 100% DM)
13.09.07	Herbicide - 2.0 l/ha Roundup (glyphosate, not monitored)
28.09.07	Ploughed - 22 cm depth
29.09.07	Rolled with a concrete roller
01.10.07	Winter wheat sown – cv. Ambition
29.10.07	Herbicide - 0.133 g/ha Pico 750 WG (picolinafen)
03.12.07	Fungicide - 1.0 l/ha Folicur EC 250 (tebuconazole)
07.05.08	Irrigation - 42 mm
14.05.08	Irrigation - 27 mm
21.05.08	Irrigation - 27 mm
30.05.08	Irrigation - 30 mm
05.06.08	Irrigation - 35 mm
11.06.08	Fungicide - 1.0 l/ha Amistar (azoxystrobin)
25.06.08	Irrigation - 35 mm
08.07.08	Irrigation - 30 mm
30.08.08	Winter wheat harvested (seed yield 68.1 hkg/ha 85% DM, straw yield 28.1 hkg/ha 100% DM)
17.03.09	Ploughed - 22 cm depth
18.03.09	Rolled with a concrete roller
18.03.09	Spring barley sown cv. Simba
27.04.09	Herbicide - 1.2 l/ha Fox 480 SC (bifenox)
11.05.09	Herbicide - 1.5 l/ha Basagran M75 (bentazone+ MCPA)
26.05.09	Fungicide - 1.5 l/ha Bell (boscalide + epoxiconazole)
27.05.09	Irrigation - 30 mm
05.06.09	Irrigation - 27 mm
29.06.09	Irrigation - 27 mm
07.08.09	Harvest of spring barley. Grain yield 64.0 hkg/ha 85% DM, straw yield 19.5 hkg/ha 100% DM
14.04.10	Ploughed. Depth 24 cm
15.04.10	Rolled with concrete roller
22.04.10	Seedbed preparation - 9 cm depth
04.05.10	Planting of potatoes - cv. Kuras
04.05.10	Ridging
27.05.10	Herbicides - 1.0 l/ha Fenix (aclonifen) + 10 g/ha Titus WSB (rimsulfuron)
08.06.10	Herbicide - 20 g/ha Titus WSB (rimsulfuron)
24.06.10	Irrigation - 25 mm
28.06.10	Fungicide - 0.2 l/ha Ranman (cyazofamid)
30.06.10	Irrigation - 25 mm

Table A3.2 Management practice at **Jyndevad** during the 2007 to 2010 growing seasons. The active ingredients of the various pesticides are indicated in parentheses.

Date	Management practice
13.04.07	Herbicide - 100 ml/ha Hussar OD (iodosulfuron)
13.04.07	Growth retardant - 1.2 l/ha Cycocel 750 (chlormequat-chloride)
07.06.07	Fungicide - 1.0 l/ha Opus (epoxiconazole)
24.08.07	Winter wheat harvested (seed yield 100.7 hkg/ha 85% DM, straw yield 40.8 hkg/ha 100% DM,
	shredded at harvest
29.08.07	Stubble harrowed, heavy disk harrow (Dalbo) - 5 cm depth
12.11.07	Ploughed - 27 cm depth
07.05.08	Fodder beet sown - cv. Kyros
22.05.08	Herbicide - 30 g/ha Safari (triflusulfuron) + 0.5 l/ha Goliath (metamitron) + 1.5 l/ha Betanal
	(phenmedipham)
30.05.08	Herbicide - 30 g/ha Safari (triflusulfuron) + 0.5 l/ha Goliath (metamitron) + 1.5 l/ha Betanal
	(phenmedipham) + 0.07 l/ha Tramat 500 SC (ethofumesate)
17.06.08	Herbicide - 30 g/ha Safari (triflusulfuron) + 0.5 l/ha Goliath (metamitron) + 1.5 l/ha Betanal
	(phenmedipham) + 0.07 l/ha Tramat 500 SC (ethofumesate)
26.06.08	Insecticide - 0.30kg/ha Pirimor G (pirimicarb)
01.07.08	Herbicide - 3.0 l/ha Fusilade Max (fluazifop-P-butyl)
04.07.08	Herbicide - 30 g/ha Safari (triflusulfuron) + 0.5 l/ha Goliath (metamitron) + 1.5 l/ha Betanal
	(phenmedipham)
09.07.08	Insecticide - 0.300 kg/ha Pirimor G (pirimicarb)
27.10.08	Fodder beet harvested. Yield of root 17.3 t/ha 100% DM, yield of top 5.15 t/ha 100% DM
15.12.08	Ploughed - 23 cm depth
02.04.09	Tracer - 31.5 kg/ha potassium bromide
11.04.09	Rolled with Cambridge roller
11.04.09	Spring barley sown - cv. Keops; undersown red fescue cv. Jasperina
19.05.09	Herbicide -1.25 l/ha Fighter 480 (bentazone)
24.06.09	Fungicide - 1.0 l/ha Amistar (azoxystrobin)
16.07.09	Wholecrop harvest of spring barley - 94.6 hkg/ha 100% DM
24.08.09	Herbicide - 0.020 l/ha Hussar OD (iodosulfuron)
09.09.09	Herbicide - 1.5 l/ha Fox 480 SC (bifenox)
02.05.10	Herbicide - 1.5 l/ha Fusilade Max (fluazifop-P-butyl)
05.05.10	Herbicides - 0.1 l/ha Hussar OD (iodosulfuron) + 0.7 l/ha SweDane MCPA 750 (not analyzed)

Table A3.3 Management practice at **Silstrup** during the 2007 to 2010 growing seasons. The active ingredients of the various pesticides are indicated in parentheses.

Date	Management practice
11.04.07	Growth retardant - 1.2 l/ha Cycocel 750 (chlormequat-chloride)
31.05.07	Fungicide - 1.0 l/ha Opus (epoxiconazole)
07.08.07	Winter wheat harvested (seed yield 81.5 hkg/ha, 85% DM)
08.08.07	Straw shredded (47.4 hkg/ha, 100% DM)
14.09.07	Herbicide - 1.5 l/ha Roundup Max (glyphosate)
02.10.07	Ploughed - depth 20 cm (packed with a ring roller)
03.10.07	Winter wheat sown – cv. Frument.
30.10.07	Herbicide - 0.133 g/ha Pico 750 WG (picolinafen)
22.11.07	Fungicide - 1.0 l/ha Folicur EC 250 (tebuconazole)
13.06.08	Fungicide - 1.0 Amistar (azoxystrobin)
16.08.08	Winter wheat harvested (seed yield 83.8 hkg/ha 85% DM)
16.08.08	Straw shredded - 40.7 hkg/ha 100% DM
12.03.09	Ploughed - depth 18 cm - packed with a ring roller
06.04.09	Tracer - 30 kg/ha potassium bromide
08.04.09	Spring barley sown - cv. Keops
08.04.09	Rolled with a cambridge roller
01.05.09	Herbicide - 1.2 l/ha Fox 480 SC (bifenox)
14.05.09	Herbicide - 1.5 l/ha Basagran M75 (bentazone/MCPA)
04.06.09	Fungicide - 1.0 l/ha Amistar (azoxystrobin)
07.08.09	Spring barley harvested. Grain yield 71.4 hkg/ha, 85% DM
07.08.09	Straw shredded. 39.9 hkg/ha, 100% DM
24.08.09	Ploughed - 20 cm depth - packed with a ring roller
24.08.09	Rotor harrowed - 4 cm depth
24.08.09	Winter rape sown - cv. Cabernet
25.08.09	Herbicide - 0.33 l/ha Command CS (clomazone)
30.09.09	Herbicide - 0.75 l/ha Fox 480 SC (bifenox)
09.10.09	Insecticide - 0.15 l/ha Cyperb (cypermethrin) (not analysed)
20.04.10	Field partially resown with spring rape - cv. Pluto
10.05.10	Insecticide - 0.3 l/ha Biscay OD 240 (thiacloprid)

Table A3.4 Management practice at **Estrup** during the 2007 to 2010 growing seasons. The active ingredients of the various pesticides are indicated in parentheses.

1	L
Date	Management practice
10.08.07	Stubble cultivation - 15 cm depth
22.08.07	Stubble cultivation - 15 cm depth
18.09.07	Ploughed and packed - 25 cm depth
18.09.07	Winter wheat sown – cv. Ambition
09.10.07	Herbicide - 5.0 l/ha Stomp (pendimethalin)
20.11.07	Fungicide - 1.0 l/ha Folicur 250 (tebuconazole)
20.08.08	Winter wheat harvested (seed yield 89.6 hkg 85% DM, straw yield 65.2 hkg/ha 100% DM)
26.08.08	Tracer - 30 kg/ha potassium bromide
01.12.08	Ploughing - 23 cm depth
05.04.09	Sugar beet sown - cv. Palace
24.04.09	Herbicide - 1.5 l/ha Betanal (phenmedipham) + 1.0 l/ha Goliath (metamitron)
30.04.09	Herbicide - 10 g/ha Safari (triflusulfuron) + 1.5 l/ha Betanal (phenmedipham) + 1.0 l/ha Goliath
11.05.09	Herbicide - 10 g/ha Safari (triflusulfuron) + 1.5 l/ha Betanal (phenmedipham) + 1.0 l/ha Goliath
14.05.09	Herbicide - 1.0 l/ha Focus Ultra (cycloxydim)
17.06.09	Herbicide - 1.0 l/ha Focus Ultra (cycloxydim)
06.10.09	Harvest of sugar beet. Root yield 147.9 hkg/ha 100% DM, top yield 40.1 hkg/ha 100% DM
01.11.09	Ploughing - 20 cm depth
07.04.10	Seedbed preparation - 6 cm depth
22.04.10	Spring barley sown - mixture of varieties. Undersown red fescue - cv. Maximum
01.06.10	Herbicide - 1.25 l/ha Fighter 480 (bentazone)

Table A3.5 Management practice at **Faardrup** during the 2007 to 2010 growing seasons. The active ingredients of the various pesticides are indicated in parentheses.



Figure A4.1. Monthly precipitation at all localities for the monitoring period July 2000 – June 2010. Normal values (1961 – 1990) are included for comparison.

	Vertical screens				Suction cups			
	n.d.	det.<0.1	det.>=0.1	n.d.	det.<0.1	det.>=0.1		
		μg/l	μg/l		μg/l	μg/l		
AIBA	191			72				
2-hydroxy-desethyl-terbuthylazine	190	1		67	5			
2-hydroxy-terbuthylazin	191			71	1			
Azoxystrobin	163			67				
Bentazone	277			108	1			
Bromoxynil	192			72				
CGA 322704	175			64				
Clomazone	224			82				
Clopyralid [*]	6			63				
СуРМ	163			67				
Desethyl-terbuthylazine	191			70	2			
Desisopropyl-atrazine	190	1		55	17			
Dimethoate	176			65				
Epoxiconazole	199			74				
ĒTU	198	2		37	7			
Fenpropimorph	307			89				
Fenpropimorph-acid	276			73				
Flamprop-M (free acid)	176			65				
Flamprop-M-isopropyl	176			65				
Fluazifop-P (free acid)	178			65				
Fluroxypyr	194			70				
FMC65317	208			74				
PPU	507	2		70	119	3		
PPU-desamino	509			157	35			
Ioxynil	198			72				
Linuron	270			67				
Metribuzin	386	1		89	2			
Metribuzin-desamino	365			85				
Metribuzin-desamino-diketo	289	231	5	168	30	51		
Metribuzin-diketo	71	136	317	73	179	63		
Pendimethalin	430			144				
Pirimicarb	295			82				
Pirimicarb-desmethyl	295			81				
Pirimicarb-desmethyl-formamido	167			52				
Propiconazole	307			89				
Propyzamide	221			82				
RH24580	221			82				
RH24644	221			82				
RH24655	157			58				
Rimsulfuron	178			65				
Tebuconazole	195	1		77				
Terbuthylazine	179			72				
TFMP	3							
Thiamethoxam	175			64				
Triasulfuron	295			82				
Triazinamin	285			75				
Triazinamin-methyl	440			137				

Table A5.1 Number of samples where pesticides were either not detected (n.d.), detected in concentrations below 0.1 μ g/l (det<0.1 μ g/l) or detected in concentrations above 0.1 μ g/l (det>=0.1 μ g/l) at **Tylstrup**. Numbers are accumulated for the entire monitoring period, and pesticides monitored for less than one year are not included.

*Number of analysed samples collected from the monitoring wells was reduced (see Appendix 2 for explanation).

		Vertical sc	reens	Suction cups				
	n.d.	det.<0.1	det.>=0.1	n.d.	det.<0.1	det.>=0.1		
		μg/l	μg/l		μg/l	μg/l		
AIBA	178			45	2			
4-chlor-2-methylphenol	189			52				
Amidosulfuron	88			20	2	1		
AMPA	221	2		68	1			
Azoxystrobin	233			65				
Bentazone	377			68	34	5		
Bifenox	98	2		28	2			
Bifenox acid	48			11		1		
Bromoxynil	218			61				
CL153815 [*]	35			36				
Chlormequat [*]	14			28				
СуРМ	233			65				
Desethyl-terbuthylazine	472	24		128	18			
Desmethyl-amidosulfuron	88			23				
Dimethoate	169			48				
Epoxiconazole	323	1		90				
Fenpropimorph	246	1		76	1			
Fenpropimorph-acid	259			79				
Flamprop-M (free acid)	12			4				
Flamprop-M-isopropyl	12			4				
Florasulam	191			54				
Florasulam-desmethyl				28				
Fluazifop-P (free acid)	190			51				
Fluroxypyr	193			55				
Glyphosate	223			69				
PPU	402	282	2	37	99	55		
PPU-desamino	616	71		102	85	4		
Ioxynil	218			61				
MCPA	189			52				
Mesosulfuron [*]	12			45				
Mesosulfuron-methyl	285			78				
Metribuzin	26			6				
Metribuzin-desamino	26			4				
Metribuzin-desamino-diketo	6	7	13	6				
Metribuzin-diketo		7	19	3	3			
Nitrofen	100			30				
Pendimethalin	257			71				
РНСР	184			59				
Picolinafen [*]	35			35	1			
Pirimicarb	251			69				
Pirimicarb-desmethyl	251			68	1			
Pirimicarb-desmethyl-	_							
formamido	251			69				
Propiconazole	230			73				
Pyridate	116			39				
Rimsulfuron	168			48				
Tebuconazole	213	1		58				
Terbuthylazine	239			75				
TFMP	3							
Triazinamin-methyl	247			77				

Table A5.2 Number of samples where pesticides were either not detected (n.d), detected in concentrations below 0.1 μ g/l (det<0.1 μ g/l) or detected in concentrations above 0.1 μ g/l (det>=0.1 μ g/l) at **Jyndevad**. Numbers are accumulated for the entire monitoring period, and pesticides monitored for less than one year are not included.

*Number of analysed samples collected from the monitoring wells was reduced (see Appendix 2 for explanation).

	Drainage		Horizontal			Vertical screens			Suction cups			
	n.d.	det	det	n.d.	det	det	n.d.	det	det	n.d.	det	det
		$<0.1 \mu g/l$	>=0.1		< 0.1	>=0.1		< 0.1	>=0.1		< 0.1	>=0.1
			μg/l		$\mu g/l$	μg/l		μg/l	μg/l		µg/l	μg/l
AIBA	64			74			131					
2-hydroxy-desethyl-												
terbuthylazine	43	27	1	84			151	1				
2-hydroxy-terbuthylazin	45	26		84			152					
3-aminophenol	53			70			170			36		
4-chlor-2-methylphenol	51			66			124					
AMPA	25	107	15	123	5		226	10		8		
Azoxystrobin	55	10	-	91	0		168	17	2			
Bentazone	59	40	5	117	8	1	213	17	3			
Bifenox Difenov said	14	1	12	10	1	4	27	4	1			
Clopyrolid	1		15	62	1	4	119	1	/			
Chlormequat	20	1		36			66					
CvPM	20	59	5	120	8		223	19	1			
Desethyl-terbuthylazine	8	64	44	101	32		113	127	2			
Desisopropyl-atrazine	28	43		84	02		148	4	-			
Desmedipham	101	_		107	1		240			58		
Dimethoate	81		1	73	1		147			27		
EHPC	68			62			118			20		
Epoxiconazole	36			62			117					
Ethofumesate	119	14	1	162	2		339	3		54	3	2
Fenpropimorph	82			74			148			27		
Fenpropimorph-acid	81	1		74			147			27		
Flamprop-M (free acid)	/3	/ 11	1	74	1		148			26		
Fluggifon D (free acid)	106	11	1	122	1		148			21 56		
Fluazilop-P (lifee acid)	100			133	1		203			50		
Glyphosate	79	52	15	128			232	4		8		
IN-D8526	33	52	15	56			102	-		0		
IN-E7710	28	5		56			102					
IN-M7222	33			55	1		102					
Iodosulfuron-methyl-	52			78			149					
MCPA	51			66			123					
Metamitron	103	27	4	154	10		323	17	2	40	9	9
Metamitron-desamino	88	43	3	158	3	3	318	23	1	40	15	4
Metsulfuron methyl	52			78			149					
MHPC	100	1		106			234			55		
Pendimethalin	00	1/		121			222					
PHCP	62	14	4	66	2		109	8	4			
Phenmedipham	101			108			240	0		59		
Pirimicarb	160	14		209			433	3		59		
Pirimicarb-desmethyl	173	1		209			436			59		
Pirimicarb-desmethyl-										• •		
formamido	141			159			308			20		
Propiconazole	76	6		74			148	-		27		
Propyzamide	43	17	6	75	2	1	143	5	1			
Prosulfocarb	69	4	1	/8	1		14/					
RH24580 DH24644	04 51	15		78	1		149	1				
RH24655	51	15		79	1		148	1				
Terbuthylazine	31	51	Q	107	5		149	30	1			
TFMP	11	4	17	42	10		57	29	9			
Triazinamin	41		17	72	10		135	_/				
Triazinamin-methyl	82			74			148			27		
Triflusulfuron	33			56			102					

Table A5.3 Number of samples where pesticides were either not detected (n.d), detected in concentrations below 0.1 μ g/l (det<0.1 μ g/l) or detected in concentrations above 0.1 μ g/l (det>=0.1 μ g/l) at **Silstrup**. Numbers are accumulated for the entire monitoring period, and pesticides monitored for less than one year are not included.

	Drainage			Horizontal			Vertical screens			Suction cups		
	C		screens						-			
	n.d.	det	det	n.d.	det	det	n.d.	det	det	n.d.	det	det
		< 0.1	>=0.1		< 0.1	>=0.1		< 0.1	>=0.1		< 0.1	>=0.1
		µg/l	μg/l		μg/l	μg/l		µg/l	μg/l		µg/l	μg/l
AIBA	235	1		79	1		271			5		
2-hydroxy-desethyl-												
terbuthylazine	44	61	25	50			180					
2-hydroxy-terbuthylazine	43	70	17	50			180					
4-chlor-2-methylphenol	101	1		34			112					
Amidosulfuron	98			34			109					
AMPA	44	258	98	158	1		546	7		23		
Azoxystrobin	118	85	15	96			323	1				
Bentazone	169	133	12	107	15		408	1		3	2	2
Bifenox	23	2	1	19			50					
Bifenox acid	12	3	8	18			51					
Bromoxynil	135	1	2	41			125			3		
CL153815	49	20	11	40			118					
Clopyralid	1											
Chlormequat	44	1		18			56					
СуРМ	34	100	84	92	4		319	5				
Desethyl-terbuthylazine	18	108	37	59	7		232					
Desisopropyl-atrazine	89	70	1	62	1		197	26				
Dimethoate	88			42			159			23		
Epoxiconazole	35	11	2	19			69					
Ethofumesate	91	27	8	46			158					
Fenpropimorph	82	1		39			150			23		
Fenpropimorph-acid	82			34			124			17		
Flamprop-M (free acid)	118	13		55			208			23		
Flamprop-M-isopropyl	111	20		55			208			23		
Florasulam	91			35			125					
Florasulam-desmethyl	80			30			100					
Fluroxypyr	87	1	2	34			120	1				
Glyphosate	149	162	89	155	2		514	34	3	23		
Ioxynil	118	14	6	41			125			3		
MCPA	91	9	2	34			111	1				
Mesosulfuron	73			24			83					
Mesosulfuron-methyl	61	13		27			99					
Metamitron	81	27	15	46			158					
Metamitron-desamino	76	38	11	46			157					
Metsulfuron methyl	130			55			208			22	1	
Nitrofen	26			19			50					
Picolinafen	63	17		40			118					
Pirimicarb	159	39		67			225	1		6		
Pirimicarb-desmethvl	191			66			223			6		
Pirimicarb-desmethyl-												
formamido	198	13	13	76			261			5		
Propiconazole	192	22	3	86			309	2		23		
Tebuconazole	39	24	17	39			118	3	2			
Terbuthylazine	49	76	35	63			222	1				
Triazinamin	125			52			195	1		22		
Triazinamin-methyl	1											

Table A5.4 Number of samples where pesticides were either not detected (n.d), detected in concentrations below 0.1 μ g/l (det<0.1 μ g/l) or detected in concentrations above 0.1 μ g/l (det>=0.1 μ g/l) at **Estrup**. Numbers are accumulated for the entire monitoring period, and pesticides monitored for less than one year are not included.

	Drainage			Horizontal			Vertical screens			Suction cups		
	U U			screens						•		
	n.d.	det	det	n.d.	det	det	n.d.	det	det	n.d.	det	det
		< 0.1	>=0.1		< 0.1	>=0.1		< 0.1	>=0.1		< 0.1	>=0.1
		μg/l	μg/l		μg/l	μg/l		μg/l	μg/l		μg/l	μg/l
AIBA	68	1		61			132					
2-hydroxy-desethyl-	61	7	1	60	1		126	6				
2-hydroxy-terbuthylazine	90	20	1	85	4		164	30				
4-chlor-2-methylphenol	143		1	109			254					
AMPA	131	9	1	110			282	2		57	5	
Azoxystrobin	107			92			194					
Bentazone	60	12	6	62	3	1	136	4	3			
Bromoxynil	101			81			225			73		
CGA 322704	68			58			126					
Clomazone	84		1	69			166					
CvPM	103	4		92			194					
Desethyl-terbuthylazine	22	82	7	68	21		149	14	31			
Desisopropyl-atrazine	86	24	1	57	32		166	28				
Desmedinham	99	- ·	-	66	0-		165	-0		29		
Dimethoate	77			58			148					
EHPC	83			52			123			16		
Epoxiconazole	81			66			143			10		
Ethofumesate	114	7	6	90			188	24	7	27	2	
Fenpronimorph	70	,	0	58	1		156	27	/	54	2	
Fenpropimorph-acid	70			59	1		156			54		
Flampron M (free acid)	76	1		58			1/18			54		
Flamprop M isopropyl	70	1		56			140					
Fluarifon P (free acid)	01	1	4	66			142	5	1	26	3	
Fluarifon P butyl	00	4	+	66			165	5	1	20	5	
Flurovypyr	155		1	128	1		305			55		
FMC65217	155		1	120	1		303			55		
Clyphosata	137	4	1	100	1		282	2		61	1	
IN D8526	137	4		109	1		202	2		01	1	
IN-D0320	27			24			54					
IN-E7710 IN M7222	27			24			54					
IN-MI/222	27	1		24 91			24	1		72		
MCDA	142	1	1	100			224	1		15		
Metamitron	142	1	1	109			105	10	5	20		
Metamitron decomine	113	9	5	90			193	19	10	29		
	07	11	J 1	90			1/1	30	12	29		
MITIC Dandimathalin	91	1	1	55			105	1		29		
Pendimethalin	22	2		33			123	n		20		
Phenmedipham Divincia a sh	99	7		00			103	2		29 50		
Pirimicard	115			90			243	2		52 20		
Pirimicarb-desmethyl	94	0		00			102	3		29		
Pirimicarb-desmethyl-	97	3		00			103	2		29		
Propiconazole	14/	2	2	110	1		303	1		54		
Propyzamide	70	2	2	68	1		100					
Prosulfocarb	/9			61			126					
RH24580	74	4		69			155					
RH24644	/0	4		69			155					
КН24633	73	1		69			155	4				
I ebuconazole	50	4		53	_		120		~ ~			
I erbuthylazine	70	30	11	83	5	1	149	24	21			
TFMP				1			2					
Thiamethoxam	68			58			126					
Triazinamin-methyl	77			57			147					
Triflusulfuron	27			24			54			1		

Table A5.5 Number of samples where pesticides were either not detected (n.d), detected in concentrations below 0.1 μ g/l (det<0.1 μ g/l) or detected in concentrations above 0.1 μ g/l (det>=0.1 μ g/l) at **Faardrup**. Numbers are accumulated for the entire monitoring period, and pesticides monitored for less than one year are not included.



Figure A6.1 Quality control data for pesticide analysis by laboratory 1. <u>Internal laboratory control</u> samples are indicated by square symbols and the nominal level is indicated by the solid grey line (\blacksquare IQ measured, — IQ nominal concentration). <u>External control</u> samples are indicated by circles. Open circles indicate the nominal level (\bigcirc EQ nominal low, \bigcirc EQ nominal high), and closed circles the observed concentration (\bigcirc EQ measured low, \bigcirc EQ measured high).



Figure A6.1 continued. Quality control data for pesticide analysis by laboratory 1. <u>Internal laboratory control</u> samples are indicated by square symbols and the nominal level is indicated by the solid grey line (\blacksquare IQ measured, — IQ nominal concentration). <u>External control</u> samples are indicated by circles. Open circles indicate the nominal level (\bigcirc EQ nominal low, \bigcirc EQ nominal high), and closed circles the observed concentration (\bigcirc EQ measured low, \bigcirc EQ measured high).